

Supplementary data

The circulating T helper subsets and regulatory T cells in patients with common variable immunodeficiency with no monogenic disease

Azizi et al.

Table S1. Excluded 373 genes variants in whole exome sequencing of COVID patients.

Genes symbols							
<i>ACP5</i>	<i>CD27</i>	<i>DEPTOR</i>	<i>IGHA2</i>	<i>LIG4</i>	<i>NOLA2</i>	<i>ROBLD3</i>	<i>TLR3</i>
<i>ACT1</i>	<i>CD28</i>	<i>DGCR</i>	<i>IGHE</i>	<i>LPIN2</i>	<i>NOLA3</i>	<i>RPSA</i>	<i>TMC6</i>
<i>ACTB</i>	<i>CD3D</i>	<i>DKC1</i>	<i>IGHG1</i>	<i>LRBA</i>	<i>NOP10</i>	<i>RORC</i>	<i>TMC8</i>
<i>ADA</i>	<i>CD3E</i>	<i>DNMT3B</i>	<i>IGHG2</i>	<i>LRRC8A</i>	<i>NOTCH1</i>	<i>RSPA</i>	<i>TMEM173</i>
<i>ADAR1</i>	<i>CD3G</i>	<i>DOCK2</i>	<i>IGHG3</i>	<i>LYST</i>	<i>PNP</i>	<i>RTEL1</i>	<i>TNFAIP3</i>
<i>ADAM17</i>	<i>CD40</i>	<i>DONSON</i>	<i>IGHG4</i>	<i>MAP3K14</i>	<i>NRAS</i>	<i>RLTPR</i>	<i>TNFRSF11A</i>
<i>AICDA</i>	<i>CD40LG</i>	<i>DOCK8</i>	<i>IGHM</i>	<i>MAGT1</i>	<i>NSMCE3</i>	<i>RTP4</i>	<i>TNFRSF13B</i>
<i>AIRE</i>	<i>CD46</i>	<i>DRB1</i>	<i>IGKC</i>	<i>MALT1</i>	<i>ORAI1</i>	<i>SAMHD1</i>	<i>TNFRSF13C</i>
<i>AK2</i>	<i>CD55</i>	<i>E47</i>	<i>IGLL1</i>	<i>MASP1</i>	<i>OX40</i>	<i>SBDS</i>	<i>TNFRSF1A</i>
<i>AP3B1</i>	<i>CD59</i>	<i>ELANE</i>	<i>IKAROS</i>	<i>MASP2</i>	<i>PARN</i>	<i>SEC16A</i>	<i>TNFRSF6</i>
<i>AP3D</i>	<i>CD70</i>	<i>ELF4</i>	<i>IKBA</i>	<i>MBL2</i>	<i>PAX5</i>	<i>SERPING1</i>	<i>TNFSF6</i>
<i>APOL1</i>	<i>CD79A</i>	<i>EPG5</i>	<i>IKBKB</i>	<i>MCM10</i>	<i>PGM3</i>	<i>SH2D1A</i>	<i>TRAC</i>
<i>ARPC1B</i>	<i>CD79B</i>	<i>EVER1</i>	<i>IKBKG</i>	<i>MCM4</i>	<i>PIGA</i>	<i>SH3BP2</i>	<i>TRAF3</i>
<i>ATM</i>	<i>CD81</i>	<i>EVER2</i>	<i>IKZF1</i>	<i>MDA5</i>	<i>PIK3CD</i>	<i>SLC11A1</i>	<i>TREX1</i>
<i>BCL10</i>	<i>CD8A</i>	<i>EVER3</i>	<i>IL10</i>	<i>MEFV</i>	<i>PIK3R1</i>	<i>SLC29A3</i>	<i>TRNT1</i>
<i>BLM</i>	<i>CDCA7</i>	<i>F12</i>	<i>IL10RA</i>	<i>mircl</i>	<i>PLCG2</i>	<i>SLC35C1</i>	<i>TTC37</i>
<i>BLNK</i>	<i>CEBPE</i>	<i>FADD</i>	<i>IL10RB</i>	<i>MKL1</i>	<i>PLDN</i>	<i>SLC37A4</i>	<i>TTC7A</i>
<i>BTK</i>	<i>CECRI/ADA2</i>	<i>FANCA</i>	<i>IL12B</i>	<i>MLPH</i>	<i>PMM2</i>	<i>SLC46A1</i>	<i>TWEAK</i>
<i>B2M</i>	<i>CFB</i>	<i>FANCE</i>	<i>IL12RB1</i>	<i>MOGS</i>	<i>PMS2</i>	<i>SMC5/6</i>	<i>TYK2</i>
<i>C16orf57</i>	<i>CFD</i>	<i>FAS</i>	<i>IL17F</i>	<i>MPO</i>	<i>POLE1</i>	<i>SMARCAL1</i>	<i>TPP1</i>
<i>C1QA</i>	<i>CFH</i>	<i>FASLG</i>	<i>IL17RA</i>	<i>MRE11A</i>	<i>PRF1</i>	<i>SP110</i>	<i>TPP2</i>
<i>C1QB</i>	<i>CFHR1</i>	<i>FCGR1A</i>	<i>IL1RN</i>	<i>MSH6</i>	<i>PRKCD</i>	<i>SPINK5</i>	<i>UNC119</i>
