CASE REPORT

Monosensitivity to Pangasius and Tilapia Caused by Allergens Other Than Parvalbumin

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Abstract

Fish allergy is one of the most common food allergies in populations where fish is a major part of the diet. Most fish-allergic patients react to the panallergen parvalbumin present in multiple fish species. Our aim was to investigate the clinical case of a patient with oral allergy syndrome to pangasius and Nile tilapia but tolerance of other fish and seafood. The temporal relationship between fish consumption and allergic symptoms, the positive skin prick tests, and the basophil activation test results for both fish species strongly supported the diagnosis of an immunoglobulin (Ig) E-mediated allergy. This was confirmed by the detection of specific IgE to 18-kDa and 45-kDa proteins in immunoblot analysis. Notably, the patient was not sensitized to parvalbumin, as shown by enzyme-linked immunosorbent assay using purified allergens.

Cross-reactivity between fish species can result from sensitization to allergens other than parvalbumin. This case report emphasizes the applications of flow cytometry–assisted analysis in the diagnosis of food allergy.

Keywords: BAT assay. Fish allergy. Monosensitivity. Pangasius. Tilapia.

Resumen

La alergia al pescado es una de las alergias alimentarias más habituales en poblaciones donde el pescado forma parte importante de la alimentación. La mayoría de los pacientes alérgicos al pescado reaccionan al panalérgeno parvalbúmina presente en muchas especies de peces. El objetivo del presente estudio fue investigar el caso clínico de un paciente con síndrome de alergia oral al pangasio y a la tilapia del Nilo, pero que presentaba tolerancia a otras clases de pescado y marisco. La relación temporal entre el consumo de pescado y los síntomas alérgicos, las pruebas de punción positivas y los resultados del test de activación de basófilos para ambas especies de pescado corroboraron firmemente el diagnóstico de una alergia mediada por la inmunoglobulina E (IgE). Esto se confirmó tras la detección de IgE específica a proteínas de 18 kDa y 45 kDa en los análisis de inmunotransferencia. En particular, el paciente no mostró sensibilización a la parvalbúmina, según se demostró por medio de un enzimoinmunoanálisis de adsorción (ELISA) con alérgenos purificados.

La reactividad cruzada entre especies de peces puede deberse a la sensibilización a alérgenos distintos de la parvalbúmina. Este caso clínico pone el énfasis en las aplicaciones del análisis por citometría de flujo en el diagnóstico de la alergia alimentaria.


Introduction

The majority of patients with fish allergy develop clinical symptoms to a range of fish species. The reason for this broad immunoglobulin (Ig) E cross-reactivity is sensitization to the fish panallergen parvalbumin [1,2]. Parvalbumins are low-molecular-weight, calcium-binding muscle proteins that are highly conserved across fish species. In immunological and molecular studies, parvalbumins have been characterized from a range of common fishes such as cod, salmon, and carp [3-5]. Fish allergens other than parvalbumin have also been reported. Examples are aldehyde phosphate dehydrogenase (~41 kDa) from codfish and fish collagen (~110 kDa) and transferrin-like protein (~94 kDa) from tuna [6-8]. Putative fish allergens such as IgE-reactive proteins of different molecular weights have also been described in swordfish, sole, eel, and snapper [9-12].
In recent years, different fish species from African and Asian countries have gained in importance on the European market. Pangasius (Pangasius hypophthalmus) and Nile tilapia (Oreochromis niloticus) account for a considerable market share as they are attractive alternatives to traditional and pricey fish species [13]. To date, no information has been published on species-specific fish allergy to pangasius and Nile tilapia.

In the present study, we analyzed the IgE-binding profile of a fish-allergic patient exclusively sensitized to pangasius and Nile tilapia in order to detect the causative allergen(s).

Case Description

We report on a patient with oral allergy syndrome characterized by itching and swelling of the lips, tongue, and oropharynx to both pangasius and Nile tilapia. The patient, a 27-year-old atopic woman with a history of house dust mite (HDM) allergy and documented allergy to sesame, had experienced several episodes of itching and swelling of the oropharynx within minutes of eating both fish species. She tolerated other seafood including cod, salmon, tuna, crustaceans, and molluscs.

Total IgE was 69 kU/L. Specific IgE was 1.87 kUa/L for HDMs, 0.43 kUa/L for sesame, 0.41 kUa/L for tilapia, and 0.46 kUa/L for cod. Specific IgE was negative for lobster, crab, oyster, mussel, and recombinant parvalbumin from carp (rCyp c 1) (<0.35 kUa/L, ImmunoCAP FEIA, Phadia, Uppsala, Sweden).

