Anaphylaxis: Mediators, Biomarkers, and Microenvironments

Fernandez-Bravo S¹, Palacio-Garcia L¹, Requena-Robledo N¹, Yuste-Montalvo A¹, Nuñez-Borque E^{1*}, Esteban V^{1,2*}

¹Department of Allergy and Immunology, IIS-Fundación Jiménez Díaz, UAM, Madrid, Spain ²Faculty of Medicine and Biomedicine, Alfonso X El Sabio University, Madrid, Spain *Equal contribution to this work.

J Investig Allergol Clin Immunol 2022; Vol. 32(6): 419-437 doi: 10.18176/jiaci.0854

Abstract

The life-threatening nature of anaphylactic reactions has increased interest in discovering new biomarkers that could improve diagnosis and prevention. However, the diverse nature of the clinical features and the etiology and pathogenesis of anaphylaxis hinder the identification of valuable molecular indicators of disease.

Most studies on anaphylaxis focus on the immune system. Anaphylactic reactions are characterized primarily by IgE-mediated activation of mast cells and basophils and release of mediators. Determination of serum tryptase levels is the main in vitro test used to confirm the reaction, although there are no biomarkers that can predict it. Nevertheless, recent research has postulated that alternative pathways, cell types, and systems are involved. Consequently, various molecular products have been explored and considered potential biomarkers, although none of them are yet used in clinical practice. The products that are altered in patients with anaphylaxis include vasoactive agents, proteases, proteoglycans, lipids, interleukins, cytokines, products of the complement-contact and coagulation systems, circulating proteins, extracellular vesicles, microRNAs, and metabolites. The recognition of biological processes and molecular pathways affecting the microenvironments involved in anaphylaxis will considerably improve clinical practice and the identification of better molecular markers. We offer a broad review of the various mediators described in anaphylaxis, consider their usefulness as potential biomarkers of this pathological event, and examine their role in the molecular basis of the reaction.

Key words: Anaphylaxis. Biomarker. Mast cells. Mediators. Microenvironments.

Resumen

La naturaleza potencialmente mortal de las reacciones anafilácticas promueve un creciente interés por descubrir nuevos biomarcadores que puedan ayudar a su diagnóstico y prevención. Sin embargo, tanto las características clínicas como la etiopatología de la anafilaxia son muy diversas, lo que dificulta la identificación de marcadores moleculares precisos.

La mayoría de los estudios sobre la anafilaxia se han centrado en el sistema inmunitario. En concreto, estas reacciones se caracterizan principalmente por la activación de mastocitos y basófilos mediada por IgE y la liberación de diferentes mediadores. Entre ellos, la determinación de los niveles de triptasa sérica es la principal prueba *in vitro* utilizada para confirmar la reacción y no existen biomarcadores con capacidad predictiva. Sin embargo, estudios recientes han postulado que existen otros tipos celulares, sistemas y vías de señalización alternativos implicados en la anafilaxia. En consecuencia, se han explorado e identificado diferentes moléculas como potenciales biomarcadores, pero ninguno de ellos se ha trasladado aún a la práctica clínica. Precisamente, los agentes vasoactivos, las proteasas, los proteoglicanos, los derivados lipídicos, las interleuquinas, las citoquinas, los productos de los sistemas de complemento-contacto y de coagulación, las proteínas circulantes, las vesículas extracelulares, los microARN y los metabolitos, se encuentran alterados en pacientes con anafilaxia. El reconocimiento de los procesos biológicos y de las vías moleculares que interactúan en la magnitud de los microambientes implicados en la anafilaxia mejorará notablemente la práctica clínica y el reconocimiento de marcadores moleculares. Por lo tanto, este artículo abarca una amplia revisión de los distintos mediadores descritos en la anafilaxia y de su propuesta como biomarcadores de este evento patológico, así como su implicación en las bases moleculares de la reacción.

Palabras clave: Biomarcador. Mastocitos. Mediadores. Microambientes.

Introduction

Anaphylaxis is an acute systemic reaction and the most severe manifestation of allergic disorders. The incidence of this pathological event is underdiagnosed owing to the lack of effective biomarkers. The main molecular marker used in clinical practice is serum tryptase (sT), which is not elevated in most cases. In this regard, anaphylactic reactions are classified into different phenotypes and endotypes, with mast cells (MCs) and basophils as the main releasers of this biomarker. However, other molecular and cellular components have also been characterized in human anaphylaxis [1].