<i>C1QC</i>	<i>CFHR3</i>	<i>FCGR3A</i>	<i>IL21</i>	<i>MST1</i>	<i>PRKDC</i>	<i>STAT1</i>	<i>UNC13D</i>
<i>C1R</i>	<i>CFHR5</i>	<i>FCN3</i>	<i>IL21R</i>	<i>MSN</i>	<i>PSMB8</i>	<i>STAT2</i>	<i>UNC93B1</i>
<i>C1S</i>	<i>CFI</i>	<i>FERMT3</i>	<i>IL23</i>	<i>MTHFD1</i>	<i>PTEN</i>	<i>STAT3</i>	<i>UNG</i>
<i>C2</i>	<i>CFP</i>	<i>FH</i>	<i>IL2RA</i>	<i>MUC2</i>	<i>PSTPIP1</i>	<i>STAT5B</i>	<i>VAV1</i>
<i>C3</i>	<i>CHD7</i>	<i>FH3</i>	<i>IL2RG</i>	<i>CD20</i>	<i>PTPRC</i>	<i>STIM1</i>	<i>VPREB1</i>
<i>C4A</i>	<i>CIAS1</i>	<i>FOXP1</i>	<i>IL36RN</i>	<i>MVK</i>	<i>RAB27A</i>	<i>STK4</i>	<i>VPS13B</i>
<i>C4B</i>	<i>CIITA</i>	<i>FOXP3</i>	<i>IL7R</i>	<i>MYD88</i>	<i>RAC2</i>	<i>STN1</i>	<i>VPS45</i>
<i>C4BPA</i>	<i>CLEC16A</i>	<i>FPR1</i>	<i>INO80</i>	<i>MYO5A</i>	<i>RAG1</i>	<i>STX11</i>	<i>WAS</i>
<i>C4BPB</i>	<i>CLEC7A</i>	<i>FUCT1</i>	<i>IRAK4</i>	<i>NBN</i>	<i>RAG2</i>	<i>STXBP2</i>	<i>WASF2</i>
<i>C5</i>	<i>CLPB</i>	<i>G6PC</i>	<i>IRF8</i>	<i>NBS1</i>	<i>RASGRP1</i>	<i>SEC61A1</i>	<i>WIPF1</i>
<i>C6</i>	<i>CMCD7</i>	<i>G6PC3</i>	<i>IRF7</i>	<i>NCF1</i>	<i>RASGRP2</i>	<i>SPPL2A</i>	<i>WRAP53</i>
<i>C7</i>	<i>CMCD8</i>	<i>G6PD</i>	<i>IRFBP2</i>	<i>NCF2</i>	<i>RBCK1</i>	<i>TAP1</i>	<i>XIAP</i>
<i>C8A</i>	<i>COH1</i>	<i>G6PT1</i>	<i>ISG15</i>	<i>NCF4</i>	<i>RC3H1</i>	<i>TAP2</i>	<i>ZAP70</i>
<i>C8B</i>	<i>COLEC11</i>	<i>GATA2</i>	<i>ITCH</i>	<i>NEIL1</i>	<i>RECQL4</i>	<i>TAPBP</i>	<i>ZBTB24</i>
<i>C8G</i>	<i>CORO1A</i>	<i>GFII</i>	<i>ITGA3</i>	<i>NEIL3</i>	<i>RFX5</i>	<i>TAZ</i>	<i>ZNF341</i>
<i>C9</i>	<i>COPA</i>	<i>GIAMP5</i>	<i>ITGB2</i>	<i>NFAT5</i>	<i>RFXANK</i>	<i>TBK1</i>	
<i>CARD11</i>	<i>CSF2RA</i>	<i>HAX1</i>	<i>ITK</i>	<i>NFKB1</i>	<i>RFXAP</i>	<i>TBX1</i>	
<i>CARD14</i>	<i>CSF3R</i>	<i>HELLS</i>	<i>JAGN1</i>	<i>NFKB2</i>	<i>RHOH</i>	<i>TCF3</i>	
<i>CARD9</i>	<i>CTLA4</i>	<i>HOIL1</i>	<i>JAK2</i>	<i>NFKBIA</i>	<i>RIPK1</i>	<i>TCIRG1</i>	
<i>CASP10</i>	<i>CTSC</i>	<i>HOIP</i>	<i>JAK3</i>	<i>NFKBID</i>	<i>RLTPR</i>	<i>TCN2</i>	
<i>CASP8</i>	<i>CTPS1</i>	<i>ICOS</i>	<i>KMT2A</i>	<i>NHEJ1</i>	<i>RMRP</i>	<i>TERC</i>	
<i>CCBE1</i>	<i>CXCR4</i>	<i>IFNGR1</i>	<i>KINDLIN3</i>	<i>NHP2</i>	<i>RNASEH2A</i>	<i>TERT</i>	
<i>CD16</i>	<i>CYBA</i>	<i>IFNGR2</i>	<i>KRAS</i>	<i>NLRP12</i>	<i>RNASEH2B</i>	<i>TFRC</i>	
<i>CD19</i>	<i>CYBB</i>	<i>IFIH1</i>	<i>LAT</i>	<i>NLRC4</i>	<i>RNASEH2C</i>	<i>THBD</i>	
<i>CD21</i>	<i>DCLRE1B</i>	<i>IGAD1</i>	<i>LCK</i>	<i>NLRP3</i>	<i>RNU4ATAC</i>	<i>TICAM1</i>	
<i>CD247</i>	<i>DCLRE1C</i>	<i>IGHA1</i>	<i>LIG1</i>	<i>NOD2</i>	<i>RNF168</i>	<i>TINF2</i>	

