Profilin, a Change in the Paradigm

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Abstract

Profilin is a protein that is present in all eukaryotic cells and is responsible for cross-reactivity between pollen, latex, and plant foods. It has been classically acknowledged as a minor or nearly irrelevant allergen, although recent data are changing this conception. The objective of this manuscript is to provide a comprehensive review of published data on the role of this ubiquitous allergen in pollen, latex, and plant food allergy.

The patterns of recognition of this minor allergen follow a north-south gradient. Although present in all pollens and vegetables, profilin is significantly associated with allergy to grass pollen and to Cucurbitaceae fruits. Heb v 8, the latex profilin, is usually a marker of profilin allergy in plant food–allergic patients, although it has no clinical relevance in latex allergy. Sensitization to profilin jeopardizes the diagnosis of pollen allergy and selection of immunotherapy, and although component-resolved diagnosis can identify its impact, there are no tailored treatments available. In recent years, several new publications have shown how profilin should be taken into account and, under certain circumstances, considered a marker of severity, an allergen capable of inducing respiratory symptoms, and, in its natural purified form, a potential candidate for etiological treatment of food allergy.

Current data on profilin strongly support the need for a shift in the previously accepted paradigm for this allergen. More research should be done to assess the real clinical impact of sensitization in specific populations and to develop therapeutic strategies.


Resumen

La profilina es una proteína presente en todas las células eucariotas, siendo responsable de la reactividad cruzada entre polen, látex y alimentos vegetales. Ha sido reconocida clásicamente como un alérgeno menor o irrelevante; sin embargo, datos recientemente publicados están modificando esta interpretación. El objetivo de este manuscrito es realizar una revisión comprensiva de la literatura sobre el papel de este ubicuo alérgeno en el polen, látex y los alimentos vegetales.

El patrón de reconocimiento de este alérgeno menor sigue un gradiente de norte a sur, y a pesar de estar presente en todos los pólens y vegetales, está significativamente asociado al polen de gramíneas y a las frutas de la familia Cucurbitaceae. Heb v 8, la profilina del látex, es habitualmente un marcador de alergia a profilina en pacientes alérgicos a vegetales pero sin relevancia clínica en la alergia a látex. La presencia de la sensibilización a profilina dificulta el diagnóstico de alergia a pólens y la selección de la inmunoterapia, y a pesar de que el diagnóstico por componentes puede identificar su impacto, no existen tratamientos personalizados disponibles. En los últimos años, diversas publicaciones nuevas han demostrado como la profilina debe ser tenida en cuenta y considerada bajo determinadas circunstancias, como un marcador de gravedad, como un alérgeno capaz de inducir síntomas respiratorios, y en su forma natural purificada, como un potencial candidato para realizar un tratamiento etiológico para tratar la alergia a alimentos.

El conocimiento actual sobre la profilina impulsa la necesidad de cambiar el concepto que previamente se tenía sobre este alérgeno. Sería preciso investigar más para valorar el impacto clínico real de esta sensibilización en poblaciones específicas y desarrollar estrategias terapéuticas.

Introduction

Profilins are 12 to 16-kDa, actin monomer-binding proteins expressed in specific viruses and in all eukaryotic cells, with the exception of some protists [1,2]. Profilins promote polymerization of actin filaments and monomers and are thus involved in the generation of the cytoskeleton and in movement [1]. Their role in such essential processes explains their ubiquitous expression and high levels of conservation [3] (Figure). The identification of 50 additional profilin ligands suggests an important role in many more complex molecular processes, as well as in signal transduction [2,4]. The first allergenic profilin described, Bet v 2 from birch pollen, was identified in 1991 [5], and since then, many allergenic profilins have been identified in pollen, plant foods, and latex [6], thus indicating a high degree of cross-reactivity due to their common epitopes.

Some sequential and conformational profilin B-cell epitopes have been described using various approaches. The actin-binding site and the adjacent plant-specific pocket were found to comprise an immunogenic region responsible for cross-reactivity in the Arabidopsis profilin [7]. Two regions overlapping with the actin-binding site were identified as major cross-reactive epitopes, and a third site, consisting of residues 30-50, was found to be a likely cause of extensive cross-reactivity in birch profilin [8]. Several epitopes, which in fact cover most of the surface, have been identified in model structures of several profilins. Radauer et al [9] highlighted 3 main candidates: epitope 1, SWQTYVDDHQYQGL; epitope 7, PGAMVIQGEPGARGKPNE; and epitope 8, MKDEPQGVIEGEPGARKKE. Leitner et al [10] found that the circular peptide CAISGGYPVC inhibited IgE binding to mugwort pollen, birch pollen, and celery tuber profilin and speculated that this epitope might be an important epitope in plant profilins. In the case of Cuc m 2, the main watermelon allergen, the sequence S2W3A5Y6D9H10T11P12G113Q114 N116M117R121L122 [11] was described as the main epitope. This IgE-binding region was implicated in cross-reactivity with most plant profilins (eg, Phl p 12 and Bet v 2) owing to the high identity observed (Table 1). The identification of this sequence allows it to be used as a diagnostic marker for cross-reactivity mediated by the profilin family, as well as for future strategies in immunotherapy.

