

Hypersensitivity to Nonsteroidal Anti-inflammatory Drugs in Children and Adolescents: Cross-Intolerance Reactions

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■ Abstract

Nonsteroidal anti-inflammatory drugs (NSAIDs) are used worldwide and are responsible for several types of drug hypersensitivity reactions (DHRs) in all age groups. The 2 major groups of DHRs to NSAIDs are those induced by immunological mechanisms (selective reactions) and those where inflammatory mediators are released through activation of the prostaglandin-leukotriene pathway without specific immunological recognition (cross-intolerance). In the present review, we focus on cross-intolerance reactions, which are the most frequent DHRs and are becoming a topic of major interest in children and adolescents.

Paracetamol and ibuprofen are the drugs that most frequently cause DHRs in children; other NSAIDs are responsible for reactions in adolescents. *In vivo* and *in vitro* tests are of limited diagnostic value, with some exceptions for the less common selective reactions. In cross-intolerance, the clinical history and controlled administration are in many instances the only way to establish a diagnosis and look for alternatives. The clinical history is diagnostic when consistent symptoms occur repeatedly after exposure to NSAIDs with different chemical structures.

Cutaneous and respiratory symptoms often co-occur in young children. The natural history of these reactions in children is unknown, and some patients can develop tolerance over time. Atopy remains a major risk factor for cross-intolerant reactions. The increasing interest in hypersensitivity to NSAIDs with improvements in patient phenotyping and the information provided by pharmacogenetics will improve our understanding and management of these reactions in the near future.

Key words: Hypersensitivity drug reactions. NSAIDs, cross-intolerance. Cysteinyl leukotrienes. NSAID-exacerbated respiratory disease. NSAID-exacerbated cutaneous disease. NSAID-induced urticaria/angioedema.

■ Resumen

Los antiinflamatorios no esteroideos (AINEs) son ampliamente utilizados en todo el mundo y en todos los tramos de edad. Son responsables de un número importante de reacciones de hipersensibilidad a fármacos (RHF), que no sólo afectan a adultos sino también a niños y adolescentes. Existen dos grandes grupos: reacciones selectivas, inducidas por mecanismos inmunológicos específicos, y de intolerancia cruzada (IC), donde se liberan mediadores inflamatorios en ausencia de reconocimiento inmunológico específico. En esta revisión nos ocuparemos de la IC, que es la causa más frecuente de RHF y resulta de gran interés en niños y adolescentes.

El paracetamol y el ibuprofeno son los medicamentos más frecuentemente implicados en las RHF en niños. El uso diagnóstico de los tests *in vivo* e *in vitro* es muy limitado, con algunas excepciones en las reacciones selectivas. En las de IC, la historia clínica y la administración

controlada son en ocasiones la única vía para confirmar el diagnóstico y determinar las alternativas terapéuticas más adecuadas. La historia clínica tiene valor diagnóstico cuando se reproducen síntomas consistentes repetidamente tras la exposición a AINEs no relacionados estructuralmente.

En niños de corta edad es especialmente frecuente la combinación de síntomas cutáneos y respiratorios. Aunque se desconoce la historia natural de la IC en niños, es probable que se desarrolle tolerancia a lo largo de la vida.

El fenotipado detallado junto con la información proporcionada por la fármaco-genética no sólo proporcionarán un conocimiento más preciso de la IC sino que también facilitará el manejo clínico de estos pacientes.

Palabras clave: Reacciones de hipersensibilidad a fármacos. AINEs. Intolerancia cruzada. Cisteinil-leucotrienos. Enfermedad respiratoria exacerbada por AINEs. Enfermedad cutánea exacerbada por AINEs. Urticaria/angioedema inducidos por AINEs.

Introduction

Nonsteroidal anti-inflammatory drugs (NSAIDs) are among the most highly consumed drugs worldwide for all age groups [1,2]. They are used to treat pain, fever, and various inflammatory diseases [3]. Despite their beneficial effects, they can induce adverse drug reactions (ADRs) [1]. Some ADRs are dose-dependent (Type A), but others occur at therapeutic or even at low, nontherapeutic concentrations (Type B) [4]. The latter include drug hypersensitivity reactions (DHRs) [5]. Although originally reported in adults, it soon became clear that DHRs can also occur in children and adolescents [6,7].

NSAIDs, particularly paracetamol and ibuprofen, are commonly prescribed to children, among whom they have proven to be relatively safe, despite their widespread consumption [8-10]. Studies assessing the risk of serious ADRs due to ibuprofen in more than 80 000 febrile children reported only 795 hospital admissions [11,12]. Another study of 1879 febrile children with asthma showed that short-term use of ibuprofen can reduce asthma morbidity [13]. However, ibuprofen and paracetamol have recently been included in a list of 20 medications thought to be responsible for ADRs in children and adolescents [14].

This review deals specifically with DHRs to NSAIDs in children and adolescents. The many initiatives that have updated our understanding of these reactions in adults include a position statement on clinical entities [15], a new nomenclature for classification [16], and guidelines for advanced phenotyping [17]. Hypersensitivity to NSAIDs in children, however, has received much less attention [6,18].

Classification of DHRs to NSAIDs

As stated above, NSAIDs are widely consumed by patients of all ages [2,8-10] and responsible for at least 25% of all ADRs, including DHRs [5].

DHRs to NSAIDs can be caused by specific immunological mechanisms (allergic reactions) or by biochemical processes linked to arachidonic acid metabolism (nonallergic hypersensitivity or cross-intolerance [CI] reactions) [16]. Reactions induced by CI are frequent in all age groups, including children and adolescents [18,19]. DHRs to NSAIDs have more potential underlying mechanisms than DHRs to β -lactam antibiotics, which result from specific IgE or T-cell responses [20]. For example, NSAIDs-induced urticaria could

Table. Classification of Hypersensitivity Reactions to Nonsteroidal Anti-inflammatory Drugs

	Type of reaction	Clinical manifestations	Timing of the reaction	Underlying disease	Mechanism
Cross intolerance (nonallergic)	NSAID-exacerbated respiratory disease (NERD)	Bronchial obstruction, dyspnea and/or nasal congestion/rhinorrhea	Acute (immediate to several hours after exposure)	Asthma	COX-1 inhibition
	NSAID-exacerbated cutaneous disease (NECD)	Wheals and/or angioedema		Rhinosinusitis	
	NSAID-induced urticaria/angioedema (NIUA)	Wheals and/or angioedema		Chronic urticaria	COX-1 inhibition
	Single-NSAID-induced urticaria/anaphylaxis (SNIUAA)	Wheals/angioedema/anaphylaxis			Unknown, probably COX-1 inhibition
Selective response (allergic)	Single-NSAID-induced delayed reactions (SNIDR)	Maculopapular exathema	Delayed onset (usually more than 24 hours after exposure)		T cell-mediated
		Fixed drug eruption			
		Acute generalized exanthematous pustulosis			
		Drug reaction with eosinophilia and systemic symptoms			
		Stevens-Johnson syndrome/Toxic epidermal necrolysis			
		Organ-specific reactions			

be due to an IgE-dependent mechanism, a T-cell response, or CI [16]. Although some entities induced by CI occur in both adults and children [5,19], others, such as facial and lip angioedema, are more common in children [21,22].

According to the European Academy of Allergy and Clinical Immunology (EAACI) [16] the major entities induced by CI [Table] are as follows:

1. NSAIDs-exacerbated respiratory disease (NERD), which is observed in patients with underlying chronic respiratory disease (asthma/rhinosinusitis/nasal polyposis) aggravated by intake of NSAIDs. This condition was previously known as aspirin (ASA)-exacerbated respiratory disease (AERD), ASA-induced asthma (AIA), or the ASA triad.
2. NSAIDs-exacerbated cutaneous disease (NECD), which is observed in patients with a previous history of chronic spontaneous urticaria (CSU) aggravated by intake of NSAIDs.
3. NSAIDs-induced urticaria/angioedema (NIUA), in which patients develop symptoms following intake of NSAIDs in the absence of CSU.