The term *biomarker* includes any biological observation that replicates or predicts a clinically relevant endpoint. However, it can also serve to predict diseases, to identify cells involved in their etiology and pathogenesis, and to assess an individual's response to treatment. In clinical practice, a biomarker is usually a mediator involved in the cellular basis of the disease [2]. Recent key statements in anaphylaxis point to the necessity to identify reliable diagnostic, predictive, and prognostic biomarkers [3]. Therefore, the aims of this review are to delve into the different molecules altered during anaphylaxis and to determine their role in the microenvironments involved in the reaction.

1. Anaphylaxis

The World Allergy Organization defines anaphylaxis as follows: "Anaphylaxis is a serious systemic hypersensitivity reaction that is usually rapid in onset and may cause death. Severe anaphylaxis is characterized by potentially lifethreatening compromise in airway, breathing and/or the circulation, and may occur without typical skin features or circulatory shock being present" [4]. The most common signs and symptoms are observed in the skin and mucous membranes (80-90%), followed by the respiratory system (70%). Less common, however, is the involvement of the gastrointestinal (45%), cardiovascular (45%), and nervous systems (15%) [5,6].

The incidence and prevalence of anaphylaxis has increased worldwide over the last 2 decades [7-10]. This pathological event can be caused by various substances, with food and drugs being the most common triggers. Foods are the most frequent in children, while drugs are more prevalent in adults [11]. However, epidemiological data are underestimated because of the absence of biomarkers and the ambiguity of the diagnosis of anaphylaxis [4,5].

Diagnosis is based on the recognition of signs and symptoms [4,12]. Moreover, it could be complemented by measurement of sT, the main biomarker used in clinical practice [4]. Over the years, many studies have reported this molecule to be increased in patients' sera [13-17]. sT peaks appear from 15 minutes after the onset of reaction symptoms and last for several hours [18,19]. However, levels do not always correlate with the severity of anaphylaxis and remain low in 36%-50% of events [14,20], especially in mild cases, where the reaction is not well diagnosed clinically [20].

Despite the wide variability in sT values, the clinical threshold for this protein in anaphylaxis is set at \geq 11.4 ng/mL [14,21]. Nevertheless, there have also been reports of cases of MC activation with sT levels below this cut-off point [15,22,23]. Therefore, sT should be assessed at least 24 hours after the episode [4,19]. Specifically, the difference between the acute peak of this protein and its baseline value was greater in anaphylaxis than in other diseases related to MC activation [19]. Accordingly, other sT thresholds proposed for the diagnosis of anaphylaxis include an increase of 20%+2 ng/mL in the acute condition compared to baseline [15,19,24]. In addition, a study carried out in Hymenoptera venom–allergic patients defined MC activation as acute sT values $\geq 135\%$ relative to baseline [22]. One recent study points to an improved diagnostic capacity of tryptase when personalized medicine is applied based on the patient's baseline status [25].

2. Pathophysiology of Anaphylaxis

Anaphylaxis is an acute systemic reaction involving several organs and systems. It is caused by the simultaneous activation of multiple molecular mechanisms and the sudden release of mediators, giving rise to signs and symptoms [26]. For many years, studies have focused on the analysis of the immune component, although the importance of other altered systems in etiology and pathogenesis has been demonstrated [27,28]. Other, as yet unexplored mechanisms likely contribute to the release of these or other mediators (Table). Therefore, understanding the plethora of specific microenvironments is essential when attempting to identify appropriate biomarkers and apply appropriate management based on precision medicine [29].

2.1 Immune System

Anaphylaxis has traditionally been considered a type I hypersensitivity reaction based on an IgE-mediated immunological mechanism. When it occurs, mediators are released from activated MCs and basophils, thus inducing the signs and symptoms of this pathological event [18]. However, degranulation of these cells can be achieved by different surface receptors and mechanisms depending on the nature of the stimulus (Figure 1) [30,31].

IgE-mediated reactions are divided into 2 phases: a sensitization phase and a re-exposure/activation phase. In primary sensitization, allergens induce the release of IgE into the bloodstream. This then binds to the high-affinity receptor for IgE (FccRI) on the surface of MCs and basophils [32,33]. However, symptoms appear after a new exposure to the allergen, when activation of MCs and basophils and sudden release of mediators occur [34,35].

However, some patients experience anaphylaxis with undetectable allergen-specific IgE levels according to skin or in vitro testing. Furthermore, elevated levels of allergen-specific IgE have been found in patients who have not experienced a reaction [34,36-38]. Therefore, these events point to the presence of other processes and cells involved in the reaction.