The Established Concept of Profilin

Prevalence

In order to establish the prevalence of profilin, it is extremely important to clarify whether the selected population is first chosen for its pollen allergy or plant food allergy. Profilin sensitization ratios across Europe can vary widely among pollen-allergic patients, especially primary sensitizers, from 5% in a Swedish birch pollen–allergic cohort [12] to 51% in a subset of a Mercurialis annua–allergic population in Spain [13]. This geographical variability and north-south gradient is due to the predominance of various pollens across Europe [14], which has been corroborated elsewhere [3].

Profilin sensitization is assumed to be always preceded by sensitization to a major allergen, although cross-sectional studies have failed to identify the primary sensitizer in most panallergen-sensitized patients because the vast majority are sensitized to 2 or more pollen sources [15]. Grass pollen has been reported to be one of the more robustly associated profilin sensitizers [16].

Figure. 3D structures of melon Cuc m2 (A), timothy grass pollen Phl p 12(B), birch pollen Bet v 2 (C), and human I profilins (D) [5]. Upper row: molecular surfaces in the upper orientation. Bottom row: views derived from the upper row after a clockwise 90° rotation about a vertical axis. Only the differences with respect to the sequence in A are labelled in mimotope sequences in B and C.
Barber et al [17] used a panel of 13 purified allergens to study 891 pollen-allergic patients from southern Spain, of whom 15% were sensitized to apple profilin (rMal d 4). More interestingly, a geographical variation according to seasonal grass pollen load was observed, revealing areas in which 50% of the population was sensitized to profilin. A logistic multivariate analysis showed that profilin sensitivity was associated with the major grass allergens Phl p 1 (OR, 3.16; 95%CI, 1.71-5.83) and Phl p 5 (OR, 6.19; 95%CI, 3.86-9.91). In a study with a similar design, but in 1329 patients from northern Spain, the same authors [18] detected that 18.8% of the population was profilin-sensitive and found that this sensitivity was again significantly associated with Phl p 5 (OR, 5; CI not provided). As a consequence, profilin seems to play a relevant role in areas where grass allergy is predominant. These areas can be identified using epidemiological studies mapping the sensitization clusters by region, such as that of Barber et al [19] in Spain.

The role of pollens other than grass in profilin sensitization can be understood with the olive pollen allergy model, as this pollen reaches maximum known exposure levels in some areas of Andalucia [17]. In the case of olive profilin (Ole e 2), results for prevalence and impact of sensitization are contradictory. Ole e 2 is usually acknowledged as a minor allergen [20] and was not found to be associated with Ole e 1 in one of the aforementioned studies [17]. Moreover, this lack of association has already been the subject of commentary by other authors [21]. However, in the study by Quiralte et al [22], 54% of 146 olive-allergic patients displayed sIgE to Ole e 2. The authors speculate that given the extremely heavy load of olive pollen in the area studied (Jaen, Spain, average 500 to 1000 grains/m3 with peaks of 10 000 grains/m3), the patients may have become sensitized to more olive allergens than in other places, thus underscoring the relevance of the area in the patients' molecular recognition patterns. One potential weakness of this study is the lack of information regarding the patients' sensitization to other pollen sources. This heavy pollen load was also used to explain relevant sensitization to other minor olive allergens, especially Ole e 7, which was found to be linked to an increase in the prevalence of asthma and the severity of allergic disease [17,23].