CI reactions were previously known as idiosyncratic or pseudoallergic reactions [23]. The term cross-reactive should be reserved for reactions induced by specific immunological mechanisms, although many authors still use it [16]. Strong COX-1 inhibitors are usually responsible for CI, although weak COX-1 inhibitors and even selective COX-2 inhibitors can trigger CI [24]. The term *blended reaction* refers to CI with respiratory and cutaneous involvement [23], which can also affect children and adolescents following intake of NSAIDs [6,18].

Children and adolescents can be affected by any of the above-mentioned entities. However, the specific features associated with younger age groups include the type, severity, and frequency of the reaction and the drug involved [18,25].

Epidemiology

Although original epidemiological studies are somewhat scarce, several manuscripts deal with the prevalence of ADRs to NSAIDs [26-28], including DHRs [29,30], and some analyze DHRs to NSAIDs in the context of total ADRs [31-33]. Studies can also be based on spontaneous reporting [31], where the description of the entities and drug involvement are assigned by probability [34], without considering the specific mechanism (CI or selective reaction) [23,35].

Other studies examine specific conditions such as angioedema in children [22], angioedema plus urticaria [36], respiratory symptoms [37], and both skin and respiratory involvement [38]. However, most studies use mixed populations of adolescents and adults [29,39,40], and we must determine the exact percentage of children/adolescents before drawing conclusions. Some studies include patients aged ≥ 14 years [29]; few studies include children aged < 12 years and even fewer include children aged < 5 years [19,41]. Variability in study design also affects interpretation [42].

Studies from the 1970s and 1980s were neither sufficiently detailed nor supported by laboratory data [18,43] or used

imputability criteria that prevented a definitive diagnosis from being established [34,44]. In general, detailed allergological studies are scarce for both children and adults [18,45], although sufficient information is available to estimate the relevance of DHRs in children.

A recent study of 659 NSAID-hypersensitive adolescent/adult patients found that 76% had CI [29]. A retrospective study showed that antibiotics and NSAIDs were the most common triggers of DHRs [46]. As this study grouped all antibiotics together, we can infer that NSAIDs were the main culprits. A cross-sectional study in which 1015 patients were evaluated based on self-reported replies to a questionnaire showed that NSAIDs were the main cause of DHRs, followed by β -lactams and sulfonamides [47].

In a recent study of the largest series of DHRs to date, we confirmed that in 1682 of 4400 initial patients, NSAIDs were the culprit drugs in 47% of cases and β -lactam antibiotics in less than 20% [30]. Within NSAIDs, no differences in reaction patterns were found between adolescents and adults [30], a result that is consistent with that of previous studies [40].

NSAIDs were the main culprit drugs in another study of patients aged < 18 years with anaphylaxis [48]. Moreover, in a large cohort of children with anaphylaxis, most culprit drugs were NSAIDs, mainly ibuprofen [49].

Studies of DHRs in children do not usually assess the underlying mechanism, whose patterns of reactions may differ from those of adults [42]. Another study evaluating 3275 confirmed cases, of which 10% were children and 22% adolescents, found significant differences in the frequency of exanthematic reactions at younger ages [50].

The prevalence of NERD in adults ranges from 4.3% to 10.9% [51-53], although data are scarcer for children. In an early study, 28% of children with chronic asthma were intolerant to ASA [54]; however, another study found that oral administration of ASA did not lead to a significant effect on respiratory function [55].

Some studies have focused on the etiology and natural history of CSU in children, although none have assessed intolerance to NSAIDs [56-59].

NIUA was shown to account for 61% of patients with CI [29]. CI accounted for 47% of all DHRs followed by allergy to β -lactams [30]. NSAIDs frequently elicit isolated angioedema in children [60], as reported, occurring in more than 85% of children with CI after challenge [19].

Blended reactions must be differentiated from anaphylaxis in selective responses. They occur in 9%-40% of NSAID-induced reactions, ie, more frequently than NERD [29,36]. One study reported that blended reactions occur in 9.7% of children [61]. In a study of children aged 9-14 years, 14% of CI episodes following a drug provocation test (DPT) were blended reactions, in which the patients presented angioedema with asthma and/or rhinoconjunctivitis [73]. The reaction elicited (blended or respiratory) in children with CI varied with the provocation protocol used, as well as with the patient's age and genetic background. More paracetamol-induced reactions were observed in Asian children with early onset of CI, as was a lower incidence of respiratory symptoms upon challenge in younger patients [62].

Drugs Involved

All NSAIDs, including strong and weak COX-1 and selective COX-2 inhibitors, can induce CI [18,35] in all age groups [7,18]. NSAIDs are increasingly used in both children and adults owing to higher demand and the introduction of new compounds [7]. In addition to ibuprofen and paracetamol, consumption of aspirin, naproxen, indomethacin, and COX-2 inhibitors (at older ages) has increased [7].

Shortly after the first description of AIA, reactions to other NSAIDs were also described [63,64]. All strong COX-1 inhibitors can induce the same effects as ASA [35,65]. Interestingly, the first adult cases reported were considered “intrinsic asthmatics,” although the first pediatric cases were atopic [37,66,67].

Ibuprofen and paracetamol have been implicated in anaphylaxis in children, although the underlying mechanism was not assessed [49]. DHRs to paracetamol are frequent in patients with ibuprofen-induced anaphylaxis [36.8%], whereas a previous reaction to ibuprofen was reported in only a fifth of patients with paracetamol-induced anaphylaxis [49]. However, as with all DHRs, an oral DPT is frequently needed for diagnosis, as demonstrated by a study where DPT confirmed paracetamol hypersensitivity in only 4% of children [68].

Ibuprofen caused bronchospasm in a 17-year-old boy with AIA [69]. ASA, which is taken less frequently at younger ages, has been implicated in the development of urticaria and/or angioedema in atopic children [63] and in a child with a positive oral challenge result to paracetamol [68]. ASA has also been shown to worsen respiratory function in asthmatic children aged between 7 and 14 years [55].

As for NECD, adolescent data [70] indicate that patients with CSU might also experience exacerbations after intake of NSAIDs, particularly strong COX-1 inhibitors [61].

All NSAIDs including weak COX-1 and selective COX-2 inhibitors have been implicated in NIUA in adults and adolescents [24,29] and children [18]. Paracetamol and ibuprofen are the analgesics that most frequently induce DHRs in children. Paracetamol was the culprit drug in 5.5% of children [71], and up to 25% of children with CI showed a positive response to this drug [36,61,68,72]. However, the response to paracetamol can depend on age and other factors [62]. Anecdotal information suggests that hypersensitivity to paracetamol can resolve over time and that patients with positive challenge results at an early age may tolerate the drug years later. Therefore, assessment of hypersensitivity using DPT years after the incident could underestimate the true prevalence of the problem in children.

Blended reactions are common in children and have been reported for ibuprofen and other drugs [36].

Clinical Characteristics and Pathophysiology

NERD

NERD comprises a heterogeneous set of syndromes that involve the upper and/or lower airways. The ASA triad as initially described (asthma, rhinitis/nasal polyposis, and ASA

intolerance) is not fully expressed in children and adolescents but instead involves the upper/lower airways to varying degrees [18]. Asthma and nasal polyposis are extremely rare in children, although they have been reported [73,74]. Asthma and/or rhinitis induced by NSAIDs is more common [75].

Contrary to the first descriptions of AIA, where most patients were adults with a negative skin test result (diagnosed with “intrinsic asthma”), most children/adolescents with NERD are atopic, with positive skin test results to inhalant allergens, particularly house dust mites [18]. These patients usually develop mild airway symptoms with or without ocular involvement [76].

The exacerbation of asthma attacks and other respiratory manifestations is attributed to COX-1 inhibition, which shunts the arachidonic acid pathway from prostaglandins (PG) towards synthesis of cysteinyl leukotrienes (CysLTs) during inflammation. The degree of COX-1 inhibition differs according to the NSAID involved and correlates with its capacity to induce bronchospasm [65]. Inflammatory mediators, including LTC₄, LTD₄, and hydroxyeicosatetraenoic acids, also participate in the regulation of mucus glycoprotein secretion, which has a role in NSAIDs-induced asthma attacks [77]. An eicosanoid imbalance in children with NERD was recently reported [73].

