REVIEWS

Practical Guidelines for Perioperative Hypersensitivity Reactions

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Abstract

Perioperative hypersensitivity reactions constitute a first-line problem for anesthesiologists and allergists. Therefore, hospitals should have a consensus protocol for the diagnosis and management of these reactions. However, this kind of protocol is not present in many hospitals, leading to problems with treatment, reporting of incidents, and subsequent etiological diagnosis. In this document, we present a systematic review of the available scientific evidence and provide general guidelines for the management of acute episodes and for referral of patients with perioperative hypersensitivity reactions to allergy units. Members of the Drug Allergy Committee of the Spanish Society of Allergy and Clinical Immunology (SEAIC) have created this document in collaboration with members of the Spanish Anesthesia Society (SEDAR). A practical algorithm is proposed for the etiologic diagnosis, and recommendations are provided for the management of hypersensitive patients.


Resumen

Las reacciones de hipersensibilidad perioperatorias constituyen un problema de primera línea para los anestesiólogos y alergólogos, por lo que es recomendable que los hospitales tengan un protocolo de consenso para el diagnóstico y el tratamiento de estas reacciones. Sin embargo, este tipo de protocolos no está presente en muchos hospitales, lo que conlleva problemas en el tratamiento, la comunicación de incidentes y el posterior diagnóstico etiológico. En este documento, se presenta una revisión sistemática de la evidencia científica disponible y se proporcionan pautas generales para el manejo de episodios agudos y para la derivación de pacientes con reacciones de hipersensibilidad perioperatoria a los Servicios de Alergología. Se propone un algoritmo práctico para el diagnóstico etiológico y se brindan recomendaciones para el manejo de pacientes con reacciones alérgicas perioperatorias.

1. Introduction

During the perioperative period, patients are exposed to multiple agents that can induce hypersensitivity reactions, with an estimated incidence of 1 per 10,000 anesthesia procedures [1,2,3]. However, prospective studies suggest that this is an underestimate and quote incidences of 1:3180 [4] and 1:1480 [5]. In a recent prospective Spanish study, the incidence of perioperative reactions was 1:381; of these, 48% were mild, involving only the skin, and 52% were anaphylaxis [6].

Owing to its low incidence, perioperative anaphylaxis is an unexpected and severe event. This hampers identification and early treatment and partially explains its high mortality (3-10% of cases) [1,7]. Perioperative hypersensitivity reactions constitute a first-line problem for anesthesiologists and allergists, and although it is advisable for hospitals to have a consensus protocol of action for diagnosis and treatment of these reactions [8], few actually do [9-12,13] and no protocols are specifically applicable to the Spanish population. Furthermore, there are no protocols of action for patients with a prior history of perioperative hypersensitivity reactions; these patients must be identified in pre-anesthesia consultations and referred to an allergy specialist for evaluation. This lack of specific protocols can imply a greater potential risk of re-exposure.

The management of perioperative hypersensitivity reactions is extremely complex and should be a combined effort between allergists and anesthesiologists [14,15] based on 2 well-differentiated areas [16,17]: (i) the acute phase of the reaction, which is the anesthesiologist’s responsibility [14]; (ii) later diagnosis to confirm the causal agent (if possible) which is the allergist’s responsibility. The aim of the present study was to develop clinical guidelines for the management of patients with a hypersensitivity reaction during the perioperative period and for the subsequent allergological diagnosis.

2. Methodology

A literature search was performed using key words agreed on by the authors. The search was performed using electronic databases (MEDLINE and PubMed), electronic libraries (Science Direct, OVID), and a database of systematic reviews (Cochrane Library). Publications were selected from between January 1985 and March 2016. The selection took into account the prevalence, pathogenesis, clinical manifestations, diagnosis, and treatment of perioperative hypersensitivity. The key terms used were perioperative anaphylaxis, perianesthetic anaphylaxis, and perioperative hypersensitivity reactions. In addition, the names of drugs commonly involved in perioperative reactions were searched for in combination with the terms skin tests, prick test, intradermal test, in vitro tests, and drug provocation tests. This search revealed 323 publications. Original research articles and systematic reviews were included; nonsystematic reviews, comments, and other types of article were excluded. We also included studies examining incidence, prevalence, natural history, clinical manifestations, pathogenesis, diagnosis, and treatment. Studies not addressing perianesthetic perioperative hypersensitivity were excluded. Following this review process by the expert panel, 195 publications were finally selected. Moreover, the expert panel evaluated the quality of the evidence and provided grades of recommendation according to the Scottish Intercollegiate Guidelines Network [18]. Wherever evidence was lacking, a consensus was reached among the experts.

3. Mechanisms of Perioperative Hypersensitivity

Although clinical presentation and early management are similar, perioperative hypersensitivity reactions may depend on 2 mechanisms: immunological mechanisms (allergic reactions) and nonimmunological mechanisms [19]. IgE-mediated immunological reactions represent 60% of all reactions and their severity can increase in a subsequent surgery [20-23]. These reactions undergo a sensitization phase, with activation of T_{H}2 and B lymphocytes and production of specific IgE antibodies that bind to high-affinity receptors of mast cells and basophils. In a second contact with the sensitizing agent and its binding to specific IgE, mediators such as histamine, tryptase, PG2, leukotrienes, thromboxane A2, platelet activating factor, chemokines, and cytokines such as tumor necrosis factor are released, leading to the development of the reaction [24]. It should be noted that in some cases, a reaction can occur upon first contact, which could be due to cross-reactivity with other substances to which the patient is sensitized. On rare occasions, immunological reactions may not be IgE-mediated, as reported for dextrans, which create immunocomplexes with IgG and activate the complement system; in such cases, these reactions are less severe [25].

Although the mechanisms are not well established, it is accepted that nonimmunological reactions are caused by direct stimulation (pharmacological or toxic) of mast cells and basophils, which induces their degranulation [26]; therefore, previous contact with the causative agent is not required [20]. These reactions are generally milder than immunologically mediated ones [7], except for a subgroup of patients, who are over-responders to the histamine released by neuromuscular blocking agents (NMBAs) [3,27,28].