Table 1. Members of the Profilin Family Identified as Allergens

<table>
<thead>
<tr>
<th>Plant Species</th>
<th>Allergen Name</th>
<th>Id (%)</th>
<th>UNIPROT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cucumis melo (muskmelon)</td>
<td>Cuc m 2</td>
<td>100</td>
<td>Q5FX67</td>
</tr>
<tr>
<td>Actinidia delicosa (kiwi fruit)</td>
<td>Act d 9</td>
<td>68</td>
<td>FG438715</td>
</tr>
<tr>
<td>Ambrosia artemisiifolia (short ragweed)</td>
<td>Amb a 8</td>
<td>69</td>
<td>Q2KN24</td>
</tr>
<tr>
<td>Ananas comosus (pineapple)</td>
<td>Ana e 1</td>
<td>76</td>
<td>Q94JN2</td>
</tr>
<tr>
<td>Apium graveolens (celery)</td>
<td>Api g 4</td>
<td>78</td>
<td>Q9XF37</td>
</tr>
<tr>
<td>Arachis hypogaea (peanut)</td>
<td>Ara h 5</td>
<td>79</td>
<td>Q9SQ9</td>
</tr>
<tr>
<td>Artemisia vulgaris (mugwort)</td>
<td>Art v 4</td>
<td>69</td>
<td>Q8H2C9</td>
</tr>
<tr>
<td>Betula verrucosa (Betula pendula) (European white birch)</td>
<td>Bet v 2</td>
<td>74</td>
<td>P25816</td>
</tr>
<tr>
<td>Capsicum annum (bell pepper)</td>
<td>Cap a 2</td>
<td>85</td>
<td>Q93Y19</td>
</tr>
<tr>
<td>Chenopodium album (pigweed)</td>
<td>Che a 2</td>
<td>77</td>
<td>Q84V37</td>
</tr>
<tr>
<td>Citrus sinensis (sweet orange)</td>
<td>Cit s 2</td>
<td>76</td>
<td>P84177</td>
</tr>
<tr>
<td>Corylus avellana (hazel)</td>
<td>Cor a 2</td>
<td>83</td>
<td>Q9AXH5</td>
</tr>
<tr>
<td>Daucus carota (carrot)</td>
<td>Dau c 4</td>
<td>75</td>
<td>Q8SAE6</td>
</tr>
<tr>
<td>Glycine max (soybean)</td>
<td>Gly m 3</td>
<td>83</td>
<td>O65809</td>
</tr>
<tr>
<td>Helianthus annuus (sunflower)</td>
<td>Hel a 2</td>
<td>71</td>
<td>O81982</td>
</tr>
<tr>
<td>Hevea brasiliensis (para rubber tree [latex])</td>
<td>Hev b 8</td>
<td>82</td>
<td>O65812</td>
</tr>
<tr>
<td>Malus domestica (apple)</td>
<td>Mal d 4</td>
<td>77</td>
<td>Q9XF42</td>
</tr>
<tr>
<td>Olea europaea (olive)</td>
<td>Ole e 2</td>
<td>74</td>
<td>Q24169</td>
</tr>
<tr>
<td>Phleum pratense (timothy grass)</td>
<td>Phl p 1</td>
<td>76</td>
<td>P35079</td>
</tr>
<tr>
<td>Phoenix dactylifera (date palm)</td>
<td>Pho d 2</td>
<td>77</td>
<td>Q8L5D8</td>
</tr>
<tr>
<td>Prunus persica (peach)</td>
<td>Pru p 4</td>
<td>79</td>
<td>Q8GT40</td>
</tr>
<tr>
<td>Pyrus communis (pear)</td>
<td>Pyr c 4</td>
<td>77</td>
<td>Q9XF38</td>
</tr>
<tr>
<td>Salsola kali (Russian thistle)</td>
<td>Sal k 4</td>
<td>76</td>
<td>C6JWH0</td>
</tr>
<tr>
<td>Sinapis alba (yellow mustard)</td>
<td>Sin a 4</td>
<td>81</td>
<td>E6Y2M0</td>
</tr>
<tr>
<td>Solanum lycopersicum (tomato)</td>
<td>Sola l 1</td>
<td>85</td>
<td>Q93YG7</td>
</tr>
</tbody>
</table>

Sequence identity to Cuc m 2 is indicated.
On the other hand, if patients are selected based on plant food allergy as the main criterion, the geographical distribution of profilin sensitization displays a similar north-south gradient. In an interesting study by Fernandez-Rivas et al [24], component-resolved diagnosis (CRD) was used to assess area-dependent recognition patterns for Mal d 1 (major apple allergen, Bet v 1 homologue), Mal d 2 (thraumatin like protein), Mal d 3 (lipid transfer protein [LTP]), and Mal d 4 (profilin) in a group of 389 apple-allergic patients from 4 European countries (Austria, Italy, Netherlands, and Spain). Their results showed that apple allergy in individuals from the Netherlands, Austria and Italy was associated with Mal d 1 and milder symptoms, whereas in Spain, apple allergy was linked to Mal d 3 and severe manifestations. Both sensitization to and sIgE levels of profilin were higher in Spain and Italy (around 40% and 30% of patients, respectively) than in the Netherlands or Austria, where it was recognized in no more than 15% of the population. This study provides further evidence of the higher prevalence of profilin sensitization in southern countries than in northern countries, a trend that is also supported by Andersen et al [25] in their review of the panallergens involved in Rosaceae fruit allergy. After including 38 European studies with determination of several isolated allergens, the authors state that in western Mediterranean areas, sensitization to PR-10 is almost absent, with LTP being the first cause of Rosaceae fruit allergy, followed by profilin, which is also linked to non-Rosaceae fruits. In contrast, in northern and central Europe, Rosaceae fruit allergy is mostly due to class 2 fruit allergen and cross-reactivity to PR-10 (Bet v 1 homologs) with poorer profilin recognition.

**Diagnosis of Sensitization to Profilin**

Profilin allergy can be diagnosed either in vitro or, in the countries where purified profilin extract is available, in vivo. In vivo diagnosis with purified palm tree profilin, nPho d 2, at 50 μg/mL, has proven to have a high diagnostic efficiency [18,26-29]. For in vitro profilin diagnosis, a single profilin (either Bet v 2 or Phl p 12) is sufficient [26]. Variability in the recognition of profilin in in vitro diagnosis is more related to specific isoform selection and protein folding than to real recognition differences between the various allergenic sources [26]. A recent consensus document examines the use of molecular diagnosis in allergy in daily practice, including a chapter on profilin and its characteristics [30].