An alternative hypothesis is that suppression of PGE₂ in chronic viral infection induces lymphocytes to attack target cells in the respiratory tract [78]. Interestingly, meclizolam can induce asthma in adolescent girls during the follicular phase of the menstrual cycle, probably as a consequence of monthly variations in serum PGF_{2α} [79]. Although this finding is common, no comprehensive detailed studies have been carried out.

NECD

Around 30% of patients with CSU experience a worsening of their symptoms after taking strong COX-1 inhibitors [80]. Although CSU is rare in children [81], it does occur [82-85] and can be exacerbated by NSAIDs, particularly strong COX-1 inhibitors [38]. A severe anaphylactic reaction to ibuprofen in a child with CSU and NSAID hypersensitivity has been described [38]. Reports are often insufficiently detailed to enable a precise, unequivocal diagnosis to be established [45].

Patients with NECD show increased N-methylhistamine metabolites and CysLT levels in urine [86,87]. The complex interaction of factors underlying CSU and NECD includes autoimmune diseases, allergens, infections, physical factors, and other as yet unidentified triggers that are also relevant in children [88].

NIUA

NIUA is the most common type of NSAID-induced DHR for all age groups [29,30], including children [19]. The most frequent clinical condition is facial angioedema followed by generalized urticaria [36], although both urticaria and angioedema can appear simultaneously [19], especially in atopic children [63].

Clinical entities are not always fully described, and despite suggestive symptoms, NIUA can sometimes only be putatively inferred. Anaphylaxis may also be induced by CI and must

be differentiated from selective responses [29]. Clinically, severe responses including skin and respiratory symptoms (ie, blended reactions) may be considered anaphylactic according to recent guidelines of the World Allergy Organization [89]. However, most NIUA reactions, even when severe, are not associated with an early drop in blood pressure. Likewise, although abdominal pain or discomfort may be part of a severe reaction, vomiting and diarrhea are rarely reported in children [36,38,62].

Whilst similarities have been found between the mechanisms underlying NECD and NERD [87], to our knowledge, no such data have been reported for NIUA. Preliminary evidence suggests that NIUA and NERD present different urinary eicosanoid profiles [90].

Nonpruriginous isolated angioedema, which often affects the face and other soft tissues, can be caused by NSAIDs [29,30,91,92]. The mediators involved remain unknown, although it is tempting to speculate on the participation of other mechanisms such as the bradykinin pathway [93].

Blended reactions

Blended reactions can be caused by high doses of ASA, leading to local release of histamine and CysLTs and causing vasoactive effects outside the lung. Pediatricians frequently deal with this type of reaction [19,71,94].

NSAIDs and Food Allergy Reactions

Food allergy has a prevalence of 7%-8% in children, with fruits, milk, and vegetables accounting for two-thirds of reactions [95]. Lipid transfer proteins, which are present in many fruits and nuts [96-98], are major triggers in children and adolescents, as are seafood and mite-contaminated food [99]. Ingestion of food allergens alongside NSAIDs (usually strong COX-1 inhibitors) can trigger anaphylaxis and urticaria/angioedema [99-101]. Physical exercise is often required as a cofactor [102,103]. In a typical scenario, a patient sensitized to a food allergen (eg, peanut/peach), but with no clinical manifestations, takes a previously tolerated NSAID (commonly ibuprofen) and develops anaphylaxis from minutes to hours after intensive physical exercise [102].

Risk Factors

Risk factors for developing CI include a previous history of anaphylaxis, immediate and accelerated reactions, atopy, older age, and CSU [61].

In children and adolescents with NERD or NIUA, skin test positivity to inhalant allergens (eg, house dust mites, pollens, and allergens such as *Alternaria*) has been reported [19]. Doña et al [29] found that sensitization to house dust mite and grass and olive pollen can be a risk factor.

Other risk factors include the number of drugs taken and sex, with female adults at higher risk [104]. However, sex has not been shown to increase the risk for children [19]. Although atopy is more frequent in patients with AIA and patients with

selective reactions to pyrazolones [67], this association was not found in a Spanish population [29]. Reduced use of ASA in favor of paracetamol in children could contribute to the increasing prevalence of asthma, atopic eczema, and allergic rhinitis in developed countries [105]. Atopy is also frequent in patients with NIUA, who were more likely to have high levels of specific IgE to *Dermatophagoides pteronyssinus* and *Blomia tropicalis* [106]. The association between *D pteronyssinus* and NIUA has also been reported in Spain [29].

Most studies of risk factors analyze the role of concomitant factors that are previous to or simultaneous with the drug (eg, food and exercise). NSAIDs can also influence the development and/or clinical course of rhinitis and asthma [107,108]. These cases do not involve hypersensitivity to NSAIDs but an interaction between NSAIDs and atopic status. The association between atopy, asthma, and NSAIDs is controversial, since it has been shown that short-term use of ibuprofen as anti-inflammatory medication in children can actually reduce the number of acute exacerbations [13].

The role of food allergy in the development of NECD or NIUA is controversial [70,80]. There is evidence that food allergy-induced anaphylaxis can occur after intake of ASA in patients with no previous history of DHRs to NSAIDs [109]. NSAIDs and/or exercise can facilitate absorption of allergens from the gastrointestinal tract [110], and ASA can induce mast cell activation leading to anaphylaxis, as seen in the skin prick test with the causative food allergen [111]. In our experience, adolescents with NIUA are not more sensitized to prevalent food allergens such as Pru p 3 or other lipid transfer proteins [112], although further studies are needed to confirm this observation.

Familial clustering has been observed in some types of hypersensitivity to NSAIDs, although a clear Mendelian pattern has not been found [113]. Most genetic studies of CI investigated NERD and polymorphisms in candidate genes [114], although 2 genome-wide association studies have been conducted [115,116]. Data are also available for NIUA [117,118], including a genome-wide association study that suggested a role for second messenger signaling pathways [119].

Diagnosis

DHRs to NSAIDs are incorrectly diagnosed more frequently than DHRs to other drugs [120]. The diagnosis of CI is based on a thorough clinical history, as diagnostic tests are of little value, in particular skin tests and the recently developed in vitro tests [104].

Clinical History

The expertise gained from NERD and NECD in adults is valuable for adolescents and children [15,16,121]. In NIUA, there is some disagreement as to whether the clinical history is valuable for diagnosis in adults/adolescents [104,122] and children [123].

In a large series of children with histories suggestive of DHRs, only 2.5% could be confirmed, and ibuprofen was the only culprit NSAID identified [124]. However,

details of whether the ibuprofen-induced reactions were CI or selective responses were not provided. In a retrospective study of patients with a history of hypersensitivity to NSAIDs, diagnosis was confirmed by challenge in only 8% of cases of asthma and 12% of cases of angioedema [125]. Oral challenge confirmed CI in 44% of patients in a study of children with a reaction suggestive of DHRs to NSAIDs [123].

The clinical history should include the sequence of events, symptoms, time between drug intake and the reaction, time between the reaction and the study, and the reason for administration [104]. We found that diagnosis of NIUA based on the clinical history was confirmed by challenge in 76% of cases. This percentage increased to 92% when more than 2 different NSAIDs were involved and the clinical history was clear [104]. However, there is some disagreement on this matter [122].

In Vitro Tests

A limited number of in vitro tests are available, although more research is needed to compare their efficacy [126].

The cellular allergen stimulation test, which quantitates basophil LT release, has been proposed for the diagnosis of AIA and can be extended to the other NERD entities (rhinitis and nasal polyposis) [127]. Initial studies showed promising results [128], which could not be confirmed in subsequent studies [129,130]. The basophil activation test has also been proposed [131], although its specificity is low [130,132].

In vitro assays for NECD are similar to those described for NERD but more difficult to apply and very unlikely to be used for children owing to the low prevalence of NECD in this group.

The use of the cellular allergen stimulation test and the basophil activation test to diagnose NIUA has been assessed [127]. Although there has been some success, several drawbacks remain [133]. The EAACI does not consider these techniques useful for diagnosis [16].

