

Hypersensitivity Reactions to Nonsteroidal Anti-inflammatory Drugs in Children and Adolescents: Selective Reactions

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■ Abstract

Nonsteroidal anti-inflammatory drugs (NSAIDs) are used throughout the world to treat pain and inflammation; however, they can trigger several types of drug hypersensitivity reactions (DHRs) in all age groups. Although most such reactions occur through activation of the leukotriene pathway without specific immunological recognition (cross-intolerance), a significant number of DHRs to NSAIDs are due to immunological mechanisms (selective reactions [SRs]). SRs are thought to be induced by specific IgE antibodies or by T cells. In this manuscript, we focus on SRs, which are of great concern in children and adolescents and comprise a heterogeneous set of clinical pictures ranging from mild entities such as urticaria/angioedema to potentially life-threatening conditions such as Stevens-Johnson syndrome/toxic epidermal necrolysis.

Paracetamol and ibuprofen are the most frequent elicitors of IgE-mediated SRs, although pyrazolones have also been implicated. T cell-mediated reactions are infrequent in children but have been associated with ibuprofen, naproxen, and dipyrone.

In this review, we analyze the available literature on SRs in children and adolescents, with emphasis on epidemiological data, mechanisms, and drugs involved, as well as on diagnostic procedures.

Key words: Drug hypersensitivity reactions. NSAIDs. Single NSAID-induced urticaria/angioedema or anaphylaxis. Single NSAID-induced delayed reactions.

■ Resumen

A pesar de su eficacia en el tratamiento del dolor y la inflamación los antiinflamatorios no esteroideos (AINE), los medicamentos de mayor consumo mundial, también son la causa más frecuente de reacciones de hipersensibilidad a fármacos (RHF) en cualquier tramo de edad. Aunque en muchas de estas reacciones se liberan mediadores inflamatorios en ausencia de reconocimiento inmunológico específico (intolerancia cruzada), un porcentaje considerable de las RHF a AINE se producen a través de mecanismos inmunológicos (reacciones selectivas, SRs). En éstas participarían anticuerpos IgE específicos o células T. Las SRs son de gran interés en niños y adolescentes e incluyen un conjunto heterogéneo de entidades que comprenden desde manifestaciones clínicas de poca gravedad como la urticaria y el angioedema hasta otras como el síndrome de Stevens-Johnson y la necrosis epidérmica tóxica, que pueden suponer una amenaza para la vida. En niños el paracetamol y el ibuprofeno son los medicamentos más frecuentemente implicados en las SRs mediadas por IgE aunque también se ha descrito la participación de las pirazonas. Las reacciones mediadas por linfocitos T son menos frecuentes pero también se han descrito en relación con la administración de ibuprofeno, naproxeno y dipirona. En esta revisión analizaremos la literatura actual sobre las SRs en niños y adolescentes, centrándonos en los datos epidemiológicos, mecanismos y fármacos implicados, así como las pruebas disponibles para su diagnóstico.

Palabras clave: Reacciones de hipersensibilidad a fármacos. AINE. Reacciones selectivas. Urticaria/angioedema o anafilaxia inducidas por un único AINE. Reacciones tardías inducidas por un único AINE.

Introduction

Nonsteroidal anti-inflammatory drugs (NSAIDs) are widely used to control pain and fever and to treat various inflammatory diseases [1]. By inhibiting the synthesis of prostaglandins, they can induce both beneficial effects and adverse reactions [1], including type B reactions, which are not dose-dependent [2,3]. Type B reactions encompass drug hypersensitivity reactions (DHRs) to NSAIDs, which affect all age groups, including children and adolescents [4]. Paracetamol and ibuprofen are widely used in children [5-7], who can take all NSAIDs, including potent COX-1 inhibitors, as they grow older and enter adolescence.

The 2 most frequent types of hypersensitivity reactions to NSAIDs are cross-intolerance, which is related to an imbalance in the arachidonic acid pathway, and selective reactions (SRs), which are thought to be due to specific immunological mechanisms [8-11]. A recent new classification from the European Academy of Allergy and Clinical Immunology (EAACI) grouped all the clinical entities induced by hypersensitivity to NSAIDs reported to date [12]. In this manuscript, we focus on SRs, which can be differentiated into 2 groups:

1. Single NSAID-induced urticaria/angioedema or anaphylaxis (SNIUAA). This term applies when patients present urticaria and/or angioedema following the intake of a single NSAID or closely related drugs from the same group (eg, pyrazolones and diclofenac) but tolerate alternative, non-chemically related NSAIDs [8,9]. The reaction is independent of the strength of COX-1 inhibition, pointing to a specific immunological mechanism. These reactions usually occur within the first hour following drug intake, although this interval can be longer [13].
2. Single NSAID-induced delayed reactions (SNIDRs). SNIDRs comprise a highly heterogeneous group of clinical entities ranging from mild reactions, such as maculopapular exanthema (MPE), nonimmediate urticaria, and fixed drug eruption (FDE), to the potentially life-threatening Stevens-Johnson syndrome and toxic epidermal necrolysis (SJS/TEN). Reactions in this category differ from SNIUAA in terms of timing (>1 hour after intake) and clinical manifestations. The category also includes organ-specific reactions such as allergic hepatitis induced by drugs (eg, diclofenac) [14]. SNIDRs are sometimes mistaken for viral diseases [15].

The reactions included in both of these categories also appear in children and adolescents, although differences in severity, frequency, and drug involved have been detected in comparisons with adults [16,17].

In this review, we analyze current literature to discuss epidemiological data, as well as mechanisms and drugs involved, diagnostic procedures, and recommendations.

Epidemiology

Most articles dealing with adverse reactions to NSAIDs do not analyze in detail the underlying mechanisms or the various entities induced by these drugs. Up to 30% of DHRs caused

by NSAIDs belong to the SNIUAA category [10], although figures vary depending on the study and the population evaluated [18,19]. No large population studies have been published for patients aged below 14 years, and the only available findings are from isolated case reports and small series [20]. Information is also scarce for SNIDRs, likely owing to their low prevalence. MPE is the most common manifestation in children and adolescents [21-23]. However, despite its temporal association with NSAIDs, it is often caused by other agents, such as viruses [15,24].

In our view, it is also relevant to include organ-specific reactions within SNIDRs, particularly those affecting the liver [25]. These reactions may be isolated or appear in the context of drug reactions with eosinophilia and systemic symptoms (DRESS) [26]; however, no appropriate epidemiological data are available on DRESS in children. This subgroup also includes drug-induced liver injury [27], pneumonitis [28], and aseptic meningitis [29], although to our knowledge, data on the prevalence of these entities in children are lacking.

Drugs Involved

The most commonly involved drugs in IgE-mediated reactions in adults are pyrazolones, followed by ibuprofen and diclofenac, although virtually all NSAIDs have been implicated. In general, many of the principles applicable for immediate hypersensitivity reactions to β -lactams can be also applied in IgE-dependent reactions [30]. Paracetamol and ibuprofen are the most frequent elicitors of SNIUAA in children [31-34], although pyrazolones have also been implicated [20].

Despite the low frequency of SNIDRs in children, FDE has been reported with ibuprofen, naproxen, dipyron, oxicams, nimesulide, and other NSAIDs [31,35-37]. A strong association has also been reported in children between FDE affecting the labial mucosa and therapy with naproxen and oxicams [35]. Bufexamac is a potential trigger of acute generalized exanthematous pustulosis (AGEP) [38]. Aspirin (ASA) has been shown to induce TEN [39], and other NSAIDs have been associated with an increased risk of SJS/TEN [40].

Photoallergic and phototoxic reactions can also be induced by NSAIDs [41], and, while infrequent, should be considered when treating children who have taken NSAIDs orally (eg, propionic acid derivatives and oxicams) or topically (eg, propionic acid derivatives, arylacetic acid derivatives, and other drugs such as bufexamac) [41-44].

The chemical structure of some COX-2 inhibitors is similar to that of sulfonamide derivatives, and it may be the case that children sensitized to these drugs will also respond to COX-2 inhibitors [37].

Also relevant is the observation that some cutaneous symptoms induced by DHRs, including those caused by NSAIDs, are not always clinically distinguishable from those elicited by viruses. This is particularly important in children, as the reaction may be due to an interaction between an NSAID and a viral infection [45-47]. Arylacetic acid derivatives (eg, diclofenac) [48], ibuprofen, and other propionic acid derivatives have been implicated in organ-specific reactions in children and adolescents [49].