**The Role of Profilin in Respiratory Allergy**

Profilin has been accepted as a minor aeroallergen in most pollen sources [31], with little or no clinical impact and a prevalence below 50% in most cases. There are some exceptions, such as Che a 2, the Chenopodium album profilin, which was recognized by 55% (n=104) of a Spanish Chenopodium-allergic population [32]. However, its clinical impact was not fully addressed since the same group was also sensitized to Che a 3 (46%) and displayed bands for several other molecular weights in Western blots from a sample of 12 patients. Its role as a major allergen in this pollen was later supported in a population of 32 Chenopodium-allergic patients from Iran, where 81% displayed IgE to rChe a 2 [33]. Another example of profilin as a major allergen is that of Pho d 2, which triggered 56% and 64% of positive skin prick test (SPT) and ELISA results, respectively, in a population of 25 date palm-allergic patients [34]. As in the Chenopodium population mentioned above, the results of Western blots exhibited several other bands in addition to a 14.4-kDa band (supposedly profilin), thus compromising the real clinical impact of profilin.

Only 1 classic report considers the impact of profilin sensitization using a purified rBet v 2 extract for nasal challenge [35]. In a population of 24 tree and/or grass pollen-allergic patients, 10 showed sIgE to rBet v 2, and 8 also presented symptoms in the specific nasal provocation challenge with rBet v 2. Despite this hint of evidence, it is generally accepted that profilin is not a relevant respiratory allergen.

**The Role of Profilin in Food Allergy**

Allergy to profilin-containing foodstuffs is due to primary sensitization to profilin through inhalation and subsequent development of the so-called pollen-food syndrome (PFS) [36], which is based on a type II food allergy mechanism. Most syndromes involve weed pollen (eg, Ambrosia, Chenopodium, Artemisia) and grass and birch pollens (see Table 2). Although performing an extensive review of the literature in search of profilin-linked pollen-food syndromes is beyond the scope of this manuscript, it should be borne in mind that some of the classic references supporting such syndromes might only provide hints of an association and not proper evidence, either because there is no proper identification of the causative allergen or because the patients’ clinical background is missing.

The most frequent scenario is profilin recognition with little or no clinical relevance [37,38]. Given its lability in pepsin digestion [39] and thermal sensitivity [40], profilin triggers oral allergy syndrome (OAS), where symptoms involve itching of the lips, tongue, mouth, and throat, are self-limiting both in time and extension, and appear immediately after the intake of raw plant foods. Nevertheless, there are 2 reports of systemic reactions to lychee fruit [41] and zucchini [42], with profilin being the putative allergen that creates an exception to this rule.

Despite being considered a minor allergen, profilin is the major allergen of some plant foods, for example, melon (Cuc m 2) [43], orange [44], and soybean (Gly m 3) [45,46]. It can induce symptoms to virtually every plant food; however, allergy to melon, watermelon, citrus fruits, tomato, and banana has been reported to be a clinical marker of profilin hypersensitivity in a population of patients with OAS after ingestion of vegetables [38,47].

**The Role of Profilin in Latex Allergy**

To assess the role of Hev b 8 in latex allergy and associated syndromes, it is of utmost importance to clarify whether a patient became sensitized to latex in the first place, or if initial sensitization was due to other sources (pollen or plant foods). At the same time, it is useful to bear in mind that Hev b 8 is present in very low amounts or even absent in natural latex rubber gloves [48].

In primarily latex-sensitized individuals, recognition of Hev b 8 was seen to reach 40% when a purified recombinant form of rHev b 8 was used in a selected population of patients with spina bifida and latex allergy [49]. In a recent study by
ranging from 89.3% to 93.9% with Hel a 2 (sunflower profilin) similarity of its sequence with profilins from other sources, is most likely a cross-reactive phenomenon due to the high patients is not based upon a primary sensitization to latex, but sensitization to Hev b 8 found in pollen- and/or fruit-allergic generally, the role of latex profilin is questionable [56]. Generally, the patatin-like proteins (Hev b 7) [53] have been directly involved, Blanco et al [54]. Latex class I chitinases (Hev b 6) [55] and despite other allergens also being potentially involved, symptoms were first described as the latex-fruit syndrome by asthma caused by natural rubber analyzed with a panel of 12 latex allergens, Hev b 8 was only recognized in 4 patients (4.8%). Even though profilin was the only putative allergen in 2 of these patients, the authors still resist considering it to be clinically relevant. The presence of sIgE to Heb v 8 is usually a marker of nonrelevant sensitization. Using a 9–latex allergen platform, Schuller et al [51] detected monosensitization to a marker of nonrelevant sensitization. Using a 9–latex allergen platform, Schuller et al [51] detected monosensitization to (peanut profilin) and Pyr c 4 (pear profilin), respectively [57]. Garnier et al [58] reported 130 patients with positive sIgE to natural rubber latex, 97 of whom were latex-allergic. Among the 33 non–latex-allergic patients, 30 had food allergy, pollen allergy, or both, and 26 were monosensitized to rHev b 8. In contrast, in a subset of 46 latex-allergic patients without pollen allergy, only 1 displayed sIgE for rHev b 8, although he was food-allergic. This evidence reinforces the lack of impact of rHev b 8 positivity on latex allergy in fruit/pollen-allergic patients.