Skin prick tests (SPTs) were performed with fish extracts from pangasius, tilapia, and cod, which were also used for stimulation in the basophil activation tests (BATs). The skin reaction was positive with a clear wheal and flare of 14/40 mm for pangasius and 3/7 mm for tilapia. The skin test for cod was negative.

Basophil Activation Test

Flow cytometric analysis (FACScan, BD Immunocytometry Systems, Erembodegem, Belgium) of activated basophils was performed using Alexa 448-coupled anti-IgE (Sigma-Aldrich, Chemic GmbH, Steinheim, Germany) and phycoerythrin-conjugated anti-CD63 (Pharmingen, BD Biosciences, Erembodegem, Belgium) double labeling. Basophil activation tests involved a negative control (spontaneous CD63 expression without any allergen), a positive control (anti-IgE), and dialyzed extracts from pangasius, tilapia, and cod (serial dilutions of 0.01, 0.1, 1 and 10 µg/mL). For this purpose the fish extracts were prepared as described by Alenius et al [14].

The BAT results are summarized in Figure 1.
In the patient, pangasius and tilapia, but not cod, induced a consistent basophilic upregulation of CD63 above spontaneous expression. Fish-induced expression of CD63 from 2 healthy control individuals, however, remained entirely negative. In 3 patients with documented cod allergy, the BAT was positive for all 3 fish species in 2 of the patients and for cod and tilapia in the other.

**Protein Extraction and Purification**

Fish muscle extracts were prepared from pangasius (*Pangasius hypophthalmus*), Nile tilapia (*Oreochromis niloticus*), salmon (*Salmo salar*), cod (*Gadus morhua*), tuna (*Thunnus albacares*), mackerel (*Scomber scombrus*), and carp (*Cyprinus carpio*) as described elsewhere [15]. Native parvalbumins from pangasius, salmon, cod, and mackerel were purified by ion exchange and gel filtration chromatography. Recombinant β-1 parvalbumins from salmon and cod were obtained by expressing the complementary DNAs (Swiss-Prot Q91482, Q90YL0) in *Escherichia coli* M15 [3,4]. These parvalbumins were purified by affinity and gel filtration chromatography. Protein identity was confirmed by immunodetection using a commercial mouse monoclonal anti-β-parvalbumin antibody (Swant, Bellinzona, Switzerland).

**IgE Quantification by Enzyme-Linked Immunosorbent Assay**

Ninety-six-well plates (Maxisorb, Nunc, Wiesbaden, Germany) were coated overnight at 4°C with 5 µg/mL of purified native and recombinant parvalbumins in phosphate buffered saline, pH 7.2. IgE were quantified versus a characterized patient serum with a known titer to cat serum (1:3, 1:5, and 1:10). An anti-parvalbumin antibody (Swant) and serum from a fish-allergic individual (5 kUa/L for cod, ImmunoCAP) were used as positive controls.

The enzyme-linked immunosorbent assay (ELISA) results for IgE reactivity to purified fish parvalbumins are shown in the Table. The patient’s IgE antibodies did not bind to native isofrom mixtures or to recombinant single isoforms of parvalbumins from 4 common fish species.

**IgE Immunoblot and Immunoblot Inhibition**

Fish protein extract was analyzed by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto polyvinylidene difluoride membranes (Millipore, Bedford, Massachusetts, USA) [15,16]. The membranes were incubated overnight with diluted patient serum (1:3) and the anti-parvalbumin antibody (Swant). In Ca²⁺-depletion experiments, 5 mM of ethylene glycol tetraacetic acid (EGTA) was added to the primary antibodies during incubation with blotted extract. To analyze cross-reactivity, the patient’s serum was incubated with pangasius, tilapia, or cod protein extract at a final concentration of 1 mg/mL for 2 hours, prior to incubation with blotted pangasius or tilapia extract.

In the immunoblot analysis of total fish extracts, the patient’s IgE detected strong protein bands of about 18 kDa and 45 kDa in pangasius and tilapia and weak bands in cod (Figure 2). No signals were obtained for protein extracts from salmon or carp (data not shown). Notably, no IgE reactivity was detected with extracts from mackerel or tuna, both close relatives of tilapia and belonging to the same order of perch-like fish (Perciformes). The patient’s IgE did not recognize the parvalbumin-like bands detected by the monoclonal antibody in any of the fish extracts. IgE detection of the putative 18-kDa- and 45-kDa allergens was