Pathophysiology

In order to fully understand the specific immunologic mechanisms involved in SRs to NSAIDs (SNIUAA and SNIDRs), we must consider interactions between drugs and proteins. Drugs must bind to proteins to become immunogenic and induce an immune response [14]. The nature of this binding, whether covalent or not [50], is beyond the scope of this review.

NSAIDs bind to serum and cellular proteins [51,52], which often require previous generation of active metabolites [50]. For example, diclofenac, which is involved in DHRs in children, requires aromatic hydroxylation and acyl glucuronidation to bind proteins covalently [53]. The protein-drug adducts generated may be involved in anaphylactic, exanthematic, or severe skin reactions such as DRESS, as well as in organ-specific reactions such as hepatitis [50]. Although mainly reported in adults, these entities can occur in children and adolescents [26,27,38].

Propionic acid derivatives are probably the drugs most commonly involved in DHRs to NSAIDs and have been shown to produce protein adducts with cell and serum proteins. Although the structural details of these conjugates have not been sufficiently well described, it is thought that this process is required for the induction of an IgE-mediated or T-cell response [54].

SNIUAA

Evidence of specific IgE antibodies has been found for several NSAIDs. In one study, 27 of 28 ASA-sensitive patients had IgE to salicyloyl and O-methylsalicyloyl determinants [55]. Specific antibodies to ASA have also been reported [56-58].

Pyrazolones are used for analgesic purposes in adults and children in many countries, although they are not licensed in others. Where they are widely used, they are the most common culprit of SNIUAA, and the existence of specific IgE has been reported in some studies [59], but not in others [60]. Dipyron, commonly known as metamizole, also induces reactions through the formation of specific IgE [61,62]. Propyphenazone has been demonstrated to induce specific IgE [63]. Interestingly, selectivity for this drug was independent of the recognition of dipyron [63].

Diclofenac and other arylacetic acid derivatives can induce immediate selective reactions, particularly anaphylaxis [64-66]. However, a recent study was unable to find specific IgE to diclofenac conjugates in the sera of allergic individuals [67].

SNIDRs

NSAIDs involved in T cell-mediated reactions in children and adolescents include propionic acid derivatives, diclofenac, pyrazolones, ASA, and the recently introduced selective COX-2 inhibitors. Although less common in children, reactions have been reported, including FDE triggered by ibuprofen [31,68], paracetamol [68], and nimesulide [37]. DRESS induced by ASA [69] and AGEP induced by bufexamac [38] and paracetamol are associated with an increased risk of SJS/TEN [40]. Although SJS/TEN has frequently been reported to be induced by ibuprofen [70-72], it is difficult to identify

NSAIDs as the culprit, since other drugs are often administered simultaneously [72].

In addition to these 5 major groups, other less common entities have been reported and may possibly be due to additional, as yet undefined mechanisms. For example, several NSAIDs, such as paracetamol, indomethacin, and some pyrazolone derivatives, have been implicated in serum sickness-like syndrome [73]. However, the definition of this entity is usually based on clinical symptoms rather than the underlying mechanisms, which remain unknown. For β -lactams, recent evidence indicates that these responses are T cell-dependent [74]. Other reactions include organ-specific diseases (eg, hepatitis, pneumonitis, and aseptic meningitis) that can be attributed to T cells or other pathogenic mechanisms. Additional details can be seen elsewhere [25,75,76].

Characteristics of Clinical Entities

SNIUAA

Immediate selective responses that are presumably mediated by specific IgE [9] and sometimes confirmed as such [55,56] can appear at all ages [20]. In children, exposure to NSAIDs increases; this is a prerequisite for developing immediate responses [9]. As occurs with β -lactams [77] and possibly with other drugs, anaphylaxis in children is milder, and severe reactions are less frequent, although they do occur [20].

The approach for evaluating children with suspected DHRs to NSAIDs often involves the administration of a drug other than the culprit drug. Therefore, many cases where a selective immediate reaction occurs are not diagnosed. It is also important to consider the natural evolution of the disease and the fact that IgE antibody levels decrease over time [62,78], thus leading to loss of sensitivity for a given patient despite a clear clinical history of reactions [62,78].

SNIDRs

The most common clinical SNIDR is MPE, followed by FDE and other reactions, which can be severe and even life-threatening [8]. Although the clinical manifestations of these entities have been described elsewhere [8], here we provide information that could be of particular relevance in children.

MPE consists of diffuse cutaneous erythema and areas of skin elevation, sometimes accompanied by angioedema, which can evolve to vesicles. Immunohistochemistry reveals the presence of a mononuclear cell infiltrate surrounding blood vessels, with a predominance of CD4 lymphocytes expressing several activation markers [79].

Skin biopsies from patients with nonimmediate NSAID-induced urticaria show a typical T-cell infiltrate supporting a T-cell effector phenomenon. Urticaria often presents as typical wheal and flare lesions with angioedema. This type of reaction, which is usually described for amoxicillin [74], has also been reported for NSAIDs [80].

NSAIDs are among the most common causes of FDE, which consists of erythematous patches that recur mainly at the same site on re-exposure to the offending drug [81].

Intradermal CD8⁺ cells producing large amounts of IFN- γ seem to play a key role [82].

DRESS, which is also known as drug-induced hypersensitivity syndrome, is a serious reaction characterized by fever, cutaneous eruption, and the involvement of several internal organs that may lead to hepatitis, renal impairment, and hematologic abnormalities such as hypereosinophilia [83]. Triggering NSAIDs include ibuprofen and selective COX-2 inhibitors [84,85].

SJS/TEN is characterized by widespread destruction of the epithelium and mucous membranes [86] owing to massive apoptosis of keratinocytes with the formation of blisters containing natural killer and CD8⁺ cells [87]. TNF- α , granzyme B, perforin, and Fas ligand have been found in the supernatant [88-91]. Activation of keratinocytes by IFN- γ makes T cells more sensitive to T cell-mediated cytotoxicity [92]. Further information can be found in a recent review [93]. Although uncommon, respiratory and gastrointestinal involvement has been reported in children with SJS/TEN [94].

Topical NSAIDs such as ketoprofen have been associated with contact dermatitis in adults [44]; however, this drug appears to be safe in children [95].

Evolution and Risk Factors

In the case of SNIUAA, it is reasonable to suspect that patients lose sensitivity over time [62,78,96], as occurs with β -lactams and other drugs [96,97]. *In vitro* evidence indicates that patients with positive skin and basophil activation test results may have negative results in subsequent tests [62,96], although it is not clear whether this means they will be able to tolerate the drug. In the case of T-cell reactions, responses to drugs persist over long periods, and patients do not show a loss of sensitivity [98].

Although atopy was shown to be more frequent in both patients with aspirin-induced asthma and patients with selective reactions to pyrazolones [99], a recent study showed the atopic status of selective responders to be similar to that of a control group [10]. Increased use of drugs and a previous history of anaphylaxis are risk factors [100,101].

The predisposing factors for SNIDRs are not well known, although paracetamol may increase the risk of SJS/TEN [40]. Because T-cell reactions to drugs usually depend on the generation of active metabolites that can stimulate the immune system, certain metabolic patterns and specific HLA alleles may constitute risk factors. More details are given in a recent review [102].

Diagnostic Tests

The diagnostic tests used with SRs are based on immunological recognition of the drug *in vivo* or *in vitro*.

In Vivo Tests

Skin testing in SNIUAA consists of prick and intradermal tests using soluble drugs that can be applied on the back of younger children or on the forearm of adolescents [103,104]. Guidelines for skin testing have already been published [105] and are valid for all age groups, although compliance and

the rate of false positives must be taken into account in children [105]. Because SRs are particularly uncommon in children, no studies reporting sensitivity and specificity are available for patients aged less than 14 years [3]. For adolescents, some recommendations can be extrapolated from studies in adults. In the case of pyrazolones, sensitivity to dipyrone has been shown to reach 85.7% [62], although in another study it was only 42.3% [106]. For propylphenazone, serum specific IgE was found in 58% of patients [63].

In the case of SNIUAA, *in vitro* tests for determination of IgE levels have not been successful, although some studies have been published for a limited number of drugs (eg, pyrazolones) [63]. Skin testing for delayed reactions to NSAIDs is also rather limited [107], with little known about its success rate in children and adolescents [108]. Finally, although *in vitro* tests for T cell-dependent responses do exist, they are rather complex to interpret; and further evaluation of their utility is required [109].