Table S2. Oligonucleotide sequences of primers used in this study for quantitative real time PCR.

Genes	Forward	Reverse
<i>IFNG</i>	F; 5'-GAGTGTGGAGACCATCAAGGAA-3'	R; 3'-GGCGACAGTTCAGCCATCA-5'
<i>IL17</i>	F; 5'-CTACAACCGATCCACCTCACC-3'	R; 3'-CCCACGGACACCAGTATCTTC-5'
<i>IL22</i>	F; 5'-CCTTGAAGAAGTGCTGTTCCT-3'	R; 3'-CCTTCAGCTTTTGCACATTCCT-5'
<i>IL10</i>	F; 5'-GACTTTAAGGGTTACCTGGGTTG-3'	R; 3'-TCTTGGTTCTCAGCTTGGGG-5'
<i>TBET</i>	F; 5'-GTCGCGCTCAACAACCAC-3'	R; 3'-GGAACATCCGCCGTCCCT-5'
<i>RORC</i>	F; 5'-TTCCGAGGATGAGATTGCCC-3'	R; 3'-CAGCTTTGCCAGGATGCTTT-5'
<i>AHR</i>	F; 5'-GTCGTCTAAGGTGTCTGCTGG-3'	R; 3'-TATGGATGGTGGCTGAAGTG-5'
<i>FOXP3</i>	F; 5'-ACAACATGCGACCCCTTT-3'	R; 3'-TGGCGGATGGCGTTCTTC-5'
<i>RUNX1</i>	F; 5'-AATGCTACCGCAGCCATGAA-3'	R; 3'-CTTGCGGTGGGTTTGTGAAG-5'
<i>GAPDH</i>	F; 5'-GAGAAGGCTGGGGCTCATTT-3'	R; 3'-TAAGCAGTTGGTGGTGCAGG-5'

