Supplementary material

Oral food challenges with shrimp

Shrimp was obtained from the sea by fisherman, who were instructed to not add any additives

and to place the shrimp at -20oC immediately after catching. Shrimp was kept at -20oC, and at

the time of use, shrimp was thawed, had their shell removed, and boiled in water for 10 minutes.

For the double-blind-placebo-controlled shrimp challenge, shrimp was crushed using a food

processor, and added to a mixture of hazelnut cream, oat flakes, milk-based ice cream with small

bits of chocolate, powdered chocolate and vanilla extract, containing 15.4 % of fat. Prior to

challenge, patients were asked about any reactions to each of the components of the mixture. The

cumulative dose of 1g shrimp protein was the maximum we could use to allow shrimp taste to be

disguised in the food matrix. Placebo challenges were carried out using the same mixture minus

shrimp and similar timing schedule. For the open challenge, cooked shrimp prepared as described

for the DBPCFC was lightly salted and seasoned with olive oil. Active and placebo challenges

were performed one week apart.

IgE Immunoblots

Natural Lit v 1 and recombinant Per a 7 were prepared in our laboratory according to previously

published methods (Santos et al). Protein concentration of shrimp extract, Lit v 1 and Per a 7 was

measured by Bradford assay, using Bovine Serum Albumin (BSA) as standard. Fifty micrograms

of protein/lane were applied to 10% Sodium Dodecyl Sulfate Polyacrylamide Electrophoresis

(SDS-PAGE) gels. Electrophoresis was carried out under reducing conditions with beta-

mercaptoethanol. Protein transferring to nitrocellulose membranes (Millipore, Bedford, MA) was

confirmed by Ponceau S staining. Membranes were blocked with Phosphate Buffered Saline

(PBS) + 0.1% Tween + 5% skim milk (blocking solution) for 1 hour at room temperature. After

washing, sera were added at 1:10 dilution in blocking solution, and membranes incubated

overnight at 4°C under agitation. Bound specific IgE was detected by peroxidase-conjugated goat

anti-human IgE antibody (Kirkegaard and Perry, Gaithersburg, MD) diluted 1:5,000 in blocking

solution, using enhanced chemiluminescence solutions (GE Healthcare) and the ChemiDoc

Imaging System equipped with the ImageLab software (Bio-Rad Laboratories, Hercules, CA).

Reference

Santos AB, Rocha GM, Oliver C, Ferriani VP, Lima RC, Palma MS, Sales VS, Aalberse RC,

Chapman MD, Arruda LK. Cross-reactive IgE antibody responses to tropomyosins from Ascaris

lumbricoides and cockroach. J Allergy Clin Immunol. 2008 Apr;121(4):1040-6

J Investig Allergol Clin Immunol 2019; Vol. 29(4): 302-305 doi: 10.18176/jiaci.0378