A number of publications provide precise guidelines for diagnosis of SNIDRs, including patch and intradermal testing [103,104]. Some of these tests can be used with children, although false-positive results may be more frequent [107]. Patch testing with ibuprofen, naproxen, ketoprofen, and piktoprofen at 2% and 5% (wt/wt) has been performed in a patient with FDE induced by ibuprofen. Readings were taken at 24, 48, and 72 hours, and the only positive result was for ibuprofen at 5% after 72 hours [31].

With respect to the diagnosis of photoallergy, we recommend a recently published consensus document [110].

In Vitro Tests

Given that SNIUAA is presumed to be IgE-mediated, immunoassays based on experimental or commercially available prototypes may be applicable. IgE to various NSAIDs has been reported in isolated cases and small series for ASA [56] and propylphenazone [63]. Experimental prototypes have also been developed for diclofenac, which is commonly used to treat SNIUAA [111]. However, convincing data have yet to be published [67].

Although ibuprofen frequently elicits immediate selective reactions in children and adolescents, no direct evidence of specific IgE has been obtained to date [9].

The basophil activation test has proven successful for anaphylactic reactions and urticaria induced by pyrazolones. Sensitivity is around 50% [111], although some studies have found higher values [106]. Studies carried out for patients with selective immediate reactions to these drugs have shown that if patients are followed over time, positive test results can become negative [62], as occurs with β -lactams [111].

In the past, many studies were carried out to monitor acute phase responses in nonimmediate allergic reactions by taking peripheral blood and samples of the affected skin. In this type of reaction, several cell-populations home to the skin and generate the tissue infiltrate that is typical of such lesions [112]. This finding has been of value for differentiating between drug-induced and virus-induced diseases [45]. The procedures have not been sufficiently exploited to characterize the acute phase of DHRs in children. Monitoring of the acute phase can also be of value when attempting to distinguish between immediate

and nonimmediate reactions. Therefore, quantitation of tryptase in peripheral blood and N-methyl histamine in urine can prove useful when distinguishing between anaphylaxis and other types of nonallergic shock and for determining nonimmediate T cell–dependent responses [113].

Drug Provocation Test

In SNIUAA, drug provocation testing consists of the administration of incremental doses of the culprit drug at regular time intervals until an objective response that can be considered positive is obtained. In a recent study of children with a reaction suggestive of hypersensitivity to NSAIDs, oral challenge confirmed a selective mechanism in 14% of cases and cross-intolerance in 44% [114].

No consensus protocols exist for evaluating SNIUAA in children, although some studies propose algorithms that may be of value. Doña et al [10] gave patients alternative drugs. For example, in the case of anaphylaxis and/or urticaria induced by ibuprofen, the patients were given ASA; if this led to a reaction, they were classified as cross-intolerance [10]. If several episodes of anaphylaxis/urticaria occurred with the same drug (eg, ibuprofen or diclofenac) but the patient tolerated an alternative (eg, ASA), the case was classified as SNIUAA. However, if only 1 episode was reported and the alternative drug was well tolerated, controlled administration of the culprit drug was performed. Recent, related studies on children may be of interest to the reader, with additional information on dosages [20,114].

SNIDRs comprise the most heterogeneous group of entities induced by DHRs to NSAIDs. General rules are therefore not applicable, although it is now widely accepted that MPE, the most common clinical entity, may be challenged since it is a mild reaction [14,115]. Interestingly, many patients with MPE attributed to NSAIDs are later shown to have good tolerance, likely due to the wide range of other factors that can induce this entity [116]. The principle for challenge in this entity is to use increasing doses of the culprit drug until the recommended therapeutic dose is reached [117]. To reduce the risk of adverse effects, it is recommended to escalate the dose every 24–48 hours and check for symptoms indicative of a reaction [117]. In principle, challenge should be initiated in hospital, and patients should be monitored every time a dose is administered. This procedure can be carried out in an outpatient setting [117].

For FDE, providing that lesions are not too extensive, many centers also base diagnosis on challenge testing, particularly in cases where the patient is given several drugs. This procedure is more appropriate for identifying the culprit than for diagnosing the clinical entity [118,119].

Challenge testing is not recommended for the remaining clinical entities within this category, especially in the more severe reactions such as SJS/NET, AGEP, DRESS, and organ-specific reactions such as allergic hepatitis [120]. In these cases, the diagnosis should be based on the clinical history and histopathology data [14].

Differential Diagnosis

The differential diagnosis must include the time interval following contact with the trigger, which in many instances can

be a drug other than NSAIDs or an allergen, usually an orally ingested food allergen, insect sting, or any other exposure that can trigger anaphylactic or urticarial reactions.

In SNIUAA, the time interval between drug intake and the occurrence of the episode is very short (usually less than 1 hour). Therefore, it is important to consider whether other drugs and/or food allergens have been taken at the same time as the NSAID [77,121].

Diagnosis of SNIDR can be complicated by viruses and bacteria capable of inducing clinical conditions that are indistinguishable from a drug reaction. Special attention must be paid to severe T cell–mediated reactions. For example, staphylococcal scalded skin syndrome can have a very similar clinical presentation to TEN and is more frequent in pediatric populations. However, the syndrome usually has less mucosal surface and internal organ involvement than TEN [122,123]. It is also important to note that although SJS and TEN belong to the same spectrum of severe delayed reactions, prognosis for patients with SJS is better, with fewer sequelae and lower mortality [124]. Generalized bullous FDE is another classic differential diagnosis of TEN, and differentiation relies mainly on the presence of primary lesions and the absence of constitutional symptoms in FDE [124]. Although erythema multiforme in children is mostly associated with infections by herpesvirus, it can also be induced by drugs, and both typical and atypical targets often appear in association with blisters, thus making it difficult to distinguish from SJS [125]. In children, SJS can also be induced by *Mycoplasma pneumoniae*, although it is not associated with erythema multiforme [126]. Monitoring the acute phase can be of value when attempting to elucidate the mechanisms underlying the reaction [116,127].

Treatment

Administration of an alternative drug is recommended in the case of SNIUAA [9]. For example, a child who experiences anaphylaxis after taking paracetamol should be able to tolerate COX-1 inhibitors [34]. However, drugs from the same chemical group as the culprit may also induce reactions owing to cross-reactivity, as occurs with arylpropionic agents (eg, ibuprofen and ketoprofen), arylacetic derivatives (eg, diclofenac and aceclofenac), and pyrazolones (eg, propyphenazone and dipyron). Cross-reactivity between selective COX-2 inhibitors may also occur [9].

The concept of cross-reactivity in T cell–dependent reactions is more complex than for immediate IgE-dependent reactions, because reactivity in the former is often related to the drug-metabolite rather than to the parent drug [14]. The culprit drug can be readily identified based on patch and/or intradermal test results, which, if positive, indicate that the drug should be avoided [117,128]. However, given that these tests have low sensitivity, controlled administration remains necessary. Exceptions include cases such as multiple drug hypersensitivity syndrome, where patients may respond to several non-chemically related drugs [129]. However, this condition has seldom been reported in children.

Of particular interest is the treatment of severe reactions. In general, the sooner treatment and supportive measures are initiated, the better the chance of survival, although there is still

no consensus on the best treatment to use. Most expertise is from studies carried out in adults. In the case of TEN, patients must be managed in burn and/or intensive care units [130]. General measures include maintenance of fluids and electrolyte balance, regulation of temperature and evaporation, and potential administration of corticosteroids [130-132].

In a large study involving pediatric patients with SJS/TEN, the average hospital stay was more than 20 days [72]. Intravenous and oral corticosteroids were used at an equivalent dose of 1-2mg/kg/d of prednisolone. Intravenous immunoglobulin 1.1 g/kg has been used in patients with extensive skin lesions (around 70% of body area) [133]. In a series of 10 consecutive patients with TEN, specific CD95 blockade with intravenous immunoglobulin reversed disease progression in all cases [90]. However, as with adults, the lack of comparative studies prevents us from recommending the appropriate dose or from confirming the validity of this therapy. Furthermore, use in children remains controversial [134]. For TEN, avoidance of corticosteroids combined with debridement of necrotic skin and blisters and coverage of the affected area with human skin allograft could prove to be a successful and safe treatment strategy [135].

Complications of severe reactions in children include sepsis, respiratory distress, infections, and ocular injuries [130,135].