IgG-mediated anaphylaxis was first characterized in mouse models [39,40]. This pathway has since been described in humans and could also be responsible for numerous episodes of IgE-independent reactions, although the number of studies



Table. Mediators Released From Activated Cells and Systems in Anaphylaxis

Abbreviations: B, basophil; C/C/C, contact, coagulation, and complement systems; CCL-2, C-C motif chemokine ligand 2; E, erythrocyte; EC, endothelial cell; EOS, eosinophil; G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; IFN-γ, interferon gamma; IL, interleukin; LTB; C, D, E4, leukotrienes B, C, D, E4; MAC, macrophage; MC, mast cell; M-CSF, macrophage colony-stimulating factor; MON, monocyte; MPO, myeloperoxidase N, neutrophil; TL, T lymphocyte; PAF, platelet-activating factor; PF4, platelet factor 4; PGD2, prostaglandin D2; PGE2, prostaglandin E2; PGF2, prostaglandin F2; PLAT, platelet; T lymphocyte; RAS, renin-angiotensin system; SP, substance P; TGF-β, transforming growth factor beta; TNF, tumor necrosis factor (*1 TNFα members family including TNF-α, TNFRI, and TWEAK); VEGF, vascular endothelial growth factor; VSMC, vascular smooth muscle cell; P, plasma; S1P, sphingosine-1-phosphate; TXA, thromboxane A2; TXB2, thromboxane B2.



Figure 1. MCs are activated through multiple molecular mechanisms in anaphylaxis. Classically, MCs can be activated through cross-linking of FcεRI (upper left panel). They can also respond to IgG immune complexes (lower left panel). Other ligands and/or signals activate MCs through receptors such as MRGPRX2, complement receptors, and many other receptors. APC indicates antigen-presenting cell; TH cell, helper T lymphocytes; B cell, B lymphocytes; FcεRI, Fc epsilon receptor I; FcγRI, Fc gamma receptor I; MRGPRX2, Mas-related G-protein coupled receptor member X2; C3a, complement component 3a; C5a, complement component 5a; C3aR, complement component 3a receptor; C5aR, complement component 5a receptor; PAR2, protease activated receptor 2; CysLTRs, cysteinyl leukotriene receptors.

in humans remains limited [41-43]. IgG antibodies act through binding to Fc gamma receptors (Fc γ Rs), which are expressed in various cell types such as macrophages, dendritic cells, neutrophils, platelets, basophils, and MCs [43-46]. Specifically, neutrophils are thought to be the main cells involved in IgGmediated human anaphylaxis [41,42,44]. They are activated by the formation of IgG–allergen immune complexes and the subsequent binding to Fc γ R receptors on their surface, leading to the release of many protein and lipid mediators [35,36,43]. These include platelet activating factor (PAF), which is one of the main molecules stored in neutrophils and whose role in the pathogenesis of anaphylaxis has been demonstrated both in mice and humans [43,47].

2.1.1 Anaphylactic mediators released by immune cells

In anaphylaxis, the release of molecules previously stored in granules or newly synthesized molecules has traditionally been understood to be derived from MCs and basophils. However, the discovery of other cell types and the subsequent release of other mediators show anaphylactic reactions to be heterogeneous in nature. Among the many molecules released, tryptase is the main biomarker in clinical practice, although many others have been proposed.

2.1.1.1 Histamine

Histamine is a diamine prestored in granules of MCs and basophils and is considered, together with tryptase, one of

J Investig Allergol Clin Immunol 2022; Vol. 32(6): 419-437 doi: 10.18176/jiaci.0854 the main mediators of anaphylactic reactions [48,49]. The mechanism of action of this vasoactive peptide is through 4 receptors (H1, H2, H3, and H4), which are expressed in numerous cell types [49-51]. Their activation gives rise to multiple intracellular signalling pathways, thus inducing many of the symptoms of anaphylaxis, such as increased vascular dilatation and permeability, as well as constriction of the airways [34,52,53].

In anaphylaxis, the increase in plasma histamine levels shows them to be a promising biomarker [13,54,55]. A study of 97 patients visiting the emergency department shows that almost half of them present elevated serum histamine levels. Likewise, another study of 76 patients found that molecule levels are significantly higher in severe reactions than in moderate reactions [54,55]. Moreover, detection of histamine in body fluids is simple, and some studies have even proposed it as a more sensitive diagnostic molecule than tryptase. However, its use as a potential diagnostic marker has several limitations [55,56], including its short half-life (approximately 30 minutes), peaking 5-10 minutes after initiation of the reaction, thus hampering sample collection [56,57].