Table 2. Plant-Food Syndromes Involving Profilin

<table>
<thead>
<tr>
<th>Syndrome</th>
<th>Pollen</th>
<th>Plant Food</th>
</tr>
</thead>
<tbody>
<tr>
<td>USA: Prevalence of profilin sensitization, 15% [98]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ragweed-melon-banana [99]</td>
<td>Ragweed (Ambrosia)</td>
<td>Cucurbitaceae (melon), Musaceae (banana)</td>
</tr>
<tr>
<td>CENTRAL EUROPE: Prevalence of profilin sensitization, 15%-26% [100,101]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mugwort-birch-celery [41,102-106]</td>
<td>Birch (Betula), Mugwort (Artemisia)</td>
<td>Apiaceae (celery), lychee, carrot, anise, fennel, coriander and cumin</td>
</tr>
<tr>
<td>Birch-fruit [107]</td>
<td>Birch (Betula)</td>
<td>Banana, pineapple</td>
</tr>
<tr>
<td>Ragweed-melon-banana [42,108]</td>
<td>Ragweed (Ambrosia)</td>
<td>Cucurbitaceae (zucchini),</td>
</tr>
<tr>
<td>Musaceae (banana)</td>
<td>Compositae (Ambrosia, Artemisia)</td>
<td>Lychee</td>
</tr>
<tr>
<td>SOUTHERN EUROPE: Prevalence of profilin sensitization, 15%-50% [17]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goosefoot-fruit [109,110]</td>
<td>Chenopodium</td>
<td>Cucurbitaceae (melon), Musaceae (banana), Rosaceae (peach), Liliaceae (garlic)</td>
</tr>
<tr>
<td>Mugwort-spice [111]</td>
<td>Mugwort (Artemisia)</td>
<td>Liliaceae (garlic)</td>
</tr>
<tr>
<td>Mugwort-peach [112]</td>
<td>Mugwort (Artemisia)</td>
<td>Rosaceae (peach)</td>
</tr>
<tr>
<td>Ragweed-melon [111]</td>
<td>Grass, weeds, and trees</td>
<td>Cucurbitaceae (melon, zucchini)</td>
</tr>
<tr>
<td>Grass/Olive-Rosaceae and several fruits [17,18,27,38,113,114]</td>
<td>Grass and olive</td>
<td>Peach, Banana, Fig, Kiwi, Melon, Orange, Peach, Pineapple, Watermelon</td>
</tr>
<tr>
<td>Plane tree-fruit [115]</td>
<td>Plane tree (Platanus)</td>
<td>Rosaceae, other fruits, peanut, treenuts, and vegetables</td>
</tr>
</tbody>
</table>

*Syndromes are included in areas where they are most frequently described for didactic purposes, although some were also described in different areas, which are also included in the references. It should be taken into account that in most cases, other allergens such as Bet v 1, CCDs, or others, might play a relevant role, and it is not possible to clarify the culprit of clinical reactivity.

*Studies where profilin is suspected due to molecular weight, but without proper identification.

Vandenplas et al [50] including 82 patients with occupational asthma caused by natural rubber analyzed with a panel of 12 latex allergens, Hev b 8 was only recognized in 4 patients (4.8%). Even though profilin was the only putative allergen in 2 of these patients, the authors still resist considering it to be clinically relevant. The presence of sIgE to Heb v 8 is usually a marker of nonrelevant sensitization. Using a 9–latex allergen platform, Schuller et al [51] detected monosensitization to latex profilin in 2 out of 14 latex-allergic patients (14.2%) and in 19 out of 28 nonallergic, latex-sensitive patients (67.8%). Overall, Hev b 8 is not considered to have a clinical impact in latex allergy and, according to guidelines [52], patients sensitized to this allergen alone do not need a latex-free setting during surgical procedures.

Around 30% to 50% of latex-allergic patients have IgE-mediated symptoms to many plant foods [53], most frequently to avocado, banana, kiwi, chestnut, and papaya. These symptoms were first described as the latex-fruit syndrome by Blanco et al [54]. Latex class I chitinases (Hev b 6) [55] and patatin-like proteins (Hev b 7) [53] have been directly involved, and despite other allergens also being potentially involved, the role of latex profilin is questionable [56]. Generally, the sensitization to Hev b 8 found in pollen- and/or fruit-allergic patients is not based upon a primary sensitization to latex, but is most likely a cross-reactive phenomenon due to the high similarity of its sequence with profilins from other sources, ranging from 89.3% to 93.9% with Hel a 2 (sunflower profilin) and Ole e 2 (olive profilin) or 88.6% to 95.5% with Ara h 5 (peanut profilin) and Pyr c 4 (pear profilin), respectively [57]. Garnier et al [58] reported 130 patients with positive sIgE to natural rubber latex, 97 of whom were latex-allergic. Among the 33 non–latex-allergic patients, 30 had food allergy, pollen allergy, or both, and 26 were monosensitized to rHev b 8. In contrast, in a subset of 46 latex-allergic patients without pollen allergy, only 1 displayed sIgE for rHev b 8, although he was food-allergic. This evidence reinforces the lack of impact of rHev b 8 positivity on latex allergy in fruit/pollen-allergic patients.

The Shift in the Perception of Profilin

Previously, profilin was shown to be a prevalent panallergen that is seemingly unable to unleash remarkable food allergic reactions and does not induce respiratory symptoms or latex allergy. However, in recent years, several publications have raised significant doubts about some of these concepts, leading to the belief that profilin is an allergen that should no longer be overlooked.