Drug Provocation Test

The principles used for DPT in children have been extrapolated from adults, except for the dose [5,16,18]. In many instances, the approach for evaluating hypersensitivity to NSAIDs is to assess tolerance rather than to identify the culprit drug [33]. We must also consider whether the alternative drug is a strong, weak, or selective COX-2 inhibitor. Since the reaction is expected with DPTs, low doses should be administered at regular intervals until the cumulative dose reaches the necessary therapeutic dose [36]. Furthermore, given that CI is the effect of an abnormal pharmacologic response, it is reasonable to increase the time between doses from the 30 minutes recommended for most challenges. Most published pediatric protocols use an interval of 60-90 minutes [28,50]. Some extremely sensitive patients may respond to lower doses than those usually recommended [134]. Guidelines for DPT have recently been published [135].

In NERD, DPT can be performed by nasal inhalation and bronchial or oral administration, with the latter considered the gold standard [135]. In adolescents aged 14-20 years, nasal inhalation (sensitivity of 80%-90%) is recommended [136].

If the result is negative, the bronchial provocation test can be considered. Bronchial provocation with lysine-ASA has long been used for the diagnosis of AIA and rhinitis in adolescents and adults, but less so in children [135].

Younger children should undergo oral DPT [61,63]. In one study, the NSAID was given incrementally every 20-30 minutes until the recommended dose was reached [125]. In a typical 25-kg patient, a proposed schedule for diagnosis of AIA is 25, 50, 100, and 250 mg every 30 minutes [137]. An alternative is to administer a constant dose at 1-hour intervals until the recommended therapeutic dose for age is reached [36,137]. According to this schedule, a therapeutic dose of 20 mg/kg for a 25-kg child is given as 4 doses of 125 mg at 1-hour intervals.

As stated above, NECD is uncommon in children. However, if younger patients with CSU must receive paracetamol or ibuprofen, a DPT should be performed to assess tolerance [38], as described below for NIUA.

In patients with NIUA, other drugs, such as paracetamol, tramadol, and selective COX-2 inhibitors, have been shown to exert little or no COX-1 inhibition [138]. Oral antihistamine has proven useful for preventing urticaria [138]. In otherwise healthy children with confirmed ibuprofen-induced NIUA, a DPT with paracetamol up to a cumulative dose of 15-20 mg/kg is imperative to enable safe antipyretic treatment as needed. In younger patients, most protocols use either equal or incremental doses at 1-hour intervals until the appropriate dose is reached [28,50]. In children with a positive result to a DPT with paracetamol or a convincing history of repeated reactions induced by paracetamol-containing formulations, the only alternative antipyretic would be off-label use of a specific COX-2 inhibitor. Although most such drugs are not indicated for fever or for children aged under 12 years, their safety has been proven in most patients in this age group [62,134].

Further Research

Childhood is the period during which we generally receive NSAIDs for the first time. As we grow older, the drugs taken in the first years of our lives may play a role in the development of future reactions and act as risk factors. Although data are available mainly for NERD and, to a lesser extent, NIUA, no well-defined genetic markers have been identified. The study of biological samples (urine, nasal and bronchoalveolar lavage fluid, saliva, and skin biopsy specimens) will be necessary to improve our understanding of the mechanisms involved in DHRs to NSAIDs and, subsequently, diagnosis of these reactions. Gene expression studies can also be useful for unraveling these mechanisms. Precise, advanced patient phenotyping [17] combined with molecular data will shed new light on our understanding of these diseases and thus facilitate patient management.

Concluding Remarks

NSAIDs are the drugs most commonly involved in DHRs in childhood [19]. They are frequently prescribed for the management of fever, pain, and other processes, and there are few alternatives, thus making understanding and management

of DHRs crucial for health care professionals. DHRs to NSAIDs are more complex than reactions to other drugs, such as β -lactams, owing to the variety of triggering mechanisms (eg, IgE-dependent reactions, T-cell effector responses, and common CI-induced reactions). A precise diagnosis is often difficult owing to the limitations of *in vitro* and *in vivo* tests. Their complexity is demonstrated by the fact that exanthema-like reactions or urticaria, which are frequent in children and adults, may belong to any of these categories. We hope that this review will not only improve our understanding of DHRs to NSAIDs, but that it will also stimulate further studies of clinical and basic aspects of these diseases.

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Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- Moore TJ, Weiss SR, Kaplan S, Blaisdell CJ. Reported adverse drug events in infants and children under 2 years of age. *Pediatrics*. 2002;110(5):e53.
- Conaghan PG. A turbulent decade for NSAIDs: update on current concepts of classification, epidemiology, comparative efficacy, and toxicity. *Rheumatol Int*. 2012;32(6):1491-1502.
- Roberts LJM, Morrow JD. Analgesic-antipyretic and antiinflammatory agents and drugs employed in the treatment of gout. In: Hardman JG, Limbird LL, Goodman Gilman A, eds. *The pharmacological basis of therapeutics*, 10th edn. New York: McGraw-Hill. 2001:687-731.
- Rawlins MDT, Thompson W. Mechanisms of adverse drug reactions. In: Davies DM ed. *Textbook of adverse drug reactions*. New York, NY, Oxford University Press. 1991:18-45.
- Torres MJ, Barrionuevo E, Kowalski M, Blanca M. Hypersensitivity reactions to nonsteroidal anti-inflammatory drugs. *Immunol Allergy Clin North Am*. 2014;34(3):507-24.
- Solensky RM, Mendelson LM. Drug allergy. In: Leung DYM, Sampson HA, Geha R, Szefer SJ, eds. *Pediatric Allergy, Principles and practice* 2nd edn. St Louis: Mosby Elsevier. 2010:616-30.
- Titchen T, Cranswick N, Beggs S. Adverse drug reactions to nonsteroidal anti-inflammatory drugs, COX-2 inhibitors and paracetamol in a paediatric hospital. *Br J Clin Pharmacol*. 2005;59(6):718-23.
- Eustace N, O'Hare B. Use of nonsteroidal anti-inflammatory drugs in infants. A survey of members of the Association of Paediatric Anaesthetists of Great Britain and Ireland. *Paediatr Anaesth*. 2007;17(5):464-9.
- Neubert A, Verhamme K, Murray ML, Picelli G, Hsia Y, Sen FE, Giaquinto C, Ceci A, Sturkenboom M, Wong IC. The prescribing of analgesics and non-steroidal anti-inflammatory drugs in paediatric primary care in the UK, Italy and the Netherlands. *Pharmacol Res*. 2010;62(3):243-8.
- Valkhoff VE, Schade R, t Jong GW, Romio S, Schuemie MJ, Arfe A, Garbe E, Herings R, Lucchi S, Picelli G, Schink T, Straatman H, Villa M, Kuipers EJ, Sturkenboom MC. Population-based analysis of non-steroidal anti-inflammatory drug use among children in four European countries in the SOS project: what size of data platforms and which study designs do we need to assess safety issues? *BMC Pediatr*. 2013;13:192.
- Lesko SM. The safety of ibuprofen suspension in children. *Int J Clin Pract Suppl*. 2003(135):50-3.
- Bigby M, Jick S, Jick H, Arndt K. Drug-induced cutaneous reactions. A report from the Boston Collaborative Drug Surveillance Program on 15,438 consecutive inpatients, 1975 to 1982. *JAMA*. 1986;256(24):3358-63.
- Lesko SM, Louik C, Vezina RM, Mitchell AA. Asthma morbidity after the short-term use of ibuprofen in children. *Pediatrics*. 2002;109(2):E20.
- Lee WJ, Lee TA, Pickard AS, Caskey RN, Schumock GT. Drugs associated with adverse events in children and adolescents. *Pharmacotherapy*. 2014;34(9):918-26.
- Kowalski ML, Makowska JS, Blanca M, Bavbek S, Bochenek G, Bousquet J, Celik G, Demoly P, Gomes ER, Nizankowska-Mogilnicka E, Romano A, Sanchez-Borges M, Sanz M, Torres MJ, De Weck A, Szczeklik A, Brockow K. Hypersensitivity to nonsteroidal anti-inflammatory drugs (NSAIDs) - classification, diagnosis and management: review of the EAACI/ENDA and GA2LEN/HANNA. *Allergy*. 2011;66(7):818-29.
- Kowalski ML, Asero R, Bavbek S, Blanca M, Blanca-Lopez N, Bochenek G, Brockow K, Campo P, Celik G, Cernadas J, Cortellini G, Gomes E, Nizankowska-Mogilnicka E, Romano A, Szczeklik A, Testi S, Torres MJ, Wohrl S. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*. 2013;68(10):1219-32.
- Ayuso P, Blanca-Lopez N, Dona I, Torres MJ, Gueant-Rodriguez RM, Canto G, Sanak M, Mayorga C, Gueant JL, Blanca M, Cornejo-García JA. Advanced phenotyping in hypersensitivity drug reactions to NSAIDs. *Clin Exp Allergy*. 2013;43(10):1097-109.
- Sanchez-Borges M, Capriles-Behrens E, Caballero-Fonseca F. Hypersensitivity to non-steroidal anti-inflammatory drugs in childhood. *Pediatr Allergy Immunol*. 2004;15(4):376-80.
- Zambonino MA, Torres MJ, Munoz C, Requena G, Mayorga C, Posadas T, Urda A, Blanca M, Corzo JL. Drug provocation tests in the diagnosis of hypersensitivity reactions to non-steroidal anti-inflammatory drugs in children. *Pediatr Allergy Immunol*. 2013;24(2):151-9.
- Blanca M, Romano A, Torres MJ, Fernandez J, Mayorga C, Rodriguez J, Demoly P, Bousquet PJ, Merk HF, Sanz ML, Ott H, Atanaskovic-Markovic M. Update on the evaluation of hypersensitivity reactions to betalactams. *Allergy*. 2009;64(2):183-93.