2.1.1.2 Enzymes: proteases and peroxidases

Among the proteases released by the degranulation of effector cells, sT is the most relevant and easily detectable molecule in blood samples from patients with anaphylaxis. However, other proteases have also been considered candidate biomarkers [36].

Carboxypeptidase A is a zinc-dependent metalloprotein that can also be found as a preformed mediator in basophils [56,58,59]. The levels of this protein are elevated in the serum of patients with anaphylaxis and in those who die from drug-induced reactions [60]. However, their levels do not necessarily correlate with those of sT and may even be increased in cases where sT levels remain low [36].

Chymase is a serine proteinase whose biological effects include activation of the renin angiotensin system. Chymase activity is heparin-dependent, and levels are elevated in the serum of anaphylactic patients up to 24 hours after onset of the reaction [61,62]. In addition, a study of patients who died of anaphylaxis showed not only increased chymase levels, but also that their value correlated with those of sT [63]. Moreover, the renin-angiotensin system is the main homeostatic regulator of blood pressure, contributing to other processes, such as proliferation, fibrosis, and inflammation [64]. One study of 50 patients with anaphylaxis due to hymenoptera venom showed that renin, angiotensinogen, angiotensin I, and angiotensin II levels decreased significantly and in accordance with the severity of the reaction [65].

Elastase is a serine protease released mainly by neutrophils, although it may also be present in the granules of basophils and MCs [66]. This protein can cleave the kinin light chain activating the contact system [61,67]. Furthermore, myeloperoxidase is a potent oxidizing enzyme found in neutrophil azurophilic granules. Neutrophil extracellular traps, which are detected as DNA-myeloperoxidase complexes together with elastase, have been described as increased during human anaphylaxis in line with the severity of the episodes [41,43,47].

2.1.1.3 Lipid molecules

Bioactive lipids are involved in immune function and vascular biology. Consequently, their relevance in allergic diseases and anaphylaxis has also been recognized, both in terms of the underlying molecular basis and as a complementary biomarker of sT [68,69].

PAF is a biologically active phospholipid that induces platelet aggregation and plays key roles in cardiovascular pathophysiology [70]. This mediator is released by multiple cell types and is involved in processes such as inflammation, proliferation, and cell adhesion [71,72]. In anaphylaxis, PAF induces the release of several cytokines and participates in bronchoconstriction, hypotension, and endothelial permeability [73]. In addition, its elevation is associated with hemodynamic changes that occur in severe cases, including alterations in right ventricular pressure and total pulmonary resistance [44]. Consequently, this molecule has been proposed as a potential therapeutic target in this pathological event [53]. PAF is one of the most widely evaluated mediators as a biomarker in anaphylaxis, although it has not been extrapolated to clinical practice. Studies carried out by Vadas et al [16,74] revealed that PAF increased with the severity of the reaction in the plasma of adults and children. Furthermore, a higher percentage of patients were diagnosed based on PAF than with other biomarkers, such as histamine and tryptase [16]. However, this mediator has a short half-life because it is

rapidly inactivated, thus hampering its application in clinical practice, since patients with anaphylaxis often attend the emergency department within minutes or hours of the onset of symptoms [74]. The main enzyme involved in degradation of PAF is PAF acetylhydrolase [75,76], which has also been proposed as a possible molecular marker for anaphylaxis, with various studies suggesting that its levels drop during the reaction [17,74,77].

Sphingosine-1-phosphate (S1P) is a bioactive lipid involved in a broad spectrum of cellular processes and is supplied to plasma from MCs, erythrocytes, platelets, and endothelial and smooth muscle cells [78]. It plays a key role in the signaling of vascular homeostasis and allergic disorders [68]. In addition, the balance between intracellular and extracellular S1P and their functional receptors attributes relevant features to several human inflammatory and allergic responses [79,80]. Intracellular levels of this molecule can be regulated by sphingosine kinases and are essential for maintaining endothelial barrier function [81]. In vivo systemic anaphylactic response has been associated with rapid depletion of circulating S1P concentrations [78,82].

Arachidonic acid-derived lipid molecules are synthesized in anaphylaxis and grouped into families including prostaglandins (PGE2, PGD2, PGF2a), prostacyclins (PGI2), cysteinylleukotrienes (LTC4, LTD4, and LTE4), and thromboxanes (TXA2) [83,84]. These molecules are derived from membrane bilayers through action of cyclooxygenase or the lipoxygenase pathway. Early studies in anaphylaxis observed the presence of slow-reacting substances with biologic activity after antigenic stimulation on sensitized tissues [85]. Years later, these substances are considered a blend of prostaglandins and cysteinyl-leukotrienes released mainly through MC activation [35,86]. In addition, some of the molecules are considered valuable biomarkers in anaphylaxis [84,87,88]. Interestingly, nonsteroidal anti-inflammatory drugs have been associated with increased PGE2 and the production and overexpression of adenosine receptor 3 [89,90].