Profilin, a Marker of Severity and Clinical Course

Large epidemiological studies analyzing molecular recognition in pollen from different areas of Spain have shown profilin to be a marker of disease severity and polysensitization

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in grass allergy [17-19]. In the aforementioned population of 146 olive-allergic patients, sensitization to Ole e 2 was statistically associated with asthma (OR, 2.2; 95%CI, 0.9-5.1; \( P = .04 \)), although the confidence interval includes the null effect [22]. Similarly, in a cohort of 1271 pollen-allergic children, of whom 296 (23%) were sensitized to profilin, sensitization to rPhl p 12 was statistically associated with longer disease duration and OAS to plant foods, but not with more severe disease [21].

Given the cross-sectional design of previous studies, stronger evidence of profilin as a maker of long-term allergic disease could only be provided by longitudinal studies, such as that of Hatzler et al [59], where a group of 820 newborns were followed until the age of 13 years. Serum sampling and clinical evaluation of all patients were performed regularly, and the authors found that in the 177 patients who finally developed seasonal allergic rhinitis to grass, profilin sensitization invariably appeared in the latter stages of the disease course and never as an early marker, thus supporting its role as an indicator of longer-term disease.

Taking an opposite approach, that is, from clinical behavior to molecular recognition, a subgroup of pediatric patients [21] were assessed for molecular characterization according to a predefined clinical profile. The authors selected 300 pollen-allergic children who reported OAS within 5 minutes of ingestion of pollen-related foods and were diagnosed with PFS [60]. IgE antibodies to PR-10 (rBet v 1), LTP (rPru p 3), and timothy grass profilin (rPhl p 12) were determined to classify the patients. A cluster analysis revealed 2 profilin-related clinical endotypes. One group comprised 63 profilin-monosensitized children whose main distinctive characteristics were as follows: sensitization to grass, plane, olive, and plantain pollen; OAS caused by peach, kiwi, banana, and fruits from the Cucurbitaceae family; and a high frequency of asthma. The other, a more interesting group, included 85 children, in whom more than 1 panallergen (38% profilin-sensitized) was detected. These patients recognized birch and grass pollen, experienced symptoms with all the plant foods studied, and had several comorbidities such as asthma, urticaria/angioedema, and atopic dermatitis. This study provides compelling evidence that profilin sensitization is itself associated with a more marked presence of asthma, and, when accompanied by other panallergens, with more severe allergic disease.

A Symptomatic Aeroallergen

The impact of profilin in respiratory allergy has traditionally been considered low or nonexistent [3], and recent clinical studies support this statement [61]. However, in recent years, isolated case reports suggest that profilin may be the culprit allergen in patients with pollinosis. Favré et al [62] report the case of a grass pollen–allergic patient who later developed symptoms during the birch pollen season with negative sIgE to rBet v 1, rBet v 4, and rBet v 6, but with positive sIgE to rBet v 2. The patients also had a positive nasal challenge result with nBet v 2, suggesting that birch pollen symptoms were produced by sensitization to Bet v 2 alone. Asero et al [63] describe the case of a 32-year-old woman with allergic rhinocconjunctivitis and positive SPT results to all whole-pollen and profilin extracts (nPho 2, purified natural date palm pollen profilin). Surprisingly, in both ImmunoCAP and ISAC, all major pollen allergens were only weakly recognized or not recognized at all, although strong positivity for profilin was detected (rPhl p 12: 12.6 kU/L), leading the authors to conclude that profilin was the most probable culprit allergen for the patient’s respiratory symptoms.

The evidence suggested in these scarce case reports is reinforced by a few very well-designed studies with in vivo and ex vivo provocation tests. Núñez et al [64] demonstrated that profilin can induce ocular symptoms by performing conjunctival challenges with nPho d 2 in 2 groups of pollen-allergic patients: one group comprised profilin-sensitized patients (n=17), and the other was a control group comprising individuals not sensitized to profilin (n=14). None of the control patients reacted, while 65% (11/17) of the profilin-sensitized patients had a positive response. Two dilutions were used (50 and 5 µg/mL), and most of the reacting patients needed the higher dose to produce positive test results (8/11). Ruiz et al [65] showed how profilin (nPho d 2) induced positive nasal and bronchial challenges, respectively, in 43% and 77% of a profilin-sensitized cohort (n=23), but not in 5 non–profilin-sensitized pollen-allergen controls, thus providing evidence that profilin can trigger nasal and bronchial symptoms in sensitized patients. A recent publication [66] demonstrated how stimulation with Bet v 2 and Phl p 12 induced dose-dependent basophil activation in 40 Bet v 2–sensitized birch-allergic patients.

The above-mentioned data support the notion that despite the misguided perception of clinical irrelevance, profilin acts as a clinically relevant aeroallergen. Moreover, given the ubiquity of this protein in pollens and plants, sensitized subjects might react clinically to multiple allergen sources, presenting perennial symptoms and, potentially, a more severe allergic phenotype.