4. Clinical Symptoms

The clinical manifestation of anaphylaxis due to anesthesia is similar to that of other forms of anaphylaxis, although it does present specific aspects [16,29]. Given that the patient is generally unconscious and covered by surgical drapes and cannot express what is happening, the prodromal symptoms (pruritus, dyspnea, or discomfort) may not be recognized. Instead, the reaction is often first recognized by the anesthesiologist, who notices nonspecific symptoms such as a drop in blood pressure and arterial saturation, difficulty in mechanical ventilation, severe arrhythmias, and cardiovascular collapse [1,30,31]. Consequently, some mild cases may recover spontaneously, meaning that the reaction goes unnoticed. Subsequently, re-exposure can lead to a more severe, potentially life-threatening reaction [4].

Reactions can occur at any time during anesthesia [32], although around 90% have been shown to occur suddenly during the induction phase after intravenous administration.
of the culprit agent (especially antibiotics, NMBAs, and hypnotic drugs) [32]. Sometimes, reactions can occur with a more delayed latency period, depending on several factors: (i) specific agents, such as dyes [33] and colloids; (ii) route of administration (cutaneous, mucosal, intraperitoneal, or subcutaneous), which delay absorption [34,35] of agents such as latex [9], chlorhexidine, or surgical glues [9,10,36]; (iii) some surgical procedures, such as gynecological procedures, owing to the release of latex particles in utero after the injection of oxytocin [31], and some orthopedic procedures, after the release of the tourniquet used in surgeries with ischemia [37].

Considering the organs involved, cutaneous symptoms, such as erythema, urticaria, and angioedema, are observed in 66%-70% of IgE-mediated reactions and in more than 90% of non–IgE-mediated reactions [38], with up to 10%-20% of cases not having any cutaneous symptoms [39,40]. Cardiovascular symptoms often include hypotension and tachycardia, which may progress rapidly to severe arrhythmia and cardiovascular collapse if they are not treated immediately [31,41-43]. These are the most frequent signs of severe anaphylaxis, and cardiovascular collapse or cardiorespiratory arrest may be the initial presentation symptoms [40,44]. In some cases, bradycardia might be the first sign of anaphylaxis; this is problematic, because the reaction could be confused with other, nonallergic anesthesia-related effects, especially if the patient is being treated with β-blockers. Other rare symptoms include acute coronary events associated with an immediate hypersensitivity reaction, such as Kounis syndrome, which is caused by the release of mediators from the cardiac mast cells [45,46]. Respiratory symptoms such as bronchospasm are less frequent and are observed in only around half of all patients [47], particularly those with a prior diagnosis of asthma [47]. The first sign may be an increase in pulmonary resistance or a decrease in oxygen saturation [48]. Other symptoms, such as alteration of coagulation [49], pulmonary edema, and rhabdomyolysis are very rare and are usually related to severe and prolonged anaphylactic shock. These symptoms are usually associated with the coexistence of cardiac disease, β-blockers, or angiotensin-converting enzyme inhibitors (ACE inhibitors). Given that many clinical symptoms of anaphylaxis reactions can be unspecific and may resemble the symptoms of other problems that can occur during anesthesia [9], it is critical to perform a differential diagnosis (Table 1).

The factors that have been identified as the main contributors to the clinical severity of anaphylaxis [11,40] include the following: (i) age, which is related to lower pulmonary capacity; (ii) prior diseases, especially cardiac or respiratory disease; (iii) systemic mastocytosis or elevated baseline tryptase [50]; (iv) current treatment, which may alter the patient’s response to catecholamine treatment, thereby potentially increasing mortality for medicines such as β-blockers, ACE inhibitors, angiotensin II receptor antagonists, monoaminoxidase inhibitors, tricyclic antidepressants, and serotonin uptake inhibitors; (v) form of administration, with the reaction occurring more rapidly when the drug is administered intravenously; (vi) initial presentation of the reaction (such as vascular collapse or cardiorespiratory arrest); (vii) speed of the clinical course (the faster the evolution, the more severe the reaction and the higher the risk of a fatal outcome); (viii) delay in administering epinephrine in the case of severe anaphylaxis [51,52].

Several classification systems have been proposed to evaluate the severity of reactions occurring during anesthesia [11].

### Table 1. Differential Diagnosis of Perioperative Anaphylaxis

<table>
<thead>
<tr>
<th>Pharmacological effect of anesthetic agents</th>
<th>Effect of local anesthetics or nerve block</th>
<th>Effect of the surgical technique</th>
<th>Effect of airway manipulation</th>
<th>Complications of surgery</th>
<th>Underlying disease</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hypotension, bradycardia</td>
<td>Sympathetic blockade, overdosage, accidental intravascular administration</td>
<td>Laparoscopy, eye surgery</td>
<td>Laryngospasm, bronchospasm</td>
<td>Pulmonary: Pulmonary edema, pulmonary embolism, amniotic fluid, fat or air embolism, pneumothorax</td>
<td>Systemic mastocytosis, hereditary angioedema, malignant hyperthermia, neuroleptic malignant syndrome, serotonin syndrome, carcinoid, or pheochromocytoma</td>
</tr>
</tbody>
</table>

5. Immediate Management of Perioperative Reactions

The anesthesiologist has a major role to play in both the prevention and the treatment of hypersensitivity reactions.

5.1. Preventive Measures

Prior to surgery, the anesthesiologist should evaluate the clinical history to identify previous history of allergy (especially allergy relating to medicines and latex), previous reactions during surgical procedures, and concomitant diseases and their treatments. Any of these factors can affect the development of an allergic reaction during anesthesia and will influence the effect of anesthesia on patient management.

If there is suspicion of latex allergy, the patient must be referred to an allergist for an allergology study prior to surgery. In the case of emergency surgery for a patient with suspected latex allergy, surgery must be performed in a latex-free environment. Similarly, in patients treated for a suspected drug allergy in the emergency department, the suspect drugs should...
be avoided. Locoregional anesthesia is preferred in these cases. Premedication with corticosteroids and antihistamines should be considered, especially if there is a suspicion of a non-immune-mediated reaction.

5.2. Management of Reactions

In the event of a reaction, a step-by-step process must be followed:

5.2.1. Recognizing the allergic reaction

The anesthesiologist must evaluate the patient’s signs and decide whether they are indicative of anaphylaxis by performing a differential diagnosis (Table 1), establishing the severity of the reaction, and identifying possible culprit agents.