2.1.1.4 Cytokines and chemokines

Cytokines are released by a wide variety of cells, although the main contributors are those of the immune system. These proteins present a diversity of functions, play a key role in immune responses, and are mainly involved in inflammatory processes [91,92]. In anaphylaxis, increased levels of cytokines have been observed in the most severe cases [17].

Tumor necrosis factor α (TNF- α) is one of the most studied cytokines. During anaphylaxis, it is released mainly by the activation and degranulation of MCs [93]. Indeed, increased levels of TNF- α have been described in experimental models and in serum samples of patients with anaphylaxis [54,94,95]. In turn, increased values of its receptor, TNF receptor I (TNFRI), have also been observed in most cases of severe anaphylaxis [17,54]. Accordingly, circulating levels of TNF-like weak inducer of apoptosis and of its receptor (Fn14) have been shown to increase in patients with anaphylaxis. These proteins play a crucial role in the alterations in vascular permeability underlying this pathological event [94].

Tumor growth factor β (TGF- β) is produced mainly by activated MCs and has an inhibitory effect on the anaphylactic

reaction by blocking the release of TNF- α [93,96]. TGF- β is increased in the serum of patients with anaphylaxis [97].

Interferon- γ (IFN- γ) is a proinflammatory cytokine that plays a key role in the T_H1 response [98]. In anaphylaxis, peripheral blood mononuclear cells from patients have been shown to release higher levels of this protein [99]. In addition, elevated values of IFN- γ have been observed in the serum of patients with anaphylaxis [54].

Granulocyte macrophage colony-stimulating factor (GM-CSF) is a hematopoietic growth factor that is increased in patients with anaphylaxis. GM-CSF modulates the maturation of several immune and inflammatory cells, including the main effector cells of the reaction [54]. In addition, it plays a role in inflammation by mediating communication between tissues and immune cells [100].

Platelet factor 4 is released by platelets following activation of the coagulation system [101], and its levels have been found to be increased in rabbit anaphylactic models [102].

Chemokines play a role in attracting and guiding various families of leukocytes [103], including CCL-2, also known as monocyte chemoattractant protein 1, which is increased in mice models and human anaphylactic samples [104-106]. In addition, a study conducted in serum from pediatric patients with anaphylaxis also demonstrated increased values for this chemokine and CCL-11 [107].

2.1.1.5 Interleukins

The term interleukin (IL) was coined to describe cytokines produced by leukocytes that play the role of messengers in different parts of the immune system [108]. However, these mediators can be produced by many other cell types (including resident cells) and exert pleiotropic effects [109,110].

Serum levels of IL-2, IL-4, IL-5, IL-6, IL-10, and IL-13 are increased in patients with severe anaphylaxis [54]. IL-2, IL-6, and IL-10 correlate with the severity of the episodes, histamine concentration, and hypotension [54,106,111]. Specifically, mouse anaphylactic models show that IL-6 is released within the first few hours after allergen exposure [112]. In humans, high levels of IL-6 have been observed in hypersensitivity reactions and anaphylaxis induced by monoclonal antibodies, as well as in perioperative patients [106,113]. IL-33 promotes oral anaphylaxis after epicutaneous sensitization in experimental murine models [114].

2.1.2 Complement and contact/coagulation systems

Complement and contact/coagulation systems are involved in the pathophysiology of anaphylaxis [61,115]. These molecular cascades are in constant crosstalk and in continuous amplification and inhibition loops, thus keeping their effect balanced during inflammation. For example, components of the coagulation system amplify complement activation, which, in turn, magnifies coagulation and inhibits fibrinolysis [116]. Factors and molecules derived from their proteolytic processes are altered in anaphylaxis, thus indicating not only a potential function of these systems in hypersensitivity reactions, but also the possibility of evaluating them as candidate diagnostic biomarkers.

2.1.2.1 Complement system

The complement system is composed of a complex cascade of proteases that participate in both the innate and the adaptive

immune responses through the activation of 3 distinguishable pathways: the classical, the alternative, and the lectin pathways [117].