Table S3. The absolute counts of the investigated lymphocyte subsets in COVID patients and Healthy controls.

Subset; cell/ μ L	COVID patients (n=13)	Healthy controls (n=13)	P value
Lymphocyte($\times 10^3$)	2.00 (1.30-2.70)	2.20 (1.80-2.40)	0.459
CD4 ⁺ T cells	567 (359-750)	936 (842-1027)	0.003*
CD4 ⁺ IFN- γ ⁺ IL-17 ⁻ (Th1)	72 (29-176)	112 (101-143)	0.174
CD4 ⁺ IFN- γ ⁺ IL-17 ⁺ (Th1-like Th17)	4.66 (1.21-6.29)	4.62 (3.62-5.92)	0.596
CD4 ⁺ IFN- γ ⁻ IL-17 ⁺ (Th17)	1.50 (0.46-4.10)	5.92 (4.89-7.67)	0.001*
CD4 ⁺ IFN- γ ⁻ IL-17 ⁻ IL-22 ⁺ (Th22)	4.68 (0.99-13.48)	4.88 (3.69-5.92)	0.800
CD4 ⁺ CD25 ⁺ FoxP3 ⁺ (Treg)	3.47 (1.76-10.11)	19.40 (14.62-22.90)	<0.001*
CD4 ⁺ CD25 ⁺ FoxP3 ⁺ CD127 ^{low/-} (CD127 ^{low/-} Treg)	2.40 (1.05-12.19)	17.69 (12.55-22.48)	<0.001*

Data are presented as median (75th-25th interquartile range).

*P values <0.05 were regarded significant.

Figure S1.