A not so Mild Food Allergen

Profilin is thought to be a clinically irrelevant food allergen that mostly elicits mild symptoms, although exceptions have been reported [41,42]. No other cases of systemic profilin allergy had been reported until recently, when 9 out of a cohort of 26 grass pollen–sensitized adults from an area with a high grass pollen load reported systemic reactions after ingestion of plant foods [67]. In the study, only 18 individuals (8 with a previously reported systemic reaction) agreed to undergo a double-blind, placebo-controlled food challenge with nPho d 2 at a maximum cumulative dose of 822.2 µg, which was equivalent to the profilin in 283 g of melon. All 18 patients reacted in the challenge (median, 81.24 µg; range, 0.074-821.24), and systemic symptoms were elicited in 11 patients (61.1%), with adrenaline being used in 5 cases. The authors speculate that the very high levels of grass in the atmosphere during the pollen season (peaks of 2000 grains/m³ and sustained levels above 300 grains/m³) and the high degree of sensitization to grass allergens in the patients in this geographic area are critical determinants of their severe profilin reactivity phenotype. In 2 recent presentations delivered at the 2016 annual EAACI meeting in Vienna [68,69], the authors described extensive oral mucosa remodeling together with a 10-fold increase in effector cell sensitivity associated with severe
food profilin-mediated reactions. This is the first evidence that the oral mucosa can be an effective route for eliciting severe food reactions, with a potential impact on sublingual immunotherapy mechanisms and evolution from respiratory to food allergy.

As previously mentioned, melon and watermelon are the foods most frequently involved in profilin-induced food allergy [63], probably because the higher pH of melon compared with other fruits and vegetables [67] increases profilin stability and allows for a more efficient mucosal interaction. It has yet to be elucidated why patients who previously tolerated and ingested profilin daily develop such a severe allergic phenotype after presenting with severe grass respiratory allergy. The study of this particular population, which represents a unique clinical model, may provide an opportunity to understand the evolution of allergic disease and the increasingly widespread allergy pandemic [70] and to explore new biomarker strategies in allergy.

Impact of Sensitization to Profilin on the Selection of Allergen Immunotherapy

Profilin sensitization jeopardizes diagnosis and treatment in pollen-polysensitized patients. Moreno et al [71] reported a discrepancy in 56% of allergen immunotherapy (AIT) prescriptions when 1263 pollen-allergic patients were diagnosed based on SPT to whole extracts, compared with CRD based on the major grass and olive pollen allergens. Using a similar approach, Sastre et al [72] previously described a change in the selection of the composition of up to 54% of AIT prescriptions after CRD was applied in a group of 141 adults previously assessed only by SPT to whole extract, highlighting sensitization to profilin and polcalcin as one of the main confounding factors. Other authors have reported similar findings [73,74]. Nonetheless, these reports state how prescription of AIT might change after CRD and the assessment of major and minor allergens, although there are no data on whether application of CRD in selection of AIT improves efficacy or not.

Although CRD seems to be helpful in assessing the presence of major and cross-reactive allergens, it does not provide information on clinical relevance and might be of limited utility if more than 1 primary source allergen arises. In fact, it is not uncommon to detect profilin sensitization in patients who are primarily sensitized to 2 or more pollens [15]. It should also be borne in mind that sensitization to profilin has been associated with a higher prevalence of sensitization to "genuine" allergens from other pollen sources such as Phl p 1/5, Cup a 1, Art v 1, and Ole e 1 [21]; therefore, its presence might be considered a marker of advanced sensitization to the source, rather than just a mere finding to be ignored. Organ-specific challenges may be used to resolve this issue and assess clinical relevance [75,76] in polysensitized patients. However, the content of profilin in whole extracts might also obscure the real meaning of a positive test in organ-specific challenges, as has been suggested by some authors [64]. The amount of profilin in extracts used in organ-specific challenges is usually disregarded [76]; however, Ruiz et al [65] analyzed Pho d 2 content in 8 diagnostic pollen extracts (ALK-Abelló) and found that only grass preparations (Lolium and Phleum) seemed to have larger amounts of this protein (75 and 46.1 µg/vial of freeze-dried extract, respectively), whereas Betula, Chenopodium, Olea, Plantago, and Salsola profilin content remained far below 5 µg/vial. Compared with the major allergen content of each source per vial, the profilin percentage ranges from 0.8% for Lolium to 0.01% for Plantago. Profilin might be even less represented than shown by Ruiz et al, as seen in the results of Focke et al [77], who analyzed qualitative and quantitative allergen composition in 4 timothy grass pollen extracts and found that Phl p 12 could not be detected in any of them. In conclusion, it seems unlikely that the profilin content in challenge extracts might bias results, although better knowledge of this issue would be desirable.

Treating Profilin Allergy

The possibility of tailoring AIT at the molecular level has been speculated about for many years [78]. This approach is known as component-resolved immunotherapy. A recombinant form of Phl p 12 [79] and a mutant form of Cuc m 2 (melon profilin) [80] have been developed and proposed as candidates for profilin allergy immunotherapy. Although double-blind, placebo-controlled aeroallergen trials [81,82] with rPhl p 1, rPhl p 2, rPhl p 5a, rPhl p 5b, and rPhl p 6 showed that recombinant forms are effective and safe in respiratory allergy, recombinant forms have yet to be approved for use in humans. Despite these optimistic reports, Tripodi et al [83] describe 39 different recognition patterns for the 8 Phleum allergens studied in a population of 200 grass-allergic children, and even after ruling out polcalcin and profilin, their results still led to a significant degree of mismatch in the potential composition of AIT when compared with a previously used recombinant vaccine [81]. Considering both the minor impact of profilin in respiratory allergy and the low prevalence of sensitization compared with other allergens, it seems unlikely that an rPhl p 12 AIT product will be developed, although exploratory research is under way in the field [84].