A general approach to severe reactions where the immune system plays a predominant role involves combinations of immunosuppressors [136]. Administration of corticosteroids is controversial. Pulse therapy with dexamethasone and other corticosteroids has been administered in widespread lesions [137], although several complications have been reported [138].

Following the publication by Viardet et al [90], who suggested that intravenous immunoglobulin could be of value for the treatment of SJS/TEN, many studies have shown varied responses [138-146]. One study suggested a dose of 3-4 g/kg [140]. A detailed analysis of various corticosteroid-based approaches (with or without intravenous immunoglobulin) has been published [147]. Other therapies have also been proposed for children [148,149]. Less severe SNIDRs, such as AGEP, DRESS, and FDE, are usually self-limiting conditions that resolve once the offending drug has been withdrawn. In the acute phase, supportive measures can consist of wet-to-dry dressings with drying and application of disinfectant solutions to avoid infection [150]. Some individuals may require topical corticosteroids for symptom relief or systemic treatment in more severe cases [150]. Systemic treatment is especially effective for AGEP [151].

Desensitization

Desensitization is seldom used in SRs in children and adolescents and is preferred for adults with cardiovascular diseases, stroke, and other diseases [152]. When indicated, the general rules for adults can be applied in children after adjusting for dose [153-155], although desensitization to paracetamol or ibuprofen is rarely needed in children. The dosage should be escalated slowly to prevent anaphylactic reactions.

Further Research

Although SNIUAA is largely thought to be IgE-mediated, the precise underlying mechanism needs to be clarified and other possibilities explored. IgE has only been detected for ASA [56] and propylphenazone [63]. Neither skin testing nor *in vitro* tests have been developed as a diagnostic tool for other NSAIDs. The major difficulty in developing these tests is identification of the specific triggering metabolite and its conjugation to a solid support.

Similar difficulties are seen in T-cell reactions, where the sensitivity of skin testing (intradermal or patch testing, or photopatch for photoallergic reactions) is generally low.

In the near future, the results of genetic and transcriptomic studies and advanced phenotyping are expected to shed new light on our understanding of SRs [156].

Concluding Remarks

NSAIDs are responsible for many DHRs in children [20], including specific immune-mediated reactions, mainly SNIUAA and, albeit to a lesser extent, SNIDRs. Since children often react to a single NSAID whilst tolerating others, attempts should be made to establish the culprit. This is particularly important when the child has taken NSAIDs simultaneously with other drugs or has been exposed to allergens. Paracetamol and ibuprofen are the most common triggers of these reactions. SNIDRs are T cell-mediated and occur much less often in children than in adults. In many instances, these reactions mimic viral reactions: this is particularly problematic if children are taking paracetamol, ibuprofen, or other NSAIDs. In most such cases, the drug is not the true underlying cause of the reaction, and controlled administration is required to verify tolerance.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Conaghan PG. A turbulent decade for NSAIDs: update on current concepts of classification, epidemiology, comparative efficacy, and toxicity. *Rheumatol Int.* 2012;32:1491-502.

2. Rawlins MDT, W. Mechanisms of adverse drug reactions. In: Davies DM ed. Textbook of adverse drug reactions. New York, NY, Oxford University Press. 1991:18-45.
3. Torres MJ, Barrionuevo E, Kowalski M, Blanca M. Hypersensitivity reactions to nonsteroidal anti-inflammatory drugs. *Immunol Allergy Clin North Am.* 2014;34:507-24.
4. Solensky RM, L.M. Drug allergy. In: Leung DYM, Sampson HA, Geha R, Szefer SJ, eds. *Pediatric Allergy, Principles and practice* 2nd edn. St Louis: Mosby Elsevier. 2010;616-30.
5. Eustace N, O'Hare B. Use of nonsteroidal anti-inflammatory drugs in infants. A survey of members of the Association of Paediatric Anaesthetists of Great Britain and Ireland. *Paediatr Anaesth.* 2007;17:464-9.
6. Neubert A, Verhamme K, Murray ML, Picelli G, Hsia Y, Sen FE, Giaquinto C, Ceci A, Sturkenboom M, Wong IC, Excellence TNo. The prescribing of analgesics and non-steroidal anti-inflammatory drugs in paediatric primary care in the UK, Italy and the Netherlands. *Pharmacol Res.* 2010;62:243-8.
7. Valkhoff VE, Schade R, t Jong GW, Romio S, Schuemie MJ, Arfe A, Garbe E, Herings R, Lucchi S, Picelli G, Schink T, Straatman H, Villa M, Kuipers EJ, Sturkenboom MC. Population-based analysis of non-steroidal anti-inflammatory drug use among children in four European countries in the SOS project: what size of data platforms and which study designs do we need to assess safety issues? *BMC Pediatr.* 2013;13:192.
8. Cornejo-Garcia JA, Blanca-Lopez N, Dona I, Andreu I, Agundez JA, Carballo M, Blanca M, Canto MG. Hypersensitivity reactions to non-steroidal anti-inflammatory drugs. *Curr Drug Metab.* 2009;10:971-80.
9. Canto MG, Andreu I, Fernandez J, Blanca M. Selective immediate hypersensitivity reactions to NSAIDs. *Curr Opin Allergy Clin Immunol.* 2009;9:293-7.
10. Dona I, Blanca-Lopez N, Cornejo-Garcia JA, Torres MJ, Laguna JJ, Fernandez J, Rosado A, Rondon C, Campo P, Agundez JA, Blanca M, Canto G. Characteristics of subjects experiencing hypersensitivity to non-steroidal anti-inflammatory drugs: patterns of response. *Clin Exp Allergy.* 2011;41:86-95.
11. Dona I, Blanca-Lopez N, Torres MJ, Garcia-Campos J, Garcia-Nunez I, Gomez F, Salas M, Rondon C, Canto MG, Blanca M. Drug hypersensitivity reactions: response patterns, drug involved, and temporal variations in a large series of patients. *J Investig Allergol Clin Immunol.* 2012;22:363-71.
12. Kowalski ML, Asero R, Bavbek S, Blanca M, Blanca-Lopez N, Bochenek G, Brockow K, Campo P, Celik G, Cernadas J, Cortellini G, Gomes E, Nizankowska-Mogilnicka E, Romano A, Szczeklik A, Testi S, Torres MJ, Wohrl S, Makowska J. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy.* 2013;68:1219-32.
13. Kowalski ML, Makowska JS, Blanca M, Bavbek S, Bochenek G, Bousquet J, Bousquet P, Celik G, Demoly P, Gomes ER, Nizankowska-Mogilnicka E, Romano A, Sanchez-Borges M, Sanz M, Torres MJ, De Weck A, Szczeklik A, Brockow K. Hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs) -classification, diagnosis and management: review of the EAACI/ENDA and GA2LEN/HANNA. *Allergy.* 2011;66:818-29.
14. Pichler WJ. Delayed drug hypersensitivity reactions. *Ann Intern Med.* 2003;139:683-93.
15. Shiohara T, Kano Y. A complex interaction between drug allergy and viral infection. *Clin Rev Allergy Immunol.* 2007;33:124-33.
16. Sanchez-Borges M, Capriles-Behrens E, Caballero-Fonseca F. Hypersensitivity to non-steroidal anti-inflammatory drugs in childhood. *Pediatr Allergy Immunol.* 2004;15:376-80.
17. Blake KV, Zaccaria C, Domergue F, La Mache E, Saint-Raymond A, Hidalgo-Simon A. Comparison between paediatric and adult suspected adverse drug reactions reported to the European medicines agency: implications for pharmacovigilance. *Paediatr Drugs.* 2014;16:309-19.
18. Asero R. Oral aspirin challenges in patients with a history of intolerance to single non-steroidal anti-inflammatory drugs. *Clin Exp Allergy.* 2005;35:713-6.
19. Quiralte J, Blanco C, Delgado J, Ortega N, Alcntara M, Castillo R, Anguita JL, Saenz de San Pedro B, Carrillo T. Challenge-based clinical patterns of 223 Spanish patients with nonsteroidal anti-inflammatory-drug-induced-reactions. *J Investig Allergol Clin Immunol.* 2007;17:182-8.
20. Zambonino MA, Torres MJ, Munoz C, Requena G, Mayorga C, Posadas T, Urda A, Blanca M, Corzo JL. Drug provocation tests in the diagnosis of hypersensitivity reactions to non-steroidal anti-inflammatory drugs in children. *Pediatr Allergy Immunol.* 2013;24:151-9.
21. Khaled A, Kharfi M, Ben Hamida M, El Fekih N, El Aidli S, Zeglaoui F, Ezzine N, Fazaa B, Kamoun MR. Cutaneous adverse drug reactions in children. A series of 90 cases. *Tunis Med.* 2012;90:45-50.
22. Heelan K, Shear NH. Cutaneous drug reactions in children: an update. *Paediatr Drugs.* 2013;15:493-503.
23. Noguera-Morel L, Hernandez-Martin A, Torrelo A. Cutaneous drug reactions in the pediatric population. *Pediatr Clin North Am.* 2014;61:403-26.
24. Mayorga C, Torres MJ, Fernandez J, Canto G, Blanca M. Cutaneous symptoms in drug allergy: what have we learnt? *Curr Opin Allergy Clin Immunol.* 2009;9:431-6.
25. Castell JV, Castell M. Allergic hepatitis induced by drugs. *Curr Opin Allergy Clin Immunol.* 2006;6:258-65.
26. Goodwin SD, Glenny RW. Nonsteroidal anti-inflammatory drug-associated pulmonary infiltrates with eosinophilia. Review of the literature and Food and Drug Administration Adverse Drug Reaction reports. *Arch Intern Med.* 1992;152:1521-4.
27. Squires RH, Jr., Shneider BL, Bucuvalas J, Alonso E, Sokol RJ, Narkewicz MR, Dhawan A, Rosenthal P, Rodriguez-Baez N, Murray KF, Horslen S, Martin MG, Lopez MJ, Soriano H, McGuire BM, Jonas MM, Yazigi N, Shepherd RW, Schwarz K, Lobritto S, Thomas DW, Lavine JE, Karpen S, Ng V, Kelly D, Simonds N, Hynan LS. Acute liver failure in children: the first 348 patients in the pediatric acute liver failure study group. *J Pediatr.* 2006;148:652-8.
28. Porto Arceo JA. Special features of NSAID intolerance in children. *Allergol Immunopathol (Madr).* 2003;31:109-25.
29. Auriel E, Regev K, Korczyn AD. Nonsteroidal anti-inflammatory drugs exposure and the central nervous system. *Handb Clin Neurol.* 2014;119:577-84.
30. Rodriguez-Pena R, Antunez C, Martin E, Blanca-Lopez N, Mayorga C, Torres MJ. Allergic reactions to beta-lactams. *Expert Opin Drug Saf.* 2006;5:31-48.
31. Diaz Jara M, Perez Montero A, Gracia Bara MT, Cabrerizo S, Zapatero L, Martinez Molero MI. Allergic reactions due to ibuprofen in children. *Pediatr Dermatol.* 2001;18:66-7.