21. Kidon MI, Kang LW, Chin CW, Hoon LS, Hugo VB. Nonsteroidal anti-inflammatory drug hypersensitivity in preschool children. *Allergy Asthma Clin Immunol.* 2007;3(4):114-22.
22. Capriles-Behrens E, Caplin J, Sanchez-Borges M. NSAID facial angioedema in a selected pediatric atopic population. *J Investig Allergol Clin Immunol.* 2000;10(5):277-9.
23. Stevenson DD, Sanchez-Borges M, Szczeklik A. Classification of allergic and pseudoallergic reactions to drugs that inhibit cyclooxygenase enzymes. *Ann Allergy Asthma Immunol.* 2001;87(3):177-80.
24. Doña I, Blanca-Lopez N, Jagemann LR, Torres MJ, Rondon C, Campo P, Gómez AI, Fernandez J, Laguna JJ, Rosado A, Blanca M, Canto G. Response to a selective COX-2 inhibitor in patients with urticaria/angioedema induced by nonsteroidal anti-inflammatory drugs. *Allergy.* 2011;66(11):1428-33.
25. Blake KV, Zaccaria C, Domergue F, La Mache E, Saint-Raymond A, Hidalgo-Simon A. Comparison between paediatric and adult suspected adverse drug reactions reported to the European medicines agency: implications for pharmacovigilance. *Paediatr Drugs.* 2014;16(4):309-19.
26. Gomes ER, Demoly P. Epidemiology of hypersensitivity drug reactions. *Curr Opin Allergy Clin Immunol.* 2005;5(4):309-16.
27. Demoly P, Gomes ER. Drug hypersensitivities: definition, epidemiology and risk factors. *Eur Ann Allergy Clin Immunol.* 2005;37(6):202-6.
28. Gomes E, Cardoso MF, Praca F, Gomes L, Marino E, Demoly P. Self-reported drug allergy in a general adult Portuguese population. *Clin Exp Allergy.* 2004;34(10):1597-1601.
29. Doña I, Blanca-Lopez N, Cornejo-Garcia JA, Torres MJ, Laguna JJ, Fernandez J, Rosado A, Rondon C, Campo P, Agundez JA, Blanca M, Canto G. Characteristics of subjects experiencing hypersensitivity to non-steroidal anti-inflammatory drugs: patterns of response. *Clin Exp Allergy.* 2011;41(1):86-95.
30. Doña I, Blanca-Lopez N, Torres MJ, Garcia-Campos J, Garcia-Nunez I, Gomez F, Salas M, Rondon C, Canto MG, Blanca M. Drug hypersensitivity reactions: response patterns, drug involved, and temporal variations in a large series of patients. *J Investig Allergol Clin Immunol.* 2012;22(5):363-71.
31. Chen CJ, Cheng CF, Lin HY, Hung SP, Chen WC, Lin MS. A comprehensive 4-year survey of adverse drug reactions using a network-based hospital system. *J Clin Pharm Ther.* 2012;37(6):647-51.
32. Rebelo Gomes E, Fonseca J, Araujo L, Demoly P. Drug allergy claims in children: from self-reporting to confirmed diagnosis. *Clin Exp Allergy.* 2008;38(1):191-8.
33. Heinzerling LM, Tomsitz D, Anliker MD. Is drug allergy less prevalent than previously assumed? A 5-year analysis. *Br J Dermatol.* 2012;166(1):107-14.
34. Naranjo CA, Busto U, Sellers EM, Sandor P, Ruiz I, Roberts EA, Janecek E, Domecq C, Greenblatt DJ. A method for estimating the probability of adverse drug reactions. *Clin Pharmacol Ther.* 1981;30(2):239-45.
35. Cornejo-Garcia JA, Blanca-Lopez N, Dona I, Andreu I, Agundez JA, Carballo M, Blanca M, Canto MG. Hypersensitivity reactions to non-steroidal anti-inflammatory drugs. *Curr Drug Metab.* 2009;10(9):971-80.
36. Kidon MI, Kang LW, Chin CW, Hoon LS, See Y, Goh A, Lin JT, Chay OM. Early presentation with angioedema and urticaria in cross-reactive hypersensitivity to nonsteroidal anti-inflammatory drugs among young, Asian, atopic children. *Pediatrics.* 2005;116(5):e675-80.
37. Botey J, Navarro C, Marin A, Eserverri JL. Aspirin-induced asthma in children. *Allergol Immunopathol (Madr).* 1988;16(3):145-9.
38. Kang LW, Kidon MI, Chin CW, Hoon LS, Hwee CY, Chong NK. Severe anaphylactic reaction to ibuprofen in a child with recurrent urticaria. *Pediatrics.* 2007;120(3):e742-4.
39. Quiralte J, Blanco C, Delgado J, Ortega N, Alcántara M, Castillo R, Anguita JL, Saenz de San Pedro B, Carrillo T. Challenge-based clinical patterns of 223 Spanish patients with nonsteroidal anti-inflammatory-drug-induced-reactions. *J Investig Allergol Clin Immunol.* 2007;17(3):182-8.
40. Messaad D, Sahla H, Benahmed S, Godard P, Bousquet J, Demoly P. Drug provocation tests in patients with a history suggesting an immediate drug hypersensitivity reaction. *Ann Intern Med.* 2004;140(12):1001-6.
41. Ferrandiz-Pulido C, Garcia-Fernandez D, Dominguez-Sampedro P, Garcia-Patos V. Stevens-Johnson syndrome and toxic epidermal necrolysis in children: a review of the experience with paediatric patients in a university hospital. *J Eur Acad Dermatol Venereol.* 2011;25(10):1153-9.
42. Porto Arceo JA. Special features of NSAID intolerance in children. *Allergol Immunopathol (Madr).* 2003;31(3):109-25.
43. Canto MG, Andreu I, Fernandez J, Blanca M. Selective immediate hypersensitivity reactions to NSAIDs. *Curr Opin Allergy Clin Immunol.* 2009;9(4):293-7.
44. Strom BL. Study design available for pharmacoepidemiology studies. In: Strom BL, ed. *Pharmacoepidemiology.* New York: Churchill Livingstone. 1989:13-26.
45. Kay E, Ben-Shoshan M. Anaphylaxis to ibuprofen in a 12-year-old boy. *BMJ Case Rep.* 2013;2013.
46. Chalabianloo F, Berstad A, Schjott J, Riedel B, Irgens A, Florvaag E. Clinical characteristics of patients with drug hypersensitivity in Norway: a single-centre study. *Pharmacoepidemiol Drug Saf.* 2011;20(5):506-13.
47. Ensina LF, Amigo MH, Koch T, Guzman E, Paoli R, Nunes IC. Drug hypersensitivity in students from Sao Paulo, Brazil. *Clinics (Sao Paulo).* 2010;65(10):1009-11.
48. Faria E, Rodrigues-Cernadas J, Gaspar A, Botelho C, Castro E, Lopes A, Gomes E, Malheiro D, Cadinha S, Campina-Costa S, Neto M, Sousa N, Rodriguez-Alves R, Romeira A, Caiado J, Morais-Almeida M. Drug-induced anaphylaxis survey in Portuguese Allergy Departments. *J Investig Allergol Clin Immunol.* 2014;24(1):40-8.
49. Liew WK, Chiang WC, Goh AE, Lim HH, Chay OM, Chang S, Tan JH, Shih E, Kidon M. Paediatric anaphylaxis in a Singaporean children cohort: changing food allergy triggers over time. *Asia Pac Allergy.* 2013;3(1):29-34.
50. Rubio M, Bousquet PJ, Gomes E, Romano A, Demoly P. Results of drug hypersensitivity evaluations in a large group of children and adults. *Clin Exp Allergy.* 2012;42(1):123-30.
51. Hedman J, Kaprio J, Poussa T, Nieminen MM. Prevalence of asthma, aspirin intolerance, nasal polyposis and chronic obstructive pulmonary disease in a population-based study. *Int J Epidemiol.* 1999;28(4):717-22.
52. Vally H, Taylor ML, Thompson PJ. The prevalence of aspirin intolerant asthma (AIA) in Australian asthmatic patients. *Thorax.* 2002;57(7):569-74.
53. Kasper L, Sladek K, Duplaga M, Bochenek G, Liebhart J, Gladysz U, Malolepszy J, Szczeklik A. Prevalence of asthma