5.2.2. Treatment of the reaction

The reaction must be treated immediately, since this will influence the patient’s prognosis, especially in severe reactions. Treatment will include general and specific pharmacological treatment depending on the severity of the reaction (Figure 1). The agents used in the treatment of the reaction can be classified as first- and second-line treatments:

(a) First-line treatment

**Epinephrine.** Anaphylactic reactions involve alterations of vascular permeability, which implies that up to 35%-50%

<table>
<thead>
<tr>
<th>Grade</th>
<th>Specific Management</th>
<th>Second Line Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Dexamethasone 5 mg</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Follow the progress of the reactions</td>
<td>1. No response to adrenaline:</td>
</tr>
<tr>
<td></td>
<td></td>
<td>a. Noradrenaline: 0.05-0.1 µg/kg/min</td>
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<tr>
<td></td>
<td></td>
<td>b. Glucagon 1-2 mg iv each 5 min or perfusion 5-15 µg/min</td>
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<tr>
<td></td>
<td></td>
<td>c. Vasopressin bolus 2-10 IU, repeat or perfusion 0.2-0.4 IU/min</td>
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<tr>
<td></td>
<td></td>
<td>2. Bronchospasm not responding to adrenaline or as a unique symptoms: Salbutamol</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3. Bradycardia after cardiopulmonary resuscitation: Atropine</td>
</tr>
</tbody>
</table>

**Figure 1.** Treatment of perioperative anaphylaxis. ABC indicates airway, breathing, circulation; PACU, postanesthesia care unit; ICU, intensive care unit.
of intravascular volume can migrate to the interstitial space in 10 minutes. Epinephrine is the drug of choice for treatment of anaphylaxis, and delay in its administration negatively influences the prognosis of severe reactions. There is no contraindication for the use of epinephrine during a reaction, although dosage should be adjusted based on severity in order to avoid severe adverse effects, especially in patients with heart disease.

**Vasoactive drugs.** In cases where the patient is taking β-blockers or has heart disease, other vasoactive agents can be given. Norepinephrine, ephedrine, methoxamine, phenylephrine, and dopamine can be administered as an intravenous bolus or continuous infusion [53].

**Glucagon.** This drug can be also used as a rescue medication in patients who normally receive treatments with β-blockers and who may not respond to epinephrine. As with norepinephrine vasoactive drugs, patients with heart disease are advised not to take epinephrine [40,54].

**Vasopressin.** The use of vasopressin in anaphylactic shock is accepted, since it may be consumed during the reaction and no response to vasopressor drugs is observed.

**Methylene blue.** This drug can be useful owing to its capacity to interfere with the action of nitric oxide in the smooth muscle of vascular walls. It should be administered in combination with epinephrine [55-57].

(b) **Second-line treatment**

**Antihistamines and corticosteroids.** These drugs should not replace first-line drugs such as epinephrine for severe reactions, although they can be used in mild reactions (grade I). Corticosteroids are not indicated in the acute phase of the reaction, but may be used to avoid delayed symptoms. Systematic reviews suggest that corticosteroids are not useful for the treatment of anaphylaxis [57,58].

**Salbutamol.** Salbutamol is indicated in patients who present bronchospasm as a main symptom or if this symptom does not respond to epinephrine.

**Atropine.** Atropine is restricted to cases of severe bradycardia refractory to epinephrine and/or fluid therapy and in patients treated with β-blockers, as they can induce cardiac arrest in the early phases of anaphylaxis.

**Other treatments.** Sugammadex has been proposed as a useful treatment for anaphylaxis symptoms caused by rocuronium [59,60]. However, in a recently published case series, sugammadex did not modify the course of the reaction [61]. Magnesium sulphate may be useful in cases of bronchospasm that are refractory to other treatments [62].

5.2.3. Actions after treatment

- **Obtaining biological samples for diagnosis.** Blood should be taken during this stage in order to measure serum tryptase, an indicator of mast cell/basophil degranulation [63]. Other markers such as histamine and methyl-histamine in urine have been used, although these are not recommended in routine practice owing to difficulties in measurement and low sensitivity [64].

- **Discharge.** In the case of mild reactions (grade I), the patient can be discharged after symptoms resolve. In severe reactions, the patient should be observed for at least 24 hours in a postsurgical recovery unit, in the intensive care unit, or in a regular hospitalization area. Extreme caution must be taken with patients who present severe symptoms and/or do not respond to treatment and/or experience involvement and/or severe comorbidities, or who have poor access to emergency treatment after discharge.

- **Informing the patient.** Before the patient is discharged, the anesthesiologist should give him/her a clinical report containing all relevant information about the reaction, including its severity, any treatment administered, and possible causative agents. The patient should be warned that, if anesthesia is needed again prior to the allergology work-up, it should only be for emergency procedures. In addition, the patient must inform the hospital and present the clinical report.

- **Referral for allergy evaluation.** With respect to a future diagnosis, the anesthesiologist must contact the allergist so that all the information about the event can be properly recorded.

In order to standardize the diagnosis and treatment of anaphylactic reactions, it is advisable to have ready-to-use kits in all areas of anesthesiology departments [8,15,41] including the following: (i) Simple anaphylaxis treatment algorithms, with the dosage of different drugs, route of administration, especially epinephrine, and rescue medication in case of severe/refractory anaphylaxis (Figure 1). (ii) Instructions to obtain biological samples from the patient, including laboratory orders, sample tubes, and instructions on where the samples must be sent (Figure 2). This is crucial for establishing a chronological sequence of the events leading to the reaction. (iii) A standardized data sheet that should be filled in after the reaction has been treated and sent to the relevant allergy department. This sheet should include details of the drugs administered and the temporal sequence of symptom onset. Any treatments administered to resolve the reaction should also be described (Figure 2) [15].

### 6. Allergy Evaluation

The diagnosis of perioperative hypersensitivity reactions is based on the combination of the clinical history, in vitro determinations performed during the acute phase of the reaction, and tests performed once the reaction has resolved. These include skin tests, in vitro tests, and, eventually, drug provocation testing. The diagnostic algorithm is shown in Figure 3.