<table>
<thead>
<tr>
<th>Coating Parvalbumin</th>
<th>Ua/L (Patient)</th>
<th>kUa/L (Positive Control)</th>
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<tbody>
<tr>
<td>Native Pangasius</td>
<td>&lt;0.1</td>
<td>0.5</td>
</tr>
<tr>
<td>Cod</td>
<td>&lt;0.1</td>
<td>4.5</td>
</tr>
<tr>
<td>Salmon</td>
<td>&lt;0.1</td>
<td>3.2</td>
</tr>
<tr>
<td>Mackerel</td>
<td>&lt;0.1</td>
<td>0.3</td>
</tr>
<tr>
<td>Recombinant Cod, β-1</td>
<td>&lt;0.1</td>
<td>5.2</td>
</tr>
<tr>
<td>Salmon, β-1</td>
<td>&lt;0.1</td>
<td>3.9</td>
</tr>
</tbody>
</table>

Table: Immunoglobulin (Ig) E antibody binding to purified fish parvalbumins in the patient was quantified by enzyme-linked immunosorbent assay. Specific IgE from a fish-allergic individual were titrated as a positive control.

Figure 2. Total extracts from pangasius, tilapia, and cod were analyzed by immunoblotting. 1, anti-β-parvalbumin antibody detected parvalbumins in the fish extracts. 2, the patient’s immunoglobulin (Ig) E detected putative new allergens of 18 kDa and 45 kDa (>, but not parvalbumins. T, The patient’s IgE reactivity to the 18-kDa and 45-kDa pangasius proteins was inhibited by tilapia extract. P, the patient’s IgE reactivity to the 18-kDa and 45-kDa tilapia proteins was inhibited by the pangasius extract. C, serum incubation with cod extract did not affect IgE binding to either of the putative allergens from pangasius and tilapia.
not influenced by incubation with 5 mM EGTA (data not shown). Cross-reactivity between pangasius and tilapia was analyzed by immunoblot inhibition (Figure 2). IgE binding to putative 18-kDa allergens in pangasius and tilapia extracts was abolished upon serum cross-inhibition with the extracts and IgE detection of putative 45-kDa allergens in both extracts was markedly reduced. Serum incubated with cod extract did not affect IgE reactivity to either of the putative allergens in pangasius or tilapia.

**Discussion**

Parvalbumins are recognized as major fish allergens that cause broad IgE cross-reactivity, even in tropical fish species [1,2,17]. Recently, parvalbumin was cloned from Mozambique tilapia (*Oreochromis mossambicus*) [18], but homologue muscle proteins have not yet been characterized in pangasius or Nile tilapia.

We studied putative fish allergens in a patient with clinical symptoms exclusively due to pangasius and Nile tilapia. SPTs were positive only for pangasius and tilapia extracts, confirming the clinical history. Notably, immunoblot and ELISA analysis showed that fish parvalbumins were not the causative allergens.

Pangasius and tilapia are distantly related in taxonomy, but tilapia is a member of the same order as mackerel and tuna (*Perciformes*). No IgE reactivity was found to protein extracts from mackerel or tuna in our patient, indicating that cross-reactivity between fish species cannot necessarily be assumed on the basis of taxonomic classification.

Sensitization to single fish species caused by allergens other than parvalbumin has already been reported [8-10]. In our patient, the presence of 2 cross-reactive, homologous allergens of 18 kDa and 45 kDa in pangasius and tilapia can be assumed from cross-inhibition assays. The identity of both proteins remains to be determined. The higher molecular weight protein might even represent a dimer of the 18-kDa allergens. Cross-reactivity under denaturing conditions, a hypothesis supported by the negative BAT results and cross-inhibition assays with cod extract. In the present case, native conditions were more representative of clinical relevance than testing of IgE binding to denatured proteins.

Diagnosis of IgE-mediated fish allergy is based on a precise clinical history and corroborated by different in vitro and in vivo tests. ImmunoCAP analysis is helpful to establish cross-reactivity, but is less sensitive and not always available for single species. In our patient, in vitro diagnosis of pangasius and tilapia allergy was documented by flow-assisted analysis of in vitro activated basophils (BAT) [19]. Simulating native conditions, this technique correlated well with the patient’s history of fish allergy and allowed discrimination between clinically relevant and irrelevant specific IgE.

In summary, we have reported the clinical case of a fish-allergic patient sensitized to pangasius and tilapia, but not to other types of fish. Cross-reactivity between pangasius and tilapia was not caused by the panallergen parvalbumin, but by putative 18-kDa and 45-kDa fish allergens. Both fish allergens might be representatives of a new fish allergen family with homologous proteins in other species, which gives impact to the present report. For developing strategies in component-resolved diagnosis, the exact nature of these new fish allergens needs to be investigated.

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**References**


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