- with aspirin hypersensitivity in the adult of Poland. *Allergy*. 2003;58(10):1064-6.
54. Rachelefsky GS, Coulson A, Siegel SC, Stiehm ER. Aspirin intolerance in chronic childhood asthma: Detected by oral challenge. *Pediatrics*. 1975;56(3):443-8.
 55. Schuhl JF, Pereyra JG. Oral acetylsalicylic acid (aspirin) challenge in asthmatic children. *Clin Allergy*. 1979;9(1):83-8.
 56. Chansakulporn S, Pongpreuksa S, Sangacharoenkit P, Pacharn P, Visitsunthorn N, Vichyanond P, Jirapongsananuruk O. The natural history of chronic urticaria in childhood: A prospective study. *J Am Acad Dermatol*. 2014;71(4):663-8.
 57. Harris A, Twarog FJ, Geha RS. Chronic urticaria in childhood: natural course and etiology. *Ann Allergy*. 1983;51(2Pt1):161-5.
 58. Jirapongsananuruk O, Pongpreuksa S, Sangacharoenkit P, Visitsunthorn N, Vichyanond P. Identification of the etiologies of chronic urticaria in children: a prospective study of 94 patients. *Pediatr Allergy Immunol*. 2010;21(3):508-14.
 59. Sahiner UM, Civelek E, Tuncer A, Yavuz ST, Karabulut E, Sackesen C, Sekerel BE. Chronic urticaria: etiology and natural course in children. *Int Arch Allergy Immunol*. 2011;156(2):224-30.
 60. Ertoy Karagol HI, Yilmaz O, Bakirtas A, Topal E, Demirsoy MS, Turktaş I. Angioedema without urticaria in childhood. *Pediatr Allergy Immunol*. 2013;24(7):685-90.
 61. Hassani A, Ponvert C, Karila C, Le Bourgeois M, De Blic J, Scheinmann P. Hypersensitivity to cyclooxygenase inhibitory drugs in children: a study of 164 cases. *Eur J Dermatol*. 2008;18(5):561-5.
 62. Kidon MI, Liew WK, Chiang WC, Lim SH, Goh A, Tang JP, Chay OM. Hypersensitivity to paracetamol in Asian children with early onset of nonsteroidal anti-inflammatory drug allergy. *Int Arch Allergy Immunol*. 2007;144(1):51-6.
 63. Botey J, Navarro C, Aulesa C, Marin A, Eserverri JL. Acetyl salicylic acid induced-urticaria and/or angioedema in atopic children. *Allergol Immunopathol (Madr)*. 1988;16(1):43-7.
 64. Fahrenholz JM. Natural history and clinical features of aspirin-exacerbated respiratory disease. *Clin Rev Allergy Immunol*. 2003;24(2):113-24.
 65. Szczeklik A, Gryglewski RJ, Czerniawska-Mysik G, Zmuda A. Aspirin-induced asthma. Hypersensitivity to fenoprofen and ibuprofen in relation to their inhibitory action on prostaglandin generation by different microsomal enzymic preparations. *J Allergy Clin Immunol*. 1976;58(1Pt1):10-8.
 66. Estrada Rodriguez JL, Florido Lopez JF, Belchi Hernandez J, Martin Munoz F, Diaz Pena JM, Garcia Ara MC, Sastre Dominguez J, Ojeda Casas A. Asthma in children and ASA intolerance. *J Investig Allergol Clin Immunol*. 1993;3(6):315-20.
 67. Bochenek G, Nizankowska E, Szczeklik A. The atopy trait in hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*. 1996;51(1):16-23.
 68. Boussetta K, Ponvert C, Karila C, Bourgeois ML, Blic J, Scheinmann P. Hypersensitivity reactions to paracetamol in children: a study of 25 cases. *Allergy*. 2005;60(9):1174-7.
 69. Palmer GM. A teenager with severe asthma exacerbation following ibuprofen. *Anaesth Intensive Care*. 2005;33(2):261-5.
 70. Doña I, Blanca-Lopez N, Torres MJ, Gomez F, Fernandez J, Zambonino MA, Monteseirin FJ, Canto G, Blanca M, Cornejo-Garcia JA. NSAID-induced urticaria/angioedema does not evolve into chronic urticaria: a 12-year follow-up study. *Allergy*. 2014;69(4):438-44.
 71. Corzo JL, Zambonino MA, Munoz C, Mayorga C, Requena G, Urda A, Gallego C, Blanca M, Torres MJ. Tolerance to COX-2 inhibitors in children with hypersensitivity to nonsteroidal anti-inflammatory drugs. *Br J Dermatol*. 2014;170(3):725-9.
 72. Pastorello EA, Zara C, Riario-Sforza GG, Pravettoni V, Incorvaia C. Atopy and intolerance of antimicrobial drugs increase the risk of reactions to acetaminophen and nimesulide in patients allergic to nonsteroidal anti-inflammatory drugs. *Allergy*. 1998;53(9):880-4.
 73. Olze H, Lau S, Forster U. Samter's triad and eicosanoid imbalance in children with recurrent nasal polyps. *Pediatr Allergy Immunol*. 2012;23(5):500.
 74. Chen BS, Virant FS, Parikh SR, Manning SC. Aspirin sensitivity syndrome (Samter's Triad): an unrecognized disorder in children with nasal polyposis. *Int J Pediatr Otorhinolaryngol*. 2013;77(2):281-3.
 75. Tan Y, Collins-Williams C. Aspirin-induced asthma in children. *Ann Allergy*. 1982;48(1):1-5.
 76. Weinberger M. Analgesic sensitivity in children with asthma. *Pediatrics*. 1978;62(5 Pt 2 Suppl):910-5.
 77. Lundgren JD, Shelhamer JH, Kaliner MA. The role of eicosanoids in respiratory mucus hypersecretion. *Ann Allergy*. 1985;55(1):5-8, 11.
 78. Szczeklik A. Aspirin-induced asthma as a viral disease. *Clin Allergy*. 1988;18(1):15-20.
 79. Eliasson O, Longo M, Dore-Duffy P, Densmore MJ, DeGraff AC, Jr. Serum 13-14-diOH-15-keto-prostaglandin F2 alpha and airway response to meclofenamate and metaproterenol in relation to the menstrual cycle. *J Asthma*. 1986;23(6):309-19.
 80. Asero R. Intolerance to nonsteroidal anti-inflammatory drugs might precede by years the onset of chronic urticaria. *J Allergy Clin Immunol*. 2003;111(5):1095-8.
 81. Church MK, Weller K, Stock P, Maurer M. Chronic spontaneous urticaria in children: itching for insight. *Pediatr Allergy Immunol*. 2011;22(1 Pt 1):1-8.
 82. Tuchinda M, Srimaruta N, Habanananda S, Varenil J, Assatherawatts A. Urticaria in Thai children. *Asian Pac J Allergy Immunol*. 1986;4(1):41-5.
 83. Volonakis M, Katsarou-Katsari A, Stratigos J. Etiologic factors in childhood chronic urticaria. *Ann Allergy*. 1992;69(1):61-5.
 84. Ghosh S, Kanwar AJ, Kaur S. Urticaria in children. *Pediatr Dermatol*. 1993;10(2):107-10.
 85. Khakoo G, Sofianou-Katsoulis A, Perkin MR, Lack G. Clinical features and natural history of physical urticaria in children. *Pediatr Allergy Immunol*. 2008;19(4):363-6.
 86. Oosting E, Kardaun SH, Doeglas HM, Los P, de Monchy JG. Increased urinary excretion of the histamine metabolite N tau-methylhistamine during acetylsalicylic acid provocation in chronic urticaria patients. *Agents Actions*. 1990;30(1-2):254-7.
 87. Mastalerz L, Setkowicz M, Sanak M, Szczeklik A. Hypersensitivity to aspirin: common eicosanoid alterations in urticaria and asthma. *J Allergy Clin Immunol*. 2004;113(4):771-5.
 88. Brunetti L, Francavilla R, Miniello VL, Platzer MH, Rizzi D, Lospalluti ML, Poulsen LK, Armenio L, Skov PS. High prevalence of autoimmune urticaria in children with chronic urticaria. *J Allergy Clin Immunol*. 2004;114(4):922-7.