In anaphylaxis, complement seems to be activated mostly by the classical pathway through the formation of IgG immunocomplexes. This process leads to the release of C3a, C4a, and C5a from the proteolysis of the C3, C4, and C5 components [117,118]. These soluble products, also known as anaphylatoxins, bind to specific G protein–coupled receptors in MCs and basophils and thereby stimulate their degranulation [119-121]. Indeed, levels of C3a, C4a, and C5a have been found to be elevated in the sera of patients with anaphylaxis and seem to be correlated with the severity of the reaction [17]. In addition, there is clinical evidence demonstrating the induction of this type of hypersensitivity reaction by agents that directly activate the complement system, which would confirm that anaphylatoxins alone would be capable of triggering the reaction [122].

2.1.2.2 Contact/coagulation systems

The contact system consists of a series of proteases and coagulation factors that are activated in response to inflammatory macromolecular complexes or negatively charged surfaces. In turn, it enables formation of bradykinin and the activation of the intrinsic coagulation cascade whose goal is clot formation [61]. Heparin is a relevant glycosaminoglycan released in anaphylaxis that exhibits a key function in the activation of the contact system after degranulation of MCs and basophils. In addition, non-IgEmediated immunologic anaphylactic reactions produced by the injection of heparin contaminated with oversulfated chondroitin sulfate have been described [123]. Given its negative charge, heparin is able to provoke the binding of high-molecular-weight kininogen (HK) to factor XII (FXII) of the contact system, inducing the autoactivation of FXII and the initiation of this cascade. Activated FXII induces the transformation of prekallikrein into kallikrein, a serine protease that hydrolyzes HK into bradykinin, one of the most potent vasoactive mediators involved during anaphylaxis [61,124-126].

The contact, coagulation, and fibrinolytic systems are intimately related because FXII activation in turn induces, via factor XI, the conversion of prothrombin to thrombin. This molecule is capable of transforming fibrinogen into insoluble fibrin, which, with the help of factor XIII, causes crosslinking of the fibrin polymers, thus creating clots [61].

A study of serum samples from patients with anaphylaxis mediated by insect stings showed that levels of factor V, factor VIII, and fibrinogen were decreased during the reaction [127]. On the other hand, regarding the contact system, other studies have shown decreased levels of FXII, prekallikrein, and HK, as well as elevated bradykinin values in sera from anaphylactic patients [128,129].

2.2 Cardiovascular System

The cardiovascular system is closely related to the pathophysiology of anaphylaxis [130,131]. A recent detailed review addresses the vascular disorders associated with this event, which is characterized by an acute increase in vascular permeability, relaxation of vascular smooth muscle, and constriction of vessels in the thoracic cavity [27].

Cardiovascular disease increases the risk of severe anaphylaxis, and activated heart MCs release mediators in the thoracic cavity [130]. Kounis syndrome, a coronary hypersensitivity disorder caused by massive activation of cardiac MCs and characterized by acute myocardial damage, is associated with anaphylaxis [132,133]. Specifically, an increase in troponins has been described in patients with this syndrome and anaphylaxis [134-136]. Troponins are part of the contractile apparatus of skeletal and cardiac muscle cells and biomarkers of myocardial damage and myocardial infarction [137].

2.2.1 Cells and processes

Endothelial cells (ECs) and vascular smooth muscle cells (VSMCs) interact with each other and govern the vascular pathophysiological processes of anaphylactic reactions [27,115].

ECs form a monolayer called the endothelium, which functions as the physical barrier between blood and tissues, thus regulating vascular permeability and blood pressure [115,138-140]. In anaphylaxis, release of mediators during the reaction opens the adherens junctions, giving rise to an increase in vascular permeability and stimulating fluid extravasation [58,115].

VSMCs are responsible for the regulation of vascular tone through their contractile properties. This process is modulated mainly by changes in intracellular calcium levels and membrane depolarization [27,141,142]. In anaphylaxis, mediators act on these cells, initiating relaxing or contractile processes in bronchial and vascular smooth muscle cells [115].

2.2.2 Mediators derived from vessels

In anaphylaxis, ECs function as both receptors and effector cells releasing relevant agents such as nitric oxide (NO) [58,143-145]. This vasodilator and anti-inflammatory molecule is crucial in vascular homeostasis [146]. Specifically, mediators such as histamine and PAF induce synthesis of NO by activating endothelial NO synthase [52,58,143]. NO plays a key role in the pathophysiology of anaphylaxis by increasing the vasodilatation underlying the reaction, thus triggering intracellular calcium depletion in VSMCs. In addition, NO has a hypotensive effect and decreases the catecholamine levels [27,147]. Likewise, fractional exhaled NO is increased in patients with anaphylaxis who experience respiratory symptoms [148].