32. Hassani A, Ponvert C, Karila C, Le Bourgeois M, De Blic J, Scheinmann P. Hypersensitivity to cyclooxygenase inhibitory drugs in children: a study of 164 cases. *Eur J Dermatol.* 2008;18:561-5.
33. Kidon MI, Kang LW, Chin CW, Hoon LS, See Y, Goh A, Lin JT, Chay OM. Early presentation with angioedema and urticaria in cross-reactive hypersensitivity to nonsteroidal antiinflammatory drugs among young, Asian, atopic children. *Pediatrics.* 2005;116:e675-80.
34. Couto M, Gaspar A, Morais-Almeida M. Selective anaphylaxis to paracetamol in a child. *Eur Ann Allergy Clin Immunol.* 2012;44:163-6.
35. Ozkaya-Bayazit E. Specific site involvement in fixed drug eruption. *J Am Acad Dermatol.* 2003;49:1003-7.
36. Ozkaya-Bayazit E, Bayazit H, Ozarmagan G. Drug related clinical pattern in fixed drug eruption. *Eur J Dermatol.* 2000;10:288-91.
37. Sarkar R, Kaur C, Kanwar AJ. Extensive fixed drug eruption to nimesulide with cross-sensitivity to sulfonamides in a child. *Pediatr Dermatol.* 2002;19:553-4.
38. Belhadjali H, Ghannouchi N, Njim L, Mohamed M, Moussa A, Bayou F, Chakroun M, Zakhama A, Zili J. Acute generalized exanthematous pustulosis induced by buprenorphine in an atopic girl. *Contact Dermatitis.* 2008;58:247-8.
39. Babala J, Brozman M, Blazovsky J. Lyell's syndrome: histological, immunohistochemical and serological observations. *Acta Paediatr Acad Sci Hung.* 1976;17:9-15.
40. Levi N, Bastuji-Garin S, Mockenhaupt M, Roujeau JC, Flahault A, Kelly JP, Martin E, Kaufman DW, Maisson P. Medications as risk factors of Stevens-Johnson syndrome and toxic epidermal necrolysis in children: a pooled analysis. *Pediatrics.* 2009;123:e297-304.
41. Andreu I, Mayorga C, Miranda MA. Generation of reactive intermediates in photoallergic dermatitis. *Curr Opin Allergy Clin Immunol.* 2010;10:303-8.
42. Guillaume JC, Roujeau JC, Chevais M, Albengres E, Revuz J, Touraine R. Lyell's syndrome and ectodermosis pluriorificialis during treatment with oxycams: 11 cases. *Ann Dermatol Venereol.* 1985;112:807-12.
43. Serrano G, Fortea JM, Latasa JM, Millan F, Janes C, Bosca F, Miranda MA. Photosensitivity induced by fibric acid derivatives and its relation to photocontact dermatitis to ketoprofen. *J Am Acad Dermatol.* 1992;27:204-8.
44. Diaz RL, Gardeazabal J, Manrique P, Raton JA, Urrutia I, Rodriguez-Sasiain JM, Aguirre C. Greater allergenicity of topical ketoprofen in contact dermatitis confirmed by use. *Contact Dermatitis.* 2006;54:239-43.
45. Torres MJ, Corzo JL, Leyva L, Mayorga C, Garcia-Martin FJ, Antunez C, Posadas S, Jurado A, Blanca M. Differences in the immunological responses in drug- and virus-induced cutaneous reactions in children. *Blood Cells Mol Dis.* 2003;30:124-31.
46. Pavlos R, Mallal S, Ostrov D, Pompeu Y, Phillips E. Fever, rash, and systemic symptoms: understanding the role of virus and HLA in severe cutaneous drug allergy. *J Allergy Clin Immunol Pract.* 2014;2:21-33.
47. Mayorga C, Pena RR, Blanca-Lopez N, Lopez S, Martin E, Torres MJ. Monitoring the acute phase response in non-immediate allergic drug reactions. *Curr Opin Allergy Clin Immunol.* 2006;6:249-57.
48. Lai M, Huang YC, Yao PC. Tissue necrosis after intramuscular administration of diclofenac: report of one case. *Acta Paediatr Taiwan.* 2005;46:24-6.
49. Srivastava M, Perez-Atayde A, Jonas MM. Drug-associated acute-onset vanishing bile duct and Stevens-Johnson syndromes in a child. *Gastroenterology.* 1998;115:743-6.
50. Faulkner L, Meng X, Park BK, Naisbitt DJ. The importance of hapten-protein complex formation in the development of drug allergy. *Curr Opin Allergy Clin Immunol.* 2014;14:293-300.
51. Meng X, Jenkins RE, Berry NG, Maggs JL, Lane CS, Stachulski AV, French NS, Naisbitt DJ, Pirmohamed M, Park BK. Direct evidence for the formation of diastereoisomeric benzylpenicilloyl haptens from benzylpenicillin and benzylpenicillenic acid in patients. *J Pharmacol Exp Ther.* 2011;338:841-9.
52. Meng X, Howarth A, Earnshaw CJ, Jenkins RE, French NS, Back DJ, Naisbitt DJ, Park BK. Detection of drug bioactivation in vivo: mechanism of nevirapine-albumin conjugate formation in patients. *Chem Res Toxicol.* 2013;26:575-83.
53. Kretz-Rommel A, Boelsterli UA. Diclofenac covalent protein binding is dependent on acyl glucuronide formation and is inversely related to P450-mediated acute cell injury in cultured rat hepatocytes. *Toxicol Appl Pharmacol.* 1993;120:155-61.
54. Ikegawa S, Isriyanthi NM, Nagata M, Yahata K, Ito H, Mano N, Goto J. The enantioselective immunoaffinity extraction of an optically active ibuprofen-modified peptide fragment. *Anal Biochem.* 2001;296:63-72.
55. Zhu D, Becker WM, Schulz KH, Schubeler K, Schlaak M. The presence of specific IgE to salicyloyl and O-methylsalicyloyl in aspirin-sensitive patients. *Asian Pac J Allergy Immunol.* 1992;10:25-32.
56. Blanca M, Perez E, Garcia JJ, Miranda A, Terrados S, Vega JM, Suau R. Angioedema and IgE antibodies to aspirin: a case report. *Ann Allergy.* 1989;62:295-8.
57. Zhu DX, Zhao JL, Mo L, Li HL. Drug allergy: identification and characterization of IgE-reactivities to aspirin and related compounds. *J Investig Allergol Clin Immunol.* 1997;7:160-8.
58. Daxun Z, Becker WM, Schulz KH, Schlaak M. Sensitivity to aspirin: a new serological diagnostic method. *J Investig Allergol Clin Immunol.* 1993;3:72-8.
59. Zhu D, Becker WM, Schulz KH, Schubeler K, Schlaak M. Detection of IgE antibodies specific for 1-phenyl-2,3-dimethyl-3-pyrazoline-5-one by RAST: a serological diagnostic method for sensitivity to pyrazoline drugs. *Asian Pac J Allergy Immunol.* 1992;10:95-101.
60. Kowalski ML, Bienkiewicz B, Woszczyk G, Iwaszkiewicz J, Poniatowska M. Diagnosis of pyrazolone drug sensitivity: clinical history versus skin testing and in vitro testing. *Allergy Asthma Proc.* 1999;20:347-52.
61. Ariza AT, Torres MJ, García-Martín E, Montañez MI, Agúndez JAG, Mayorga C, Doña I, Blanca-Lopez N, Zambonino MA, Andreu I, Blanca M. Basophil activation test with dipyrone metabolites in the evaluation of immediate allergic reactions to dipyrone. *Allergy.* 2014;69 (Suppl 99):133:276.
62. Gomez E, Blanca-Lopez N, Torres MJ, Requena G, Rondon C, Canto G, Blanca M, Mayorga C. Immunoglobulin E-mediated immediate allergic reactions to dipyrone: value of basophil activation test in the identification of patients. *Clin Exp Allergy.* 2009;39:1217-24.

