SUPPLEMENTARY MATERIAL

Basophil activation test

BAT was performed on lithium-heparin whole blood samples within 2 hours after collection. Samples were stimulated for 20 minutes at 37°C with a 10-fold serial dilution (100 µg/mL to 0.1 µg/mL) of five whole-egg extracts dissolved in a calcium-containing aqueous buffer supplemented with 60 ng/mL interleukin (IL-)3 (Preprotech). This resulted in 9 ng/mL IL-3 in the final reaction volume, considering the 1/10 dilution and incubation of 30 µl of IL-3 buffer diluted stimulus in 150 µl blood. These extracts from progressively less heated forms of egg being cake (35' at 165°C), hard-boiled egg (10' at 100°C), omelet (4' at 120°C), soft-boiled egg (5' at 100°C), and raw egg were prepared following the same protocol as previously described (see section below) (20). Buffer without stimulus, polyclonal goat anti-human IgE (5 µg/mL, Sigma-Aldrich), monoclonal mouse anti-human IgE (5 µg/mL, BD Biosciences) and formyl-methionyl-leucyl-phenylalanine (fMLP, 2 µM, Sigma-Aldrich) were used as controls. Basophil degranulation was halted by placing the samples on ice followed by staining with anti-CD123 PE (clone 6H6, Biolegend), anti-HLA-DR AF647 (clone L243, Biolegend) and anti-CD63 FITC (Clone H5C6, Biolegend). After red blood cell lysis and washing, a minimum of 500 basophils, gated as SSC^{low} CD123⁺ HLADR⁻ CD63⁺ cells, were acquired on the LSR Fortessa SORP flow cytometer running DIVA software (BD biosciences) and analysed with FlowJo 10.8.1 for Windows. Basophil activation was measured as the %CD63⁺ basophils, corrected for baseline CD63-expression by subtracting the %CD63⁺ basophils in the unstimulated control condition. Generally, we observed that baseline CD63-expression did not exceed 2.5% in any of the unstimulated control conditions (9). Children with less than 5% activated basophils in response to both monoclonal and polyclonal anti-IgE were considered non-releasing, henceforth termed non-responders.

Preparation of the five egg extracts

Cake was prepared by mixing 4 eggs with 200 g wheat flour, 200 g sugar and 200 g plant-based butter, and baked for 35 minutes at 165°C (329°F). Hard-boiled egg was boiled for 10 minutes, soft-boiled egg for 5 minutes and raw egg was gently mixed (no heat treatment). Omelet was heated on a stove stop for 3 to 4 minutes at 120°C (250°F) on both sides. In brief, the five egg preparations were diluted 1:5 with phosphate-buffered saline. The extracts were shaken at 4°C for four hours, and centrifuged for 10 minutes at 2130 g. The extracts were disposed of and supernatants centrifuged for 10 minutes at 2130 g. The extracts were decanted and filter-sterilized through a 0.2 μ m filter. The total protein concentrations were determined using the Bicinchoninic acid assay, and can be found in our previously published work (20).