#### 6.1. Clinical History

The clinical history is the first step for establishing a diagnosis, although all the information comes from the anesthesiologist’s report (grade of recommendation, D) [9,12,65]. It is important to evaluate various aspects, as follows: clinical signs and symptoms of the reaction; grade of severity [11]; drugs administered for treating the reaction and the time needed to resolve the reaction; personal history of allergy, including atopy and allergy to other drugs; risk factors, such as age; underlying diseases and treatments, such as β-blockers and ACE inhibitors; and all possible drugs and
agents used during the perioperative period, alongside details on their temporal sequence of administration. The agents most frequently involved include the following:

6.1.1. Drugs

Antibiotics. Antibiotics are usually administered before the induction phase [12] and constitute the most common cause of perioperative anaphylaxis in Spain [5]. It would be useful to separate the administration of prophylactic antibiotic therapy and anesthetic induction in order to identify and treat the reaction as early as possible. ß-Lactams, especially cephalosporins [5], are responsible for 70% of reactions due to antibiotics [20], with cefazolin being the most frequent causal agent in Spain [66]. Vancomycin is the second most common antibiotic involved, although reactions are almost always due to a nonimmunological mechanism [67]. Quinolones are the third most important group, with a high incidence [68]. Finally, other potentially important antibiotics include gentamicin, metronidazole, and tobramycin, which are mainly used in abdominal surgery [69].

NMBAs. In some studies, NMBAs are the agents most frequently involved in reactions [20,70,71]. Diagnosis is complex, as reactions can sometimes appear in patients receiving these drugs for the first time. This could be explained by the existence of cross-reactivity with other substances.
containing tertiary or quaternary ammonium groups such as cosmetics, foods, industrial material, and disinfectants [72]. Although this hypothesis has not been proven, it was recently shown that contact with quaternary ammonium in hairdressing students is associated with an increase in the incidence of IgE antibodies against NMBAs [22]. It was also shown that the use of pholcodine, which is present in some cough medicines, is related to an increase in the incidence of NMBA allergy, thus possibly explaining differences in incidence between countries, as the consumption of such medicines is variable [37]. Finally, diagnosis is complicated by the fact that all NMBAs are histamine-releasing drugs, especially benzylisoquinoline derivatives (d-tubocurarine, atracurium, and mivacurium) [26].

Sugammadex. Sugammadex is a modified gamma cyclodextrin that acts as a blocking agent of the aminosteroid NMBAs, especially rocuronium. It can induce IgE-mediated reactions [73-76].

Hypnotics. These include 2 groups of chemically unrelated drugs (barbiturates and nonbarbiturates). Drugs from the barbiturate group are used infrequently nowadays. Thiopental is the most highly consumed, and although IgE-mediated reactions have been reported, most are induced by nonspecific histamine release [77].

The nonbarbiturate group includes propofol, ketamine, etomidate, benzodiazepines, and inhaled anesthetics. Propofol is the cause of 2.3%-2.6% of perioperative anaphylactic reactions [20,78], most of which are IgE-mediated; the antigenic determinant is the 2-isopropyl group of the molecule (2,3 diisopropylphenol). Soybean oil and egg-derived lecithin are both used during its formulation, and although it has been suggested that patients with severe anaphylaxis to egg and/or soy should undergo an allergology work-up or use alternative treatment [79], these drugs seem to be safe in most patients. One study of 99 patients with positive specific IgE to egg, soya, or peanut (only 44% had immediate clinical symptoms) found that none had an allergic reaction following exposure to propofol [78]. In a Spanish study of 52 adult patients with eosinophilic esophagitis sensitized to egg, soy, or peanut, none of the patients who received propofol before an endoscopy procedure reacted to the drug [80]. In fact, according to our literature search, no reactions to propofol have been documented for soy- or egg-allergic patients. Moreover, prick testing with soya oil and propofol has always yielded negative results, indicating that there are no reasons to contraindicate its use [79,80].

Hypersensitivity reactions to benzodiazepines are extremely rare; midazolam was the most frequent etiological agent in the few cases that have been reported [7,20]. Reactions to etomidate and ketamine are also rare; in fact etomidate is considered one of the safest drugs in anesthesia in terms of allergic reactions [7,81].

Opioids. Reactions are rare, and although there have been some IgE-mediated reactions, most are due to nonspecific histamine release [82-85]. Semisynthetic opioids such as...
fentanyl, alfentanil, remifentanil, and sufentanil do not cause histamine release by themselves. Morphine derivatives are chemically different from phenylpiperidines, and there is no cross-reactivity between them [82-86]. While infrequent, allergic reactions to opioids are an important problem because these drugs are essential for anesthesia and it is difficult to find a safe alternative.

**Nonsteroidal anti-inflammatory drugs (NSAIDs).** NSAIDs are responsible for a high number of perioperative reactions [5,20]. In general, nonimmunological reactions due to COX-1 inhibition are more common [29]. In some cases, the underlying mechanism can be immunological, most often due to pyrazolones, followed by diclofenac, although again, specific studies relating to incidence in perioperative reactions are needed [87].

**Local anesthetics.** Although widely used, local anesthetics rarely induce adverse effects related to overdose or accidental intravascular administration [88]. In exceptional cases, they induce hypersensitivity reactions, which are mainly associated with the amide group [39,89].

**Other drugs.** There have been anecdotal reports of cases of anaphylaxis induced by heparins [39], tranexamic acid [90], uterotonic drugs [91,92], atropine [93], and neostigmine [93].

### 6.1.2. High-molecular-weight agents

**Colloids.** These can cause up to 4% of perioperative anaphylactic reactions, usually appearing 20-30 minutes after initiation of the infusion [1]. Gelatins are responsible for most reactions (95%), followed by dextran, and although both can induce nonspecific histamine release, IgE-mediated reactions due to gelatins and IgG-mediated reactions due to complement activation by dextran have been reported [39,94]. Gelatins can be a cause of anaphylaxis in patients sensitized to the carbohydrate epitope α-gal [95]. The incidence of reactions is much lower for albumin and exceptional for hydroxyethyl starch [96,97].