89. Simons FE, Arduzzo LR, Dimov V, Ebisawa M, El-Gamal YM, Lockey RF, Sanchez-Borges M, Senna GE, Sheikh A, Thong BY, Worm M. World Allergy Organization Anaphylaxis Guidelines: 2013 update of the evidence base. *Int Arch Allergy Immunol*. 2013;162(3):193-204.
90. Ayuso P, Plaza MC, Doña I, Blanca-López N, Mayorga C, Barrionuevo E, Meléndez L, Godineau V, Galindo L, Torres MJ, Canto MG, Blanca M, Campo P. Urinary eicosanoids excretion profile in NIUA and NERD patients after NSAIDs challenge. *Allergy*. 2014;69 (Suppl. 99):129-30.
91. Zhu D, Becker WM, Schulz KH, Schubeler K, Schlaak M. The presence of specific IgE to salicyloyl and O-methylsalicyloyl in aspirin-sensitive patients. *Asian Pac J Allergy Immunol*. 1992;10(1):25-32.
92. Caimmi S, Caimmi D, Bousquet PJ, Demoly P. How can we better classify NSAID hypersensitivity reactions? validation from a large database. *Int Arch Allergy Immunol*. 2012;159(3):306-12.
93. Sala-Cunill A, Bjorkqvist J, Senter R, Guilarte M, Cardona V, Labrador M, Ninnaro A, Kenne E, Jamsa A, Krieger T, Schluter H, Fuchs T, Flohr S, Hassiepen U, Cumin F, McCrae K, Maas C, Stavrou E, Renne T. Plasma contact system activation drives anaphylaxis in severe mast cell-mediated allergic reactions. *J Allergy Clin Immunol*. 2015;135(4):1031-43.
94. Iancovici Kidon M, Abramovitch I, Steinberg S, Barash J. Cross-reactive hypersensitivity to COX inhibitors in a child with mild allergic rhinitis. *Isr Med Assoc J*. 2005;7(12):790-1.
95. Sicherer SH, Sampson HA. Food allergy: Epidemiology, pathogenesis, diagnosis, and treatment. *J Allergy Clin Immunol*. 2014;133(2):291-307; quiz 308.
96. Bock SA, Buckley J, Holst A, May CD. Proper use of skin tests with food extracts in diagnosis of hypersensitivity to food in children. *Clin Allergy*. 1977;7(4):375-83.
97. Novembre E, de Martino M, Vierucci A. Foods and respiratory allergy. *J Allergy Clin Immunol*. 1988;81(5 Pt 2):1059-65.
98. Pastorello EA, Ortolani C, Farioli L, Pravettoni V, Ispano M, Borgia A, Ispano M, Borgia A, Bengtsson A, Incorvaia C, Berti C, Zanussi C. Allergenic cross-reactivity among peach, apricot, plum, and cherry in patients with oral allergy syndrome: an in vivo and in vitro study. *J Allergy Clin Immunol*. 1994;94(4):699-707.
99. Sanchez-Borges M, Suarez Chacon R, Capriles-Hulett A, Caballero-Fonseca F, Fernandez-Caldas E. Anaphylaxis from ingestion of mites: pancake anaphylaxis. *J Allergy Clin Immunol*. 2013;131(1):31-5.
100. Sanchez-Borges M, Capriles-Hulett A, Capriles-Behrens E, Fernandez-Caldas E. A new triad: sensitivity to aspirin, allergic rhinitis, and severe allergic reaction to ingested aeroallergens. *Cutis*. 1997;59(6):311-4.
101. Blanco C, Quiralte J, Castillo R, Delgado J, Arteaga C, Barber D, Carrillo T. Anaphylaxis after ingestion of wheat flour contaminated with mites. *J Allergy Clin Immunol*. 1997;99(3):308-313.
102. Fujii H, Kambe N, Fujisawa A, Kohno K, Morita E, Miyachi Y. Food-dependent exercise-induced anaphylaxis induced by low dose aspirin therapy. *Allergol Int*. 2008;57(1):97-8.
103. Romano A, Scala E, Rumi G, Gaeta F, Caruso C, Alonzi C, Maggioletti M, Ferrara R, Palazzo P, Palmieri V, Zeppilli P, Mari A. Lipid transfer proteins: the most frequent sensitizer in Italian subjects with food-dependent exercise-induced anaphylaxis. *Clin Exp Allergy*. 2012;42(11):1643-53.
104. Blanca-Lopez N, M JT, Dona I, Campo P, Rondon C, Seoane Reula ME, Salas M, Canto G, Blanca M. Value of the clinical history in the diagnosis of urticaria/angioedema induced by NSAIDs with cross-intolerance. *Clin Exp Allergy*. 2013;43(1):85-91.
105. Varner AE, Busse WW, Lemanske RF, Jr. Hypothesis: decreased use of pediatric aspirin has contributed to the increasing prevalence of childhood asthma. *Ann Allergy Asthma Immunol*. 1998;81(4):347-51.
106. Sanchez-Borges M, Acevedo N, Carballo L, Capriles-Hulett A, Caballero-Fonseca F. Increased total and mite-specific immunoglobulin E in patients with aspirin-induced urticaria and angioedema. *J Investig Allergol Clin Immunol*. 2010;20(2):139-45.
107. Kumar P, Bryan C, Hwang D, Kadowitz P, Butcher B, Leech SH. Allergic rhinitis relieved by aspirin and other nonsteroidal antiinflammatory drugs. *Ann Allergy*. 1988;60(5):419-22.
108. Barr RG, Kurth T, Stampfer MJ, Buring JE, Hennekens CH, Gaziano JM. Aspirin and decreased adult-onset asthma: randomized comparisons from the physicians' health study. *Am J Respir Crit Care Med*. 2007;175(2):120-5.
109. Harada S, Horikawa T, Ashida M, Kamo T, Nishioka E, Ichihashi M. Aspirin enhances the induction of type I allergic symptoms when combined with food and exercise in patients with food-dependent exercise-induced anaphylaxis. *Br J Dermatol*. 2001;145(2):336-9.
110. Matsuo H, Morimoto K, Akaki T, Kaneko S, Kusatake K, Kuroda T, Niihara H, Hide M, Morita E. Exercise and aspirin increase levels of circulating gliadin peptides in patients with wheat-dependent exercise-induced anaphylaxis. *Clin Exp Allergy*. 2005;35(4):461-6.
111. Aihara M, Miyazawa M, Osuna H, Tsubaki K, Ikebe T, Aihara Y, Ikezawa Z. Food-dependent exercise-induced anaphylaxis: influence of concurrent aspirin administration on skin testing and provocation. *Br J Dermatol*. 2002;146(3):466-72.
112. Doña I, Salas M, Ruiz MD, Blazquez AB, Blanca-Lopez N, Torres MJ, Campo P, Canto G, Blanca M. Associations of food sensitisation and multiple hypersensitivity to non-steroidal antiinflammatory drugs. *Allergy*. 2013;68 (Suppl. 97):519-20.
113. Mastalerz L, Setkowicz M, Sanak M, Rybarczyk H, Szczeklik A. Familial aggregation of aspirin-induced urticaria and leukotriene C synthase allelic variant. *Br J Dermatol*. 2006;154(2):256-60.
114. Kim SH, Sanak M, Park HS. Genetics of hypersensitivity to aspirin and nonsteroidal anti-inflammatory drugs. *Immunol Allergy Clin North Am*. 2013;33(2):177-94.
115. Kim JH, Park BL, Cheong HS, Bae JS, Park JS, Jang AS, Uh ST, Choi JS, Kim YH, Kim MK, Choi IS, Cho SH, Choi BW, Park CS, Shin HD. Genome-wide and follow-up studies identify CEP68 gene variants associated with risk of aspirin-intolerant asthma. *PLoS One*. 2010;5(11):e13818.
116. Park BL, Kim TH, Kim JH, Bae JS, Pasaje CF, Cheong HS, Kim LH, Park JS, Lee HS, Kim MS, Choi IS, Choi BW, Kim MK, Shin S, Shin HD, Park CS. Genome-wide association study of aspirin-exacerbated respiratory disease in a Korean population. *Hum Genet*. 2013;132(3):313-21.
117. Cornejo-García JA, Jagemann LR, Blanca-Lopez N, Dona I, Flores C, Gueant-Rodriguez RM, Torres MJ, Fernandez J, Laguna JJ, Rosado A, Agundez JA, Garcia-Martin E, Canto G, Gueant JL, Blanca M. Genetic variants of the arachidonic acid