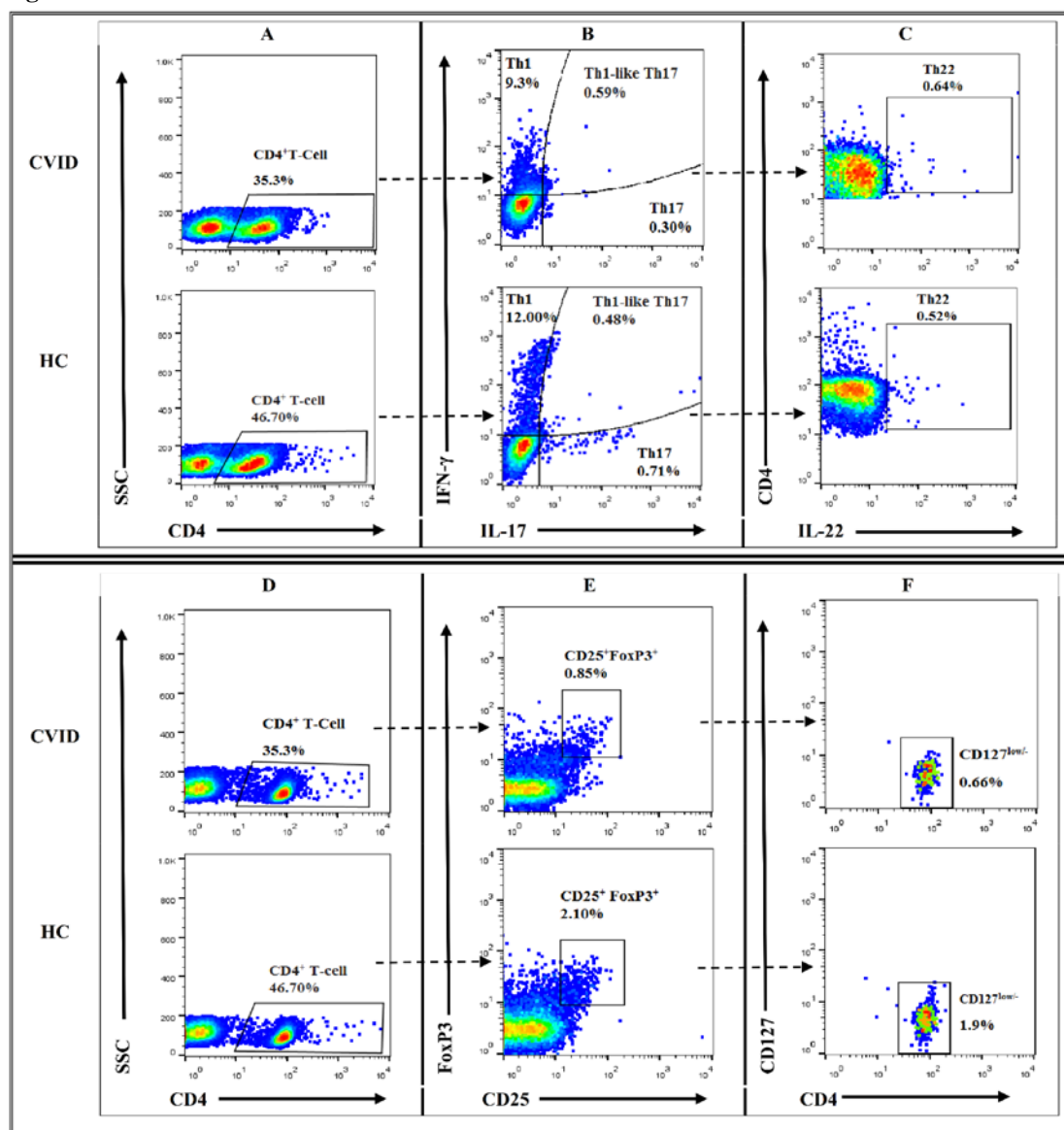


Figure S1. Flow cytometry analysis of different subsets of CD4⁺ T cells. Two examples demonstrate the flow cytometric analysis of peripheral blood samples (PBMCs) from one CVID patient and a healthy control (HC). (A-C) Th subsets: PBMCs from subjects were prepared and cultured with PMA and ionomycin for 5 hours, and then cells were harvested. The cells were stained with anti-CD4 PerCP-cy5.5, fixed, and permeabilized, followed by intracellular staining with anti-IFN- γ FITC, anti-IL-17 PE, and anti-IL-22 APC. CD4⁺ T-cells were gated within the lymphocyte scatter region (A) for analysis of the CD4⁺ IFN γ ⁺ IL-17⁻, CD4⁺ IFN γ ⁺ IL-17⁺, CD4⁺ IFN γ ⁻ IL-17⁺ cells (B). The CD4⁺ IFN γ ⁻ IL-17⁻ cells were further analyzed for CD4⁺ IFN γ ⁻ IL-17⁻ IL-22⁺ cells (C). (D-F) Treg analysis: PBMCs were isolated and stained with anti-CD4 PerCP-cy5.5, anti-CD25 APC, anti-CD127 FITC, fixed, and permeabilized, followed by intracellular staining with anti-Foxp3 PE. Then, the cells were gated first on CD4⁺ cells for analysis of the CD4⁺ CD25⁺ FoxP3⁺. CD4⁺ CD25⁺ FoxP3⁺ cells were further analyzed for CD4⁺ CD25⁺ FoxP3⁺ CD127^{low/-} cells. Data are expressed as the median values of individual participants. Arrows indicate the gated population subsequently analyzed.