63. Himly M, Jahn-Schmid B, Pittertschatscher K, Bohle B, Grubmayr K, Ferreira F, Ebner H, Ebner C. IgE-mediated immediate-type hypersensitivity to the pyrazolone drug propyphenazone. *J Allergy Clin Immunol.* 2003;111:882-8.
64. Sen I, Mitra S, Gombor KK. Fatal anaphylactic reaction to oral diclofenac sodium. *Can J Anaesth.* 2001;48:421.
65. van Puijenbroek EP, Egberts AC, Meyboom RH, Leufkens HG. Different risks for NSAID-induced anaphylaxis. *Ann Pharmacother.* 2002;36:24-9.
66. Picaud J, Beaudouin E, Renaudin JM, Pirson F, Metz-Favre C, Dron-Gonzalvez M, Moneret-Vautrin DA. Anaphylaxis to diclofenac: nine cases reported to the Allergy Vigilance Network in France. *Allergy.* 2014;69:1420-3.
67. Harrer A, Lang R, Grims R, Braitsch M, Hawranek T, Aberer W, Vogel L, Schmid W, Ferreira F, Himly M. Diclofenac hypersensitivity: antibody responses to the parent drug and relevant metabolites. *PLoS One.* 2010;5:e13707.
68. Morelli JG, Tay YK, Rogers M, Halbert A, Krafchik B, Weston WL. Fixed drug eruptions in children. *J Pediatr.* 1999;134:365-7.
69. Kawakami T, Fujita A, Takeuchi S, Muto S, Soma Y. Drug-induced hypersensitivity syndrome: drug reaction with eosinophilia and systemic symptoms (DRESS) syndrome induced by aspirin treatment of Kawasaki disease. *J Am Acad Dermatol.* 2009;60:146-9.
70. Pliskow S. Severe gynecologic sequelae of Stevens-Johnson syndrome and toxic epidermal necrolysis caused by ibuprofen: a case report. *J Reprod Med.* 2013;58:354-6.
71. Sheridan RL, Schulz JT, Ryan CM, Schnitzer JJ, Lawlor D, Driscoll DN, Donelan MB, Tompkins RG. Long-term consequences of toxic epidermal necrolysis in children. *Pediatrics.* 2002;109:74-8.
72. Ferrandiz-Pulido C, Garcia-Fernandez D, Dominguez-Sampedro P, Garcia-Patos V. Stevens-Johnson syndrome and toxic epidermal necrolysis in children: a review of the experience with paediatric patients in a university hospital. *J Eur Acad Dermatol Venereol.* 2011;25:1153-9.
73. Assem ES. Immunological and non-immunological mechanisms of some of the desirable and undesirable effects of anti-inflammatory and analgesic drugs. *Agents Actions.* 1976;6:212-18.
74. Gomez E, Blanca-Lopez N, Salas M, Canto G, Campo P, Torres MJ, Mayorga C, Blanca M. Induction of accelerated reactions to amoxicillin by T-cell effector mechanisms. *Ann Allergy Asthma Immunol.* 2013;110:267-73.
75. Moreno-Ancillo A, Gil-Adrados AC, Jurado-Palomo J. Ibuprofen-induced aseptic meningoencephalitis confirmed by drug challenge. *J Investig Allergol Clin Immunol.* 2011;21:484-7.
76. Inoue M, Horita N, Kimura N, Kojima R, Miyazawa N. Three cases of mesalazine-induced pneumonitis with eosinophilia. *Respir Investig.* 2014;52:209-12.
77. Blanca M, Romano A, Torres MJ, Fernandez J, Mayorga C, Rodriguez J, Demoly P, Bousquet PJ, Merk HF, Sanz ML, Ott H, Atanaskovic-Markovic M. Update on the evaluation of hypersensitivity reactions to betalactams. *Allergy.* 2009;64:183-93.
78. Blanca M, Torres MJ, Garcia JJ, Romano A, Mayorga C, de Ramon E, Vega JM, Miranda A, Juarez C. Natural evolution of skin test sensitivity in patients allergic to beta-lactam antibiotics. *J Allergy Clin Immunol.* 1999;103:918-24.
79. Torres MJ, Mayorga C, Fernandez TD, Cornejo-Garcia JA, Antunez C, Valenzuela M, Del Prado MF, Rodriguez-Pena R, Blanca M. T cell assessment in allergic drug reactions during the acute phase according to the time of occurrence. *Int J Immunopathol Pharmacol.* 2006;19:119-30.
80. Nettis E, Giordano D, Colanardi MC, Paradiso MT, Ferrannini A, Tursi A. Delayed-type hypersensitivity rash from ibuprofen. *Allergy.* 2003;58:539-40.
81. Mahboob A, Haroon TS. Drugs causing fixed eruptions: a study of 450 cases. *Int J Dermatol.* 1998;37:833-8.
82. Shiohara T, Mizukawa Y. Fixed drug eruption: a disease mediated by self-inflicted responses of intraepidermal T cells. *Eur J Dermatol.* 2007;17:201-8.
83. Roujeau JC, Stern RS. Severe adverse cutaneous reactions to drugs. *N Engl J Med.* 1994;331:1272-85.
84. Um SJ, Lee SK, Kim YH, Kim KH, Son CH, Roh MS, Lee MK. Clinical features of drug-induced hypersensitivity syndrome in 38 patients. *J Investig Allergol Clin Immunol.* 2010;20:556-62.
85. Lee JH, Park HK, Heo J, Kim TO, Kim GH, Kang DH, Song GA, Cho M, Kim DS, Kim HW, Lee CH. Drug Rash with Eosinophilia and Systemic Symptoms (DRESS) syndrome induced by celecoxib and anti-tuberculosis drugs. *J Korean Med Sci.* 2008;23:521-5.
86. Lyell A. Toxic epidermal necrolysis (the scalded skin syndrome): a reappraisal. *Br J Dermatol.* 1979;100:69-86.
87. Correia O, Delgado L, Barbosa IL, Campilho F, Fleming-Torrinha J. Increased interleukin 10, tumor necrosis factor alpha, and interleukin 6 levels in blister fluid of toxic epidermal necrolysis. *J Am Acad Dermatol.* 2002;47:58-62.
88. Miyachi H, Hosokawa H, Akaeda T, Iba H, Asada Y. T-cell subsets in drug-induced toxic epidermal necrolysis. Possible pathogenic mechanism induced by CD8-positive T cells. *Arch Dermatol.* 1991;127:851-5.
89. Paquet P, Pierard GE. Soluble fractions of tumor necrosis factor-alpha, interleukin-6 and of their receptors in toxic epidermal necrolysis: a comparison with second-degree burns. *Int J Mol Med.* 1998;1:459-62.
90. Viard I, Wehrli P, Bullani R, Schneider P, Holler N, Salomon D, Hunziker T, Saurat JH, Tschopp J, French LE. Inhibition of toxic epidermal necrolysis by blockade of CD95 with human intravenous immunoglobulin. *Science.* 1998;282:490-3.
91. Posadas SJ, Padial A, Torres MJ, Mayorga C, Leyva L, Sanchez E, Alvarez J, Romano A, Juarez C, Blanca M. Delayed reactions to drugs show levels of perforin, granzyme B, and Fas-L to be related to disease severity. *J Allergy Clin Immunol.* 2002;109:155-61.
92. Nassif A, Moslehi H, Le Gouvello S, Bagot M, Lyonnet L, Michel L, Boumsell L, Bensussan A, Roujeau JC. Evaluation of the potential role of cytokines in toxic epidermal necrolysis. *J Invest Dermatol.* 2004;123:850-5.
93. Tohyama M, Hashimoto K. Immunological mechanisms of epidermal damage in toxic epidermal necrolysis. *Curr Opin Allergy Clin Immunol.* 2012;12:376-82.
94. Edell DS, Davidson JJ, Muelenaer AA, Majure M. Unusual manifestation of Stevens-Johnson syndrome involving the respiratory and gastrointestinal tract. *Pediatrics.* 1992;89:429-32.
95. Carbone C, Rende P, Comberlati P, Carnovale D, Mammi M, De Sarro G. The safety of ketoprofen in different ages. *J Pharmacol Pharmacother.* 2013;4:S99-S103.