- pathway in non-steroidal anti-inflammatory drug-induced acute urticaria. *Clin Exp Allergy*. 2012;42(12):1772-81.
118. Cornejo-García JA, Flores C, Plaza-Seron MC, Acosta-Herrera M, Blanca-Lopez N, Dona I, Torres MJ, Mayorga C, Gueant-Rodriguez RM, Ayuso P, Fernandez J, Laguna JJ, Agundez JA, Garcia-Martin E, Gueant JL, Canto G, Blanca M. Variants of CEP68 gene are associated with acute urticaria/angioedema induced by multiple non-steroidal anti-inflammatory drugs. *PLoS One*. 2014;9(3):e90966.
 119. Cornejo-García JA, Liou LB, Blanca-Lopez N, Dona I, Chen CH, Chou YC, Chuang HP, Wu JY, Chen YT, Plaza-Serón MC, Mayorga C, Gueant-Rodriguez RM, Lin SC, Torres MJ, Campo P, Rondon C, Laguna JJ, Fernandez J, Gueant JL, Canto G, Blanca M, Lee MT. Genome-wide association study in NSAID-induced acute urticaria/angioedema in Spanish and Han Chinese populations. *Pharmacogenomics*. 2013;14(15):1857-69.
 120. Reisfeld S, Goldberg A, Confino-Cohen R. Management of patients with known drug hypersensitivity in an emergency department in Israel. *Int Arch Allergy Immunol*. 2011;155(4):361-6.
 121. Asero R. Oral aspirin challenges in patients with a history of intolerance to single non-steroidal anti-inflammatory drugs. *Clin Exp Allergy*. 2005;35(6):713-6.
 122. Viola M, Rumi G, Valluzzi RL, Gaeta F, Caruso C, Romano A. Assessing potential determinants of positive provocation tests in subjects with NSAID hypersensitivity. *Clin Exp Allergy*. 2011;41(1):96-103.
 123. Yilmaz O, Ertoy Karagol IH, Bakirtas A, Topal E, Celik GE, Demirsoy MS, Turktaş I. Challenge-proven nonsteroidal anti-inflammatory drug hypersensitivity in children. *Allergy*. 2013;68(12):1555-61.
 124. Atanaskovic-Markovic M, Gaeta F, Gavrovic-Jankulovic M, Cirkovic Velickovic T, Valluzzi RL, Romano A. Diagnosing multiple drug hypersensitivity in children. *Pediatr Allergy Immunol*. 2012;23(8):785-91.
 125. Cormican LJ, Farooque S, Altmann DR, Lee TH. Improvements in an oral aspirin challenge protocol for the diagnosis of aspirin hypersensitivity. *Clin Exp Allergy*. 2005;35(6):717-22.
 126. Mayorga C, Sanz ML, Gamboa P, Garcia-Aviles MC, Fernandez J, Torres MJ. In vitro methods for diagnosing nonimmediate hypersensitivity reactions to drugs. *J Investig Allergol Clin Immunol*. 2013;23(4):213-25.
 127. de Weck AL, Sanz ML. Cellular allergen stimulation test (CAST) 2003, a review. *J Investig Allergol Clin Immunol*. 2004;14(4):253-73.
 128. Sanz ML, Gamboa P, de Weck AL. A new combined test with flowcytometric basophil activation and determination of sulfidoleukotrienes is useful for in vitro diagnosis of hypersensitivity to aspirin and other nonsteroidal anti-inflammatory drugs. *Int Arch Allergy Immunol*. 2005;136(1):58-72.
 129. Bavbek S, Dursun AB, Birben E, Kalayci O, Misirligil Z. Cellular allergen stimulation test with acetylsalicylic acid-lysine is not a useful test to discriminate between asthmatic patients with and without acetylsalicylic acid sensitivity. *Int Arch Allergy Immunol*. 2009;149(1):58-64.
 130. Sanz ML, Gamboa PM, Mayorga C. Basophil activation tests in the evaluation of immediate drug hypersensitivity. *Curr Opin Allergy Clin Immunol*. 2009;9(4):298-304.
 131. Celik GE, Schroeder JT, Hamilton RG, Saini SS, Adkinson NF. Effect of in vitro aspirin stimulation on basophils in patients with aspirin-exacerbated respiratory disease. *Clin Exp Allergy*. 2009;39(10):1522-31.
 132. Mayorga C, Sanz ML, Gamboa PM, Garcia BE, Caballero MT, Garcia JM, Labrador M, Lahoz C, Longo Areso N, Lopez Hoyos M, Martinez Quesada J, Monteseirin FJ. In vitro diagnosis of immediate allergic reactions to drugs: an update. *J Investig Allergol Clin Immunol*. 2010;20(2):103-9.
 133. Ebo DG, Leysen J, Mayorga C, Rozieres A, Knol EF, Terreehorst I. The in vitro diagnosis of drug allergy: status and perspectives. *Allergy*. 2011;66(10):1275-86.
 134. Sanchez-Borges M, Caballero-Fonseca F, Capriles-Hulett A. Tolerance of nonsteroidal anti-inflammatory drug-sensitive patients to the highly specific cyclooxygenase 2 inhibitors rofecoxib and valdecoxib. *Ann Allergy Asthma Immunol*. 2005;94(1):34-8.
 135. Nizankowska-Mogilnicka E, Bochenek G, Mastalerz L, Swierczynska M, Picado C, Scadding G, Kowalski ML, Setkowicz M, Ring J, Brockow K, Bachert C, Wohrl S, Dahlen B, Szczeklik A. EAACI/GA2LEN guideline: aspirin provocation tests for diagnosis of aspirin hypersensitivity. *Allergy*. 2007;62(10):1111-8.
 136. Campo P, Ayuso P, Salas M, Plaza MC, Cornejo-García JA, Doña I, Torres MJ, Blanca-López N, Canto G, Gueant JL, Sanak M, Blanca M. Mediator release after nasal aspirin provocation supports different phenotypes in subjects with hypersensitivity reactions to NSAIDs. *Allergy*. 2013;68(8):1001-7.
 137. Nizankowska E, Bestynska-Krypel A, Cmiel A, Szczeklik A. Oral and bronchial provocation tests with aspirin for diagnosis of aspirin-induced asthma. *Eur Respir J*. 2000;15(5):863-9.
 138. Asero R. Cetirizine premedication prevents acute urticaria induced by weak COX-1 inhibitors in multiple NSAID reactors. *Eur Ann Allergy Clin Immunol*. 2010;42(5):174-7.
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