**Protamine.** Hypersensitivity reactions mediated by IgG, IgE, complement activation, and nonspecific histamine release have been reported [17,98]. Patients who have been previously exposed to this drug are at a higher risk of developing a reaction, regardless of whether the drug was given for blocking the effect of heparin or combined with insulin. Protamine is derived from fish sperm; however there is no increased risk for patients undergoing vasectomy or those allergic to fish [58]. It is important to be aware that rapid protamine infusion can induce hypotension.

**Aprotinin.** Aprotinin is a bovine-derived protease inhibitor used via the parenteral route as a fibrinolysis inhibitor or topically as a surgical glue [17]. The incidence of reactions in cardiac surgery is 0.5%, although this can increase to 2.5%-2.8% in patients who have previously undergone multiple surgeries [99,100], especially during the previous 6 months [31]. Perioperative anaphylaxis has also occurred upon the first parenteral administration in patients who had previously received topical aprotinin [17].

**Hyaluronidase.** Hyaluronidase is a bovine or ovine enzyme that degrades hyaluronic acid and can be used as a drug or fluid adjuvant. Both immediate reactions [101] and delayed reactions [102] have been reported during ocular surgery and epidural injection.

### 6.1.3. Antiseptics and sterilizers

**Antiseptics.** There are significant geographical differences with respect to the incidence of chlorhexidine-induced perioperative anaphylaxis. Reactions have been reported to be frequent in the UK and Scandinavia [71,103,104], with chlorhexidine accounting for 8.7%-9.6% of cases of perioperative anaphylaxis; however, such reactions were found to be relatively rare in France [22], perhaps because of the limited use of chlorhexidine in the operating room. The mechanism is IgE-mediated, and although reactions usually appear 20-30 minutes after administration, onset and severity depend on the administration route, with cutaneous application rarely causing severe reactions [36]. Nevertheless, when it is applied to mucous membranes (urinary catheters or oral cavity washes) or by parenteral routes (venous catheters or surgical meshes), the onset of clinical symptoms can be faster and more severe [36,104,105]. The appearance of anaphylaxis induced by povidone is unusual, and it is noteworthy that no cross-reactivity with iodinated contrast media has been reported [12,70].

**Sterilizers.** Ethylene oxide is a gas used for sterilizing multiple medical devices. Reactions have mainly been described in dialedyzed patients and in those with spina bifida and are extremely rare during perioperative reactions [1]. Administration of anesthesia without ethylene oxide is a challenge for the anesthesiologist, because in some cases it cannot be substituted. The case is similar for some intraarterial catheters, pump infusion systems, and intratracheal tubes.

### 6.1.4. Dyes

Isosulfan blue, its isomer patent blue V, and methylene blue are used for mapping sentinel lymph nodes in patients with breast cancer or melanoma. They are all capable of inducing hypersensitivity reactions, and the incidence of such reactions has been on the increase in the last decade owing to a rise in their intraoperative use [106]. Isosulfan blue and patent blue V can induce anaphylaxis in up to 1% of individuals who receive it, with reactions usually appearing 10-30 minutes after administration [107]. The typical reaction involves the appearance of blue wheals [108], although severe reactions with shock can appear in some cases. In most cases, there has been no previous medical exposure to these dyes, and it is thought that sensitization occurs through exposure to cosmetics or other objects. Anaphylaxis induced by methylene blue is less frequent, and this agent can be an alternative in patients sensitized to isosulfan blue [107], although cross-reactivity has been described in a few cases [109].

### 6.1.1. Latex

A decrease in the number of reactions to latex has been reported in several countries as a result of the reduction in latex exposure and sensitization by decreasing the protein content and stopping the use of powdered gloves [110,111]. The absorption of latex allergens usually occurs through the skin and mucous membranes, and although reactions can appear at any time during surgery, onset of symptoms is usually delayed. Abdominal, gynecological, and orthopedic surgery are usually
associated with an increased risk, and incidence is likely to be higher in the case of atopic patients, health system personnel, workers exposed to latex, patients undergoing multiple surgeries, women undergoing in vitro fertilization, children with urogenital malformation or spina bifida, and those with a history of perioperative anaphylaxis [112,113].

6.2. Skin Tests

Skin tests are the initial diagnostic approach. Interpretation of the results depends on clinical symptoms (grade of recommendation, B). In most cases, the skin prick test will be followed by an intradermal test. A skin prick test is considered positive when the mean wheal diameter is larger than 3 mm and surrounded by erythema and when the saline control is negative [87]. An intradermal test is considered positive when there is an increase greater than 3 mm of the initial wheal made by injection of the drug [87] (grade of recommendation, C). Skin testing should be performed within 4-6 weeks after the reaction. Sensitivity decreases over time; this decrease is faster for β-lactams than for NMBAs (grade of recommendation, D) [114,115].

The optimal concentration for skin tests is not clearly defined in all cases for drugs that can induce nonspecific histamine release [116,117]. Therefore, false-positive results can occur with drugs such as NMBAs (mivacurium, atracurium, cisatracurium, and succinylcholine), thiopental, opioids, and some antibiotics such as vancomycin and quinolones. A detailed investigation to determine the maximal nonreactive concentration for NMBAs in healthy individuals has been performed [118]. The concentrations recommended as nonirritant are shown in Table 2 [31,119].

Skin testing with NMBAs is considered to be highly sensitive (>95%) and specific (96-98%) and therefore mandatory for diagnosis and evaluating cross-reactivity, although this observation needs to be confirmed in other populations (grade of recommendation, C) [120]. Cross-reactivity among NMBAs has been reported, thus making it necessary to test other NMBAs when looking for a safe alternative (grade of recommendation, C) [9,12,121]. In addition, skin prick tests with latex show high sensitivity and specificity (grade of recommendation, B) (93% and 100%, respectively) [122]. Chlorhexidine is also highly sensitive and specific; 95% and 97%, respectively, for skin prick tests and 68% to 100% for intradermal tests (grade of recommendation, B) [103].

Skin testing can also be useful for diagnosing allergic reactions to antibiotics. However, except for β-lactams, in which sensitivity has been estimated to be 70% [123], sensitivity and specificity are not well established (grade of recommendation, D) [119]. In the case of NSAIDs, skin testing is only recommended for reactions to pyrazolones (grade of recommendation, C) [87,124]. Skin testing can be used to diagnose hypersensitivity reactions to hypnotic drugs [125], sugammadex [126], local anesthetics [89], and dyes [127] (grade of recommendation, C). The value of skin testing with opioids such as morphine for detection of perioperative reactions is unclear owing to their capacity to induce nonspecific histamine release (grade of recommendation, C) [116]. Given the mechanisms of reactions to dextrans (immune complex–mediated or complement activation), the value of skin tests is not established (grade of recommendation, D) [12].