96. Fernandez TD, Torres MJ, Blanca-Lopez N, Rodriguez-Bada JL, Gomez E, Canto G, Mayorga C, Blanca M. Negativization rates of IgE radioimmunoassay and basophil activation test in immediate reactions to penicillins. *Allergy*. 2009;64:242-8.
97. Aranda A, Mayorga C, Ariza A, Dona I, Blanca-Lopez N, Canto G, Blanca M, Torres MJ. IgE-mediated hypersensitivity reactions to methylprednisolone. *Allergy*. 2010;65:1376-80.
98. Moreno F, Blanca M, Mayorga C, Terrados S, Moya M, Perez E, Suau R, Vega JM, Garcia J, Miranda A, et al. Studies of the specificities of IgE antibodies found in sera from subjects with allergic reactions to penicillins. *Int Arch Allergy Immunol*. 1995;108:74-81.
99. Bochenek G, Nizankowska E, Szczeklik A. The atopy trait in hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*. 1996;51:16-23.
100. Kemp SF, Lockey RF. Anaphylaxis: a review of causes and mechanisms. *J Allergy Clin Immunol*. 2002;110:341-8.
101. Simons FE. 9. Anaphylaxis. *J Allergy Clin Immunol*. 2008;121:S402-7.
102. Pirmohamed M. Personalized pharmacogenomics: predicting efficacy and adverse drug reactions. *Annu Rev Genomics Hum Genet*. 2014;15:349-70.
103. Ponvert C. Allergic and non-allergic hypersensitivity to non-opioid analgesics, antipyretics and nonsteroidal anti-inflammatory drugs in children: epidemiology, clinical aspects, pathophysiology, diagnosis and prevention. *Arch Pediatr*. 2012;19:556-60.
104. Ponvert C, Scheinmann P. Allergic and pseudoallergic reactions to analgesics, antipyretics and non-steroidal anti-inflammatory drugs. *Arch Pediatr*. 2007;14:507-12.
105. Brockow K, Garvey LH, Aberer W, Atanaskovic-Markovic M, Barbaud A, Bilo MB, Bircher A, Blanca M, Bonadonna B, Campi P, Castro E, Cernadas JR, Chiriack AM, Demoly P, Grosber M, Gooi J, Lombardo C, Mertes PM, Mosbech H, Nasser S, Pagani M, Ring J, Romano A, Scherer K, Schnyder B, Testi S, Torres M, Trautmann A, Terreehorst I, Group EEDA. Skin test concentrations for systemically administered drugs - an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy*. 2013;68:702-12.
106. Gamboa PM, Sanz ML, Caballero MR, Antepará I, Urrutia I, Jauregui I, Gonzalez G, Dieguez I, De Weck AL. Use of CD63 expression as a marker of in vitro basophil activation and leukotriene determination in metamizol allergic patients. *Allergy*. 2003;58:312-7.
107. Barbaud A. Skin testing and patch testing in non-IgE-mediated drug allergy. *Curr Allergy Asthma Rep*. 2014;14:442.
108. Na HR, Lee JM, Jung JW, Lee SY. Usefulness of drug provocation tests in children with a history of adverse drug reaction. *Korean J Pediatr*. 2011;54:304-9.
109. Mayorga C, Sanz ML, Gamboa P, Garcia-Aviles MC, Fernandez J, Torres MJ, Spanish Society of A, Clinical, Immunology, Immunology, Drug Allergy C. In vitro methods for diagnosing nonimmediate hypersensitivity reactions to drugs. *J Investig Allergol Clin Immunol*. 2013;23:213-25.
110. Goncalo M, Ferguson J, Bonevalle A, Bruynzeel DP, Gimenez-Arnau A, Goossens A, Kerr A, Lecha M, Neumann N, Niklasson B, Pigatto P, Rhodes LE, Rustemeyer T, Sarkany R, Thomas P, Wilkinson M. Photopatch testing: recommendations for a European photopatch test baseline series. *Contact Dermatitis*. 2013;68:239-43.
111. Steiner M, Harrer A, Lang R, Schneider M, Ferreira T, Hawranek T, Himly M. Basophil activation test for investigation of IgE-mediated mechanisms in drug hypersensitivity. *J Vis Exp*. 2011;55:3263.
112. Blanca M, Posadas S, Torres MJ, Leyva L, Mayorga C, Gonzalez L, Juarez C, Fernandez J, Santamaria LF. Expression of the skin-homing receptor in peripheral blood lymphocytes from subjects with nonimmediate cutaneous allergic drug reactions. *Allergy*. 2000;55:998-1004.
113. Fernandez J, Blanca M, Moreno F, Garcia J, Segurado E, del Cano A, Aguilar F. Role of tryptase, eosinophil cationic protein and histamine in immediate allergic reactions to drugs. *Int Arch Allergy Immunol*. 1995;107:160-2.
114. Yilmaz O, Ertoý Karagol IH, Bakirtas A, Topal E, Celik GE, Demirsoy MS, Turktas I. Challenge-proven nonsteroidal anti-inflammatory drug hypersensitivity in children. *Allergy*. 2013;68:1555-61.
115. Lerch M, Pichler WJ. The immunological and clinical spectrum of delayed drug-induced exanthems. *Curr Opin Allergy Clin Immunol*. 2004;4:411-9.
116. Fernandez TD, Canto G, Blanca M. Molecular mechanisms of maculopapular exanthema. *Curr Opin Infect Dis*. 2009;22:272-8.
117. Aberer W, Bircher A, Romano A, Blanca M, Campi P, Fernandez J, Brockow K, Pichler WJ, Demoly P, European Network for Drug A, hypersensitivity Eigod. Drug provocation testing in the diagnosis of drug hypersensitivity reactions: general considerations. *Allergy*. 2003;58:854-63.
118. Ordoqui E, De Barrio M, Rodriguez VM, Herrero T, Gil PJ, Baeza ML. Cross-sensitivity among oxicams in piroxicam-caused fixed drug eruption: two case reports. *Allergy*. 1995;50:741-4.
119. Gomez-Traseira C, Rojas-Perez-Ezquerria P, Sanchez-Morillas L, Gonzalez-Mendiola R, Rubio-Perez M, Moral-Morales A, Juliolaguna-Martinez J. Paracetamol-induced fixed drug eruption at an unusual site. *Recent Pat Inflamm Allergy Drug Discov*. 2013;7:268-70.
120. Romano A, Torres MJ, Castells M, Sanz ML, Blanca M. Diagnosis and management of drug hypersensitivity reactions. *J Allergy Clin Immunol*. 2011;127:S67-73.
121. Sicherer SH, Sampson HA. Food allergy: Epidemiology, pathogenesis, diagnosis, and treatment. *J Allergy Clin Immunol*. 2014;133:291-307.
122. Hansen RC. Staphylococcal scalded skin syndrome, toxic shock syndrome, and Kawasaki disease. *Pediatr Clin North Am*. 1983;30:533-44.
123. Green D, Law E, Still JM. An approach to the management of toxic epidermal necrolysis in a burn centre. *Burns*. 1993;19:411-4.
124. Fritsch PO, Sidoroff A. Drug-induced Stevens-Johnson syndrome/toxic epidermal necrolysis. *Am J Clin Dermatol*. 2000;1:349-60.
125. Leaute-Labreze C, Lamireau T, Chawki D, Maleville J, Taieb A. Diagnosis, classification, and management of erythema multiforme and Stevens-Johnson syndrome. *Arch Dis Child*. 2000;83:347-52.
126. Tay YK, Huff JC, Weston WL. Mycoplasma pneumoniae infection is associated with Stevens-Johnson syndrome, not erythema multiforme (von Hebra). *J Am Acad Dermatol*. 1996;35:757-60.
127. Gonzalez FJ, Carvajal MJ, del Pozo V, Lahoz C, Santamaria L, Blanca M, Juarez C. Erythema multiforme to phenobarbital:

- involvement of eosinophils and T cells expressing the skin homing receptor. *J Allergy Clin Immunol.* 1997;100:135-7.
128. Romano A, Blanca M, Torres MJ, Bircher A, Aberer W, Brockow K, Pichler WJ, Demoly P, Enda, Eaaci. Diagnosis of nonimmediate reactions to beta-lactam antibiotics. *Allergy.* 2004;59:1153-60.
 129. Chiriac AM, Demoly P. Multiple drug hypersensitivity syndrome. *Curr Opin Allergy Clin Immunol.* 2013;13:323-9.
 130. Sheridan RL, Weber JM, Schulz JT, Ryan CM, Low HM, Tompkins RG. Management of severe toxic epidermal necrolysis in children. *J Burn Care Rehabil.* 1999;20:497-500.
 131. Guibal F, Bastuji-Garin S, Chosidow O, Saiag P, Revuz J, Roujeau JC. Characteristics of toxic epidermal necrolysis in patients undergoing long-term glucocorticoid therapy. *Arch Dermatol.* 1995;131:669-72.
 132. Goyal S, Gupta P, Ryan CM, Kazlas M, Noviski N, Sheridan RL. Toxic epidermal necrolysis in children: medical, surgical, and ophthalmologic considerations. *J Burn Care Res.* 2009;30:437-49.
 133. Tristani-Firouzi P, Petersen MJ, Saffle JR, Morris SE, Zone JJ. Treatment of toxic epidermal necrolysis with intravenous immunoglobulin in children. *J Am Acad Dermatol.* 2002;47:548-52.
 134. Koh MJ, Tay YK. Stevens-Johnson syndrome and toxic epidermal necrolysis in Asian children. *J Am Acad Dermatol.* 2010;62:54-60.
 135. Spies M, Sanford AP, Aili Low JF, Wolf SE, Herndon DN. Treatment of extensive toxic epidermal necrolysis in children. *Pediatrics.* 2001;108:1162-8.
 136. Yang Y, Xu J, Li F, Zhu X. Combination therapy of intravenous immunoglobulin and corticosteroid in the treatment of toxic epidermal necrolysis and Stevens-Johnson syndrome: a retrospective comparative study in China. *Int J Dermatol.* 2009;48:1122-8.
 137. Kardaun SH, Jonkman MF. Dexamethasone pulse therapy for Stevens-Johnson syndrome/toxic epidermal necrolysis. *Acta Derm Venereol.* 2007;87:144-8.
 138. Schneck J, Fagot JP, Sekula P, Sassolas B, Roujeau JC, Mockenhaupt M. Effects of treatments on the mortality of Stevens-Johnson syndrome and toxic epidermal necrolysis: A retrospective study on patients included in the prospective EuroSCAR Study. *J Am Acad Dermatol.* 2008;58:33-40.
 139. French LE, Trent JT, Kerdel FA. Use of intravenous immunoglobulin in toxic epidermal necrolysis and Stevens-Johnson syndrome: our current understanding. *Int Immunopharmacol.* 2006;6:543-9.
 140. Mittmann N, Chan B, Knowles S, Cosentino L, Shear N. Intravenous immunoglobulin use in patients with toxic epidermal necrolysis and Stevens-Johnson syndrome. *Am J Clin Dermatol.* 2006;7:359-68.
 141. Bachot N, Revuz J, Roujeau JC. Intravenous immunoglobulin treatment for Stevens-Johnson syndrome and toxic epidermal necrolysis: a prospective noncomparative study showing no benefit on mortality or progression. *Arch Dermatol.* 2003;139:33-6.
 142. Shortt R, Gomez M, Mittman N, Cartotto R. Intravenous immunoglobulin does not improve outcome in toxic epidermal necrolysis. *J Burn Care Rehabil.* 2004;25:246-55.
 143. Brown KM, Silver GM, Halerz M, Walaszek P, Sandroni A, Gamelli RL. Toxic epidermal necrolysis: does immunoglobulin make a difference? *J Burn Care Rehabil.* 2004;25:81-8.
 144. Barron SJ, Del Vecchio MT, Aronoff SC. Intravenous immunoglobulin in the treatment of Stevens-Johnson syndrome and toxic epidermal necrolysis: a meta-analysis with meta-regression of observational studies. *Int J Dermatol.* 2015;54:108-15.
 145. Morici MV, Galen WK, Shetty AK, Lebouef RP, Gouri TP, Cowan GS, Gedalia A. Intravenous immunoglobulin therapy for children with Stevens-Johnson syndrome. *J Rheumatol.* 2000;27:2494-7.
 146. Mangla K, Rastogi S, Goyal P, Solanki RB, Rawal RC. Efficacy of low dose intravenous immunoglobulins in children with toxic epidermal necrolysis: an open uncontrolled study. *Indian J Dermatol Venereol Leprol.* 2005;71:398-400.
 147. Koh MJ, Tay YK. An update on Stevens-Johnson syndrome and toxic epidermal necrolysis in children. *Curr Opin Pediatr.* 2009;21:505-10.
 148. Inamo Y, Okubo T, Wada M, Fuchigami S, Hashimoto K, Fuchigami T, Takahashi S, Sawada S, Harada K. Intravenous ulinastatin therapy for Stevens-Johnson syndrome and toxic epidermal necrolysis in pediatric patients. Three case reports. *Int Arch Allergy Immunol.* 2002;127:89-94.
 149. Mayorga C, Torres MJ, Corzo JL, Sanchez-Sabate E, Alvarez J, Vera A, Posadas S, Jurado A, Blanca M. Improvement of toxic epidermal necrolysis after the early administration of a single high dose of intravenous immunoglobulin. *Ann Allergy Asthma Immunol.* 2003;91:86-91.
 150. Fernando SL. Acute generalised exanthematous pustulosis. *Australas J Dermatol.* 2012;53:87-92.
 151. Sultan SJ, Sameem F, Ashraf M. Drug reaction with eosinophilia and systemic symptoms: manifestations, treatment, and outcome in 17 patients. *Int J Dermatol.* 2015;54:537-42.
 152. Stevenson DD. Aspirin sensitivity and desensitization for asthma and sinusitis. *Curr Allergy Asthma Rep.* 2009;9:155-63.
 153. Castells M. Rapid desensitization for hypersensitivity reactions to medications. *Immunol Allergy Clin North Am.* 2009;29:585-606.
 154. Swierczynska-Krepa M, Sanak M, Bochenek G, Strek P, Cmiel A, Gielicz A, Plutecka H, Szczeklik A, Nizankowska-Mogilnicka E. Aspirin desensitization in patients with aspirin-induced and aspirin-tolerant asthma: A double-blind study. *J Allergy Clin Immunol.* 2014;134:883-90.
 155. Woessner KM, White AA. Evidence-based approach to aspirin desensitization in aspirin-exacerbated respiratory disease. *J Allergy Clin Immunol.* 2014;133:286-7.
 156. Perkins JR, Sanak M, Canto G, Blanca M, Cornejo-Garcia JA. Unravelling adverse reactions to NSAIDs using systems biology. *Trends Pharmacol Sci.* 2015;36:172-80.

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