Following a more viable approach, profilin as it is currently used in AIT products could be the best option for targeting this allergen. Asero et al [85] recently performed profilin-inhibition assays with the sera of 18 pollen-allergic profilin-sensitized patients and commercially available birch, grass, ragweed, and olive pollen AIT extracts. The authors concluded that given the high level of inhibition (80%-90%), these products contained large amounts of profilin and were potentially able to desensitize patients to this allergen. Nevertheless, several reasons discourage the use of current AIT products to specifically treat profilin allergy: AIT products are only standardized for major allergens [86], the differences in protein content are very wide [87], and the profilin content in allergen extracts is low [65] or undetectable [77]. Supporting these considerations, in a cohort of 33 grass-allergic patients (51% profilin-sensitized), the levels of IgG4 for Phl p 12 were undetectable after 16 weeks of grass subcutaneous immunotherapy (65 µg of Phl p 5 cumulative dose, Alutard SQ, Alk-Abello). Moreover, Phl p 1, Phl p 2, and Phl p 11 IgG4 levels were low, leading the authors to suggest that the vaccine content of all 4 allergens was also so low that it was unable to elicit induction of slgG4 [88].

Considering both scenarios, and in accordance with suggestions by other authors [21,59], the best way to treat
profilin sensitization/allergy may be to use preventive administration of regular AIT in early stages of pollen sensitization, as this would halt the expected progression towards higher sIgE levels and wider recognition of other allergens from the same source. Given that profilin sensitization is mainly associated with grass pollen, as evidenced by an increase in prevalence at higher intensities in the grass pollen gradient and the observation that grass extracts have the highest profilin content [65] once the primary sensitization to Phl p 1 and or Phl p 5 is confirmed, grass monotherapy is likely the best therapeutic option to treat profilin-sensitized patients in the absence of specific profilin-based therapy. Unfortunately, owing to the lack of specificity of whole extract–based diagnosis and the underuse of CRD [89], correct identification of profilin-positive grass monoallergenic patients is limited, and patients are therefore treated with less efficacious extracts in the best case or placed at risk of de novo sensitizations in the worst case.

In type II food allergy, it has been hypothesized that symptoms due to cross-reactivity with its homologs in plant foods will also be reduced by administering pollen AIT with the primary allergen. Interesting publications on birch pollen–allergic patients with vegetable allergy due to the cross-reactivity of PR-10 proteins (Bet v 1 homologs) show both beneficial effects [90,91] and no effect [92,93] in the associated food allergy despite a favorable response with respect to respiratory symptoms. The experience with profilin is far more limited, with only 2 reports of food allergy being successfully treated with AIT [94,95], suggesting that pollen AIT is unable or too underpowered to treat the secondary food allergy.

Another route that has been explored to minimize the impact of profilin allergy is the production of plant foods with reduced allergenicity [96], although these proposals have not yet led to any real-world implementation. To date, the best option for treatment of profilin-induced food allergy is that recently discussed by Nucera et al [97]. In their study, 7 patients withprofilin-induced PFS and OAS to a wide array of foods (median number of foods triggering symptoms, 9) were treated with an nPho 2 extract (50 µg/mL) following a sublingual protocol with incremental doses up to a maximum of approximately 75 µg of profilin per week. The duration of the treatment was 9 to 10 months. Treatment was very well tolerated, and in the exit double-blind, placebo-controlled food challenge with each of the offending foods, patients increased the number of vegetables they could eat from 23% to 92.9%. This new approach needs further optimization, and although profilin usually induces only mild symptoms, the high number of implicated foods produces a significant burden for patients and represents an important therapeutic target.

Conclusions

Profilin plays a relevant role as sensitizer and as a confounding factor in both diagnosis and treatment of patients with pollen and plant food allergy. Its relevance in latex-allergic individuals remains low or nonexistent according to several publications. In controlled settings, profilin has proven able to induce symptoms at all levels of the respiratory tract, although it still has to be elucidated whether it can induce respiratory symptoms in real-world exposure and to which extent it contributes to symptoms. The role of profilin in the bothersome OAS to several fruits has been acknowledged in pollen-allergic patients. In addition, profilin can trigger systemic reactions to plant foods in selected populations who routinely face seasons with heavy grass pollen loads. From a more holistic perspective, sensitization to profilin has been significantly linked to more severe presentation of allergic disease. Therefore, its presence should be taken seriously by allergists, who should begin to consider it more than a mere confounding factor in patient evaluations. Despite the aforementioned relevance, there is no solid therapeutic approach to treat profilin allergy. Moreover, currently available AIT products are most probably underpowered and food immunotherapy insufficiently explored. Prevention strategies could be the best option for patients who are likely to become sensitized to profilin if they are identified at early stages of their disease.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

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