There is some controversy over when to perform skin tests and which agents to use. The allergist has a key role here. Various scenarios can occur in clinical practice. First, patients have no previous clinical history of perioperative allergic reaction. In these cases there is no indication to perform skin tests (grade of recommendation, B) [12,118,128]. Second, patients may have a previous history of reactions, with detailed information on reaction kinetics and drugs and agents administered. In these cases, skin testing is mandatory for all the agents administered plus latex [129] and chlorhexidine.
However, since the measurement of tryptase also accounts for a high frequency of false negatives and a low predictive value (54%), a normal tryptase value does not rule out real anaphylaxis [93]. Although tryptase is more often increased than histamine, the former suggests an IgE-mediated reaction [131]. Furthermore, tryptase can be determined in deceased patients [11,30,136].

6.3. In Vitro Tests

In vitro tests can be performed during the acute phase of the reaction or once it has subsided. The former help to understand the mechanisms involved in and the latter to identify the culprit agent (grade of recommendation, C).

6.3.1. Acute phase of the reaction

In this case, tests are mainly based on the determination of histamine and tryptase. Determination of plasma histamine. Levels of histamine usually increase in the first 5-10 minutes after symptom onset; their half-life is 15-20 minutes, which is why blood samples need to be taken during the first 15-30 minutes of the reaction [11,132]. It is important to maintain the tube at 4ºC until processing to avoid nonspecific release due to cellular lysis. This determination is not sufficiently standardized for diagnostic use (grade of recommendation, C).

Determination of serum tryptase. Tryptase stays in the blood for hours, with maximum levels appearing after 1-2 hours and remaining high for a further 4-6 hours. This means that blood samples can be obtained between 30 minutes and 6 hours after the reaction. Moreover, the samples are less sensitive to environmental conditions, as determination is performed in serum. Various cut-offs have been considered, ranging from 8.23 μg/L to 11.4 μg/L [94-96], although the best criterion is a 2-fold or 2×1.2× increase above baseline levels (grade of recommendation, B) [63,133]. Tryptase levels are especially high in more severe reactions [134,135]. However, since the measurement of tryptase also accounts for a high frequency of false negatives and a low predictive value (54%), a normal tryptase value does not rule out real anaphylaxis [93]. Although tryptase is more often increased than histamine, the former suggests an IgE-mediated reaction [131]. Furthermore, tryptase can be determined in deceased patients [11,30,136].

6.3.2. Resolution phase

In vitro methods are further described in Table 4. Specific IgE determination. This method is useful when combined with skin test results, but not in isolation (grade of recommendation, B). The most widely available method is ImmunoCAP (Thermo Fisher), which can be used with latex, suxamethonium, morphine, pholcodine, bovine gelatine, protamine, chlorhexidine, thiopental, ethylene oxide, and some antibiotics. It is also possible to determine specific IgE to quaternary ammonium using the same method, and this can be used as a marker of sensitization to NMBAs and opioids. As this determination is positive in 3%-10% of nonallergic patients, results should be interpreted with caution [137]. Basophil activation test (BAT). BAT is useful when confirming skin test results for diagnosing or assessing cross-reactivity [138] (grade of recommendation, B). Some authors have obtained promising results in reactions induced by muscle relaxants; consequently, the test is recommended for routine analysis [139]. Moreover, some authors consider that performing the BAT with muscle relaxants is useful for diagnosis, even in patients with negative skin test results, and that it can be used for the identification of safe alternatives [137,139,140]. However, identification of safe alternatives based on BAT only should be regarded with caution, taking into account that only small series with BAT have been reported. Other determinations. Histamine and sulphidoleukotriene release assays have limited utility (grade of recommendation, C) [141,142].

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Table 3. Agents and Drugs Panel to Use in Skin Testing in Those Patients Where the Possible Culprits Are not Identified by the Clinical History

<table>
<thead>
<tr>
<th>Muscle relaxants</th>
<th>Hypnotics</th>
<th>Opioids</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atracurium</td>
<td>Propofol</td>
<td>Alfentanil</td>
</tr>
<tr>
<td>Cisatracurium</td>
<td>Etomidate</td>
<td>Fentanyl</td>
</tr>
<tr>
<td>Mivacurium</td>
<td>Ketamine</td>
<td>Remifentanil</td>
</tr>
<tr>
<td>Pancuronium</td>
<td>Thiopental</td>
<td>Sufentanil</td>
</tr>
<tr>
<td>Rocuronium</td>
<td>Midazolam</td>
<td>Morphine</td>
</tr>
<tr>
<td>Vecuronium</td>
<td></td>
<td>Chlorhexidine</td>
</tr>
<tr>
<td>Suxamethonium</td>
<td></td>
<td>Latex</td>
</tr>
</tbody>
</table>

Table 4. Available In Vitro Methods for Identifying the Culprit Agent Involved in Perioperative Reactions

<table>
<thead>
<tr>
<th>Drug</th>
<th>ImmunoCAP</th>
<th>BAT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Barbiturates</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Propofol</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Local anesthetics</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Neuromuscular blocking agents</td>
<td>X</td>
<td></td>
</tr>
<tr>
<td>Latex</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Opioids</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Atropine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protamine</td>
<td>X</td>
<td></td>
</tr>
<tr>
<td>Gelatins</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Chlorhexidine</td>
<td>X</td>
<td></td>
</tr>
<tr>
<td>Ethylene Oxide</td>
<td></td>
<td>X</td>
</tr>
</tbody>
</table>

Abbreviation: BAT, basophil activation test.

Available: suxamethonium.

Available: pholcodine and morphine.
6.4. Drug Provocation Tests

Drug provocation tests are considered the gold standard for diagnosing drug hypersensitivity (grade of recommendation, C) and consist of the administration of increasing doses of the drug at 30 minute intervals in a single-blinded and placebo-controlled manner until the therapeutic dose is reached or a reaction occurs [143,144]. Many of the drugs administered during the perioperative period, such as antibiotics and NSAIDs, are studied following regular procedures and are beyond the scope of this review. However, DPT with perioperative drugs has several added limitations, and there is no consensus procedure (grade of recommendation, C) [10,145,146]. The goal is to reach a total dose of the drug needed for induction of anesthesia (propofol, etomidate, ketamine) or to be administered during the anesthetic procedure (opioids, midazolam). This dose is not standard for all patients and depends on body weight and underlying diseases (Table 5) [147,148].

Although some of these agents can be administered in allergology units, many require coordination with the anesthesiology unit. Cardiovascular and respiratory monitoring is mandatory.

- Drugs administered in the allergy unit. It is generally preferable to use the oral and subcutaneous routes (grade of recommendation, D), with close monitoring of the patient. Every attempt should be made to avoid high-risk patients such as older patients or patients with comorbidities; for example, drug exposure might provoke reactions that are hard to control [143]. Among the opioids, morphine, pethidine, and fentanyl can generally be administered without major adverse effects, and if they do appear, they can be reversed with naloxone at an initial dose of 0.4 mg and repeated every 2-3 minutes until a response is obtained or a maximum dose of 10 mg is achieved [149]. Benzodiazepine is quite safe, and its sedative effects can be reversed with flumazenil, starting with a dose of 0.2 mg, and if necessary, repeating the dose of 0.2 mg each minute until a maximum dose of 1 mg is reached [149].

- Drugs administered in the surgery area. This approach is only indicated when there are no alternatives to administering anesthesia. A risk-benefit analysis should be performed, and administration should be intravenous (grade of recommendation, D) [3,10]. It should be performed in a setting with adequate cardiovascular and respiratory monitoring and in the presence of well-trained personnel to manage cardiovascular events (hypotension, bradycardia), respiratory events (depression), and hypersensitivity reactions (grade of recommendation, C). The most adequate setting is the operating room or the postanesthesia unit. The necessary material and personnel are as follows: (i) an anesthetist and an allergist throughout the DPT procedure and recovery; (ii) nurses trained in the management of airway and cardiovascular events; (iii) individualized monitoring (ECG, noninvasive blood pressure, pulse oximetry, and respiratory rate); (iv) resuscitation trolley and airway handling material; (v) a ventilator or anesthesia machine, especially if the DPT is performed with muscle relaxants or in high-risk patients (Physical Table 5. Doses Administered in the Drug Provocation Test

<table>
<thead>
<tr>
<th>Drug</th>
<th>Setting</th>
<th>Route</th>
<th>Maximum Dose</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Opioids</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Morphine</td>
<td>AU</td>
<td>Oral</td>
<td>5 mg; 10 mg in &lt;65 years</td>
</tr>
<tr>
<td>Pethidine</td>
<td>SC</td>
<td></td>
<td>25 mg</td>
</tr>
<tr>
<td>Fentanyl</td>
<td>SC</td>
<td></td>
<td>50 µg</td>
</tr>
<tr>
<td><strong>Benzodiazepines</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Midazolam</td>
<td>AU</td>
<td>Oral</td>
<td>3.5-5 mg</td>
</tr>
<tr>
<td>Diazepam</td>
<td>AU</td>
<td>Oral</td>
<td>5-10 mg</td>
</tr>
<tr>
<td><strong>Opioids</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Morphine hydrochloride</td>
<td>PU</td>
<td>IV</td>
<td>0.1 mg/kg (10 mg)</td>
</tr>
<tr>
<td>Meperidine</td>
<td>IV</td>
<td></td>
<td>0.5 mg/kg (50 mg)</td>
</tr>
<tr>
<td>Fentanyl</td>
<td>IV</td>
<td></td>
<td>1-2 µg/kg</td>
</tr>
<tr>
<td>Alfentanil</td>
<td>IV</td>
<td></td>
<td>10-20 µg/kg</td>
</tr>
<tr>
<td>Remifentanil</td>
<td>IV</td>
<td></td>
<td>Continuous infusion (0.05-0.1 µg/kg/min)</td>
</tr>
<tr>
<td>Sufentanil</td>
<td>IV</td>
<td></td>
<td>0.1 µg/kg</td>
</tr>
<tr>
<td><strong>Hypnotics</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Propofol</td>
<td>PU</td>
<td>IV</td>
<td>1-2 mg/kg</td>
</tr>
<tr>
<td>Etomidate</td>
<td>IV</td>
<td></td>
<td>0.2-0.3 mg/kg</td>
</tr>
<tr>
<td>Ketamine</td>
<td>IV</td>
<td></td>
<td>0.5-2 mg/kg</td>
</tr>
<tr>
<td><strong>NMBAs</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cisatracurium</td>
<td>PU</td>
<td>IV</td>
<td>0.5 mg/kg</td>
</tr>
<tr>
<td>Atracurium</td>
<td>IV</td>
<td></td>
<td>0.1-0.15 mg/kg</td>
</tr>
<tr>
<td>Rocuronium</td>
<td>IV</td>
<td></td>
<td>0.6 mg/kg</td>
</tr>
<tr>
<td>Vecuronium</td>
<td>IV</td>
<td></td>
<td>0.1 mg/kg</td>
</tr>
</tbody>
</table>

Abbreviations: AU, allergology unit; IV, intravenous; NMBAs, neuromuscular blocking agent; PU, postanesthetic unit; SC, subcutaneous.
than 30 minutes; (iv) monitoring of the patient during the
administered intravenously and most reactions appear in less
intervals between each drug. This is possible, as drugs are
different drugs consecutively on the same day, with 1-hour
This should yield negative skin test and BAT results, and, if
test a muscle relaxant other than that inducing the reaction.
enable better control in case of a reaction. It is recommended
recommended to test them after other drugs, such as propofol,
tests are not valuable. As NBMAs cannot be used alone, it is
exist [144]. Testing is indicated in specific situations where the
anesthetist needs to know tolerance to NMBAs, for example,
Drug release and hypotension with certain drugs (succinylcholine,
rocuronium, atracurium, mivacurium, morphine, and
meperidine), which can hamper diagnosis [143,145,151];
(iii) the complexity of the procedure requires DPT with
different drugs consecutively on the same day, with 1-hour
recovery phase after finishing the DPT.

The drugs that usually need to be tested are opioids,
because there is no alternative and hypnotics that are essential
for anesthetic induction, particularly propofol, are the most
widely used agents for anesthesia and sedation [78,80,152].
Testing is not recommended for muscle relaxants owing to
potential effects on breathing, with the exception of those cases
where they are suspected of causing a reaction and where they
are absolutely necessary to perform surgery and no alternatives
exist [144]. Testing is indicated in specific situations where the
anesthetist needs to know tolerance to NMBAs, for example,
in transplant procedures, high-risk patients, or when skin
tests are not valuable. As NBMAs cannot be used alone, it is
recommended to test them after other drugs, such as propofol,
to control the airway by endotracheal intubation, which will
enable better control in case of a reaction. It is recommended
to test a muscle relaxant other than that inducing the reaction.
This should yield negative skin test and BAT results, and, if
possible, be from a different chemical group, especially in
the case of cisatracurium and rocuronium [9,146,151].

7. Recommendations After the Allergological Study

Once the allergological study is finished, a medical
report must be produced with a detailed description of
the drugs involved, the type of reaction, the allergology
work-up results, diagnosis, and recommendations (grade of
recommendation, D). This will be essential when deciding on
the best anesthetic procedures for the patient in the future. The
various possibilities are as follows:
– Patient with a clinical history that not suggestive and
negative allergology findings.
If the patient has not presented a hypersensitivity drug
reaction, the recommendation is to use any anesthetic
procedure with the same risk as for the general population
(grade of recommendation, D).
– Patient with a suggestive clinical history and negative
allergology results.
In this scenario, the various possibilities are as follows:
- If the allergological study has only been based on
the performance of skin tests accompanied or not by
in vitro tests and a drug provocation test cannot be
performed, then it is not possible to rule out an allergic
reaction [109]. If the drugs involved in the reaction
are known, the best approach is to avoid them as well
as any others that may cause cross-reactivity (grade of
recommendation, D). If the drugs involved are
not known, the best approach is to use locoregional
anesthesia (if possible) or general anesthesia, but
without using NMBAs or drugs with a high capacity
to induce nonspecific histamine release (grade of
recommendation, D) [6]. Moreover, it is important
to re-evaluate the patient by testing emerging agents
such as chlorohexidine, methylcellulose, polyethylene
glycol/macrogol, mannitol, dyes, and any other drugs
not initially considered due to improbability. It is also
important to assess any methodological problems in
the testing procedures (failure in concentration or skin
test reading) (grade of recommendation, D).
- If the allergological study was based on provocation
testing, drugs with confirmed tolerance will be
recommended (grade of recommendation, D).
- In cases where nonspecific histamine release is
suspected, the recommendation is to avoid drugs
with potent histamine-releasing capacity the next
time anesthesia is administered. Pretreatment with
antihistamines is also recommended [11,153], and all
drugs should be administered slowly and one by one
[10,145] (grade of recommendation, D).
– Patient with a suggestive clinical history and positive
allergology results.

The recommendation is to avoid the agent identified as
allergic and those with cross-reactivity. In these
cases, pretreatment is not useful for preventing a new
reaction [154]. The recommendations will depend on the
culprit drug, as follows:
- NMBAs. The recommendation is to use a muscle
relaxant that yields negative results in intradermal
skin tests and the BAT [12,140]. Although there is
considerable cross-reactivity between muscle relaxants
(65% if the reaction was induced by rocuronium and
29% for succinylcholine) [155], sensitization to all of
them is low (grade of recommendation, C) [146]. If this
is the case and all NMBAs yield positive results in skin
tests and/or BAT, one might consider a locoregional
anesthesia (if possible) or general anesthesia using inhalant agents or combinations of
opioids and hypnotics (midazolam, propofol, fentanyl)
plus local anesthetics such as lidocaine [154-156]
(grade of recommendation, D).
– Hypnotics. Since hypnotics do not present cross-
reactivity, it should be possible to replace one hypnotic
with another. Currently, most reactions are due to
propofol. Ketamine and etomidate provide effective
dedation with limited effects on hemodynamic function.
The better alternative in patients with cardiovascular
disease is etomidate. Ketamine induces dissociative
anesthesia with minimum respiratory depression and
no cardiodepressant effects and is especially useful
in hemodynamically unstable patients or critically
ill patients. Thiopental is rarely used nowadays. In
cases of endoscopy, a combination of midazolam and
fentanyl has been recommended [157].
- Opioids. The incidence of anaphylaxis is quite low, and most cases are due to nonspecific histamine release. In such cases, it is important to avoid morphine, meperidine, and codeine, all of which have a high capacity for stimulating mast cells in the skin. Recommended alternatives include phenylpiperidine drugs (fentanyl, alfentanil, remifentanil, sufentanil), which have a low histamine release capacity. In IgE-mediated reactions induced by morphine, it seems useful to use remifentanil or fentanyl; morphine can be used in cases of reaction to fentanyl [83-86].

- Colloids/crystalloids. Gelatins and dextrans are the substances most frequently involved [158]. Cross-reactivity between them has not been reported.

- Latex. Every hospital should have protocols for operations involving patients with latex allergy. If there is no specific operating room, then the surgery should be performed early in the day and without materials containing latex in order to avoid the presence of latex particles in the environment [113]. Similar precautions should be taken when the patient is moved to the postsurgery room or any other part of the hospital [113].

- Local anesthetic. Reactions are quite infrequent and are generally not due to hypersensitivity. If confirmed, an alternative with confirmed tolerance can be used.

- Dyes. Reactions have been mainly reported for patent blue V and isosulfan blue, which cross-react. Methylene blue can be used as an alternative if the allergological study is negative [159].

- Antiseptics and sterilizers. In cases for which hypersensitivity to chlorhexidine or povidone-iodine is confirmed, they should be avoided, and other drugs without cross-reactivity should be used. When this is not possible, as with ethylene oxide, the area should be washed several times in physiological saline before use. The same procedure should be followed with ortho-phthalaldehyde solution, although this has generally been replaced by peracetic acid owing to its higher sensitizing capacity.

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Conflicts of Interest

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References

Perioperative Hypersensitivity Reactions


Perioperative Hypersensitivity Reactions