Endothelin-1 is a molecule with vasoconstrictor capacity, since it induces an increase in the amount of intracellular calcium upon binding to its receptors on VSMCs [149]. Murine models have shown that this mediator can induce MC degranulation [150]. In addition, levels of endothelin-1 have been shown to be elevated in the trachea of guinea pigs with anaphylaxis [151].

Vascular endothelial growth factor (VEGF) is a molecule released by cells such as ECs, VSMCs, and MCs that induces vascular permeability [58,152]. Levels of this molecule have been found to be elevated in the serum of a patient with anaphylactic shock [153].

2.3 Nervous System

The nervous system comprises the central nervous system, which includes the brain and spinal cord, and the peripheral nervous system, consisting of afferent sensory and efferent motor nerves [154]. This system monitors and responds to changes in the internal and external environment [155]. Therefore, it is essential for maintaining body homeostasis and reacting to acute stressors [156,157].

Numerous studies have described the importance of the nervous system in the induction and coordination of the immune response [156,157]. In allergic processes, the nervous system participates in activities such as regulation of antigen presentation, IgE production, and MC degranulation [156]. These functions are due to close associations between autonomic nerves and immune cells, which can be activated through numerous neurotransmitters, and molecules released through their activation can also influence the neuronal activity [156].

In anaphylaxis, histamine acts through C- and A- γ fibers, stimulating the release of substance P and calcitonin gene–related peptide from the peripheral nerve endings [158]. These neuropeptides induce further activation of MCs through their specific surface receptors and intensify the inflammatory response, thus affecting arteriolar vasodilation and increasing blood flow [156]. In addition, tryptase can activate protease-activated receptor 2 on sensory neurons, thus stimulating the release of substance P and calcitonin gene–related peptide [158,159].

Furthermore, IgE- and antigen-mediated MC activation induces the release of serotonin [160], which contributes to clinical signs of anaphylaxis and is associated with the severity of the episode [161]. In mice models, serotonin is released in response to ovalbumin and acts on the parasympathetic neuronal fibers. This event leads to release of acetylcholine, which acts directly by inducing bronchoconstriction [162]. Moreover, activated human and mouse platelets contain high concentrations of serotonin and were able to induce significant hypothermia, probably through 5-hydroxytryptamine receptor 7 (5-HT7) [160,163].

Adenosine is a significant molecule that is released by the nervous system during anaphylaxis [164]. Adenosine produces vasodilatation, increases vascular permeability, and potentiates mediator release from stimulated MCs [164]. Additionally, adenosine may act on the nervous system, causing the release of tachykinins from sensory nerve endings in a similar manner to bradykinin [165].

Finally, clinical studies highlight the role of the nervous system in recovery from anaphylaxis [166,167]. Epinephrine, norepinephrine, and angiotensin II are elevated minutes after onset of an anaphylactic episode [167]. The vasopressor activity of these mediators enables them to compensate for the vasodilation and extravasation of liquids produced during the reaction [34].

2.4 Other Circulating Blood Mediators

Blood-derived circulating biomarkers have several advantages, as they reflect the individual's status, are easy to measure, and are obtained using minimally invasive methods [168]. Consequently, together with complement-, contact-, and coagulation-derived proteins, various circulating anaphylaxis-related molecules have been characterized and proposed as new molecular markers for this pathological event.

2.4.1 Circulating proteins

The blood contains many circulating proteins, among which human serum albumin stands out, since alone, it accounts for 55% of the total concentration [169]. A recent study of 112 patients with anaphylaxis showed that both serum protein concentration and human serum albumin levels decreased according to the severity of the reaction. Moreover, these measurements made it possible to indirectly quantify the extravasation underlying the event [25].

Oher circulating proteins have also been characterized in anaphylaxis. Apolipoprotein B (ApoB) was shown to inversely correlate with severity in a pediatric population of food-induced anaphylaxis [170]. Moreover, the decrease in serum apolipoprotein AI has been suggested as a promising biomarker for this pathological event [88]. In addition, a drop in this protein and apolipoprotein E levels has also been reported in the serum of anaphylactic mice models and in patients. However, apolipoprotein E has been shown to decrease in other inflammatory reactions; therefore, it could not identify anaphylaxis correctly [171]. Similarly, hemoglobin β subunit was found to be increased in 150 patients with mild hypersensitivity reactions and anaphylaxis [25].

2.4.2 MicroRNAs

microRNAs (miRNAs) are small noncoding RNA molecules (19-25 nucleotides) that govern the translation of messenger RNAs (mRNAs). Indeed, a single miRNA can modulate several mRNAs, and the same mRNA can be regulated by different miRNAs [172]. Thus, miRNAs participate in a wide range of physiological processes, such as proliferation, differentiation, and survival [173].

Alterations in circulating miRNA levels have been associated with a wide variety of diseases [174]. Furthermore, they may be associated with the individual's health status, making them noninvasive and promising biomarkers, although they are not yet used in clinical practice [173,175].

Diverse studies carried out in mouse models suggested the involvement of miRNAs (miR-155, miR-154-5p, miR-26a, miR-26b, miR-122a-5p, miR-135-5p, and miR-182-5p) in the allergic inflammatory processes underlying anaphylaxis [176-180]. In human samples, miRNA levels were shown to vary during this pathological event. One study in a pediatric population with food-induced anaphylaxis showed increased circulating serum levels of miR-21-3p and miR-487b-3p [181]. Other authors propose miR-451a as the most relevant biomarker in adult blood samples [88].

2.4.3 Extracellular vesicles

Extracellular vesicles (EVs) are lipid structures classified mainly according to their size and biogenesis into 3 subtypes: exosomes, microvesicles, and apoptotic bodies [182]. However, while they differ in terms of size, morphology, and protein composition, it remains difficult to distinguish between them [183].

EVs can be released by many cells under the physiological conditions controlling homeostasis [184]. They participate in cell–cell communication by regulating the interaction between various systems [183]. However, EVs are also released under pathological conditions [183,185,186]. Therefore, they have been proposed as a promising source of biomarkers, as they reflect the physiological or pathological state of cells. Moreover, they are very stable structures and are easy and inexpensive to obtain, since they can be isolated from many body fluids, such as blood and saliva [182,187].

The role of EVs in anaphylaxis is practically unknown. The only available study demonstrated the existence of a differential proteomic profile in EVs obtained during the anaphylactic reaction and revealed an increase in Ficolin-2, CDC42, and alarmin S100A9 levels [188].

2.4.4 Metabolites

In the last few decades, metabolomics has become increasingly important as a tool for biomarker discovery in various fields, such as allergy [189,190]. In anaphylaxis, a study carried out in guinea pigs demonstrated metabolic changes in challenged animals compared to controls [191]. Furthermore, several metabolites are generated from relevant known mediators participating in anaphylaxis, such as histamine. The rapid breakdown of this molecule gives rise to 2 metabolites (N-methylhistamine and N-methylimidazole octane), which are detectable in urine up to 24 hours after the onset of the reaction. However, although this sampling is less invasive than blood collection, its sensitivity and specificity is much lower [53,56,57]. In addition, given its stability in both urine and serum from anaphylactic patients, 9α , 11B-PGF2 appears to be a promising lipid-derived biomarker of anaphylaxis [84,87,88]. Moreover, a recently published exploratory study pointed to the relevance of other altered metabolites in anaphylaxis. Depending on the triggers or severity of the reaction, the differential molecules observed include glucose, lipids, cortisol, betaine, and oleamide [192].

3. Microenvironments

The relative contribution of mediators, cells, and systems to the pathophysiology of anaphylaxis is unknown. Furthermore, the location of effector cells and the half-life and abundance of the molecules released (local or systemic distribution) are likely to mask the recognition of reliable biomarkers. Identification of the exact mechanisms activated in these reactions would improve the diagnosis of patients based on precision medicine. Therefore, clarifying the various microenvironments established in anaphylaxis (eg, skin, lung, intestine, heart, vascular system, nervous system) will considerably enhance clinical management and recognition of better molecular markers.

3.1 Mast Cell Heterogeneity

Undoubtably, IgE-mediated anaphylactic reactions are the best characterized. However, the heterogeneity of

MCs is detrimental to finding a "universal" biomarker of anaphylaxis, and differences between humans and mice, and even between mice strains, indicate marked variations in IgE abundance, MC activity, and features of the mediators released [193,194]. Specifically, the degranulation profile of MCs, far from being simple, is not unique and uniform. Different types of stimuli and triggers induce diverse patterns of exocytosis, releasing various types of mediators to the surroundings [31]. MCs induce distinct lipid, cytokine, and chemokine secretion profiles, as well as diverse degranulation dynamics, depending on FceRI cross-linking or other G protein-coupled receptor activation [30,195]. Such is the case of gastrointestinal symptoms, where the main MC mediators involved are PAF and serotonin, rather than histamine [196,197]. Moreover, the wide variety of MC receptors activating specific cell signalling has reached considerable relevance and may even have an impact on the diversity of organs affected in anaphylaxis [198,199]. For instance, A3 is increased 3-fold in lung MCs compared to skin MCs [90,200]. Therefore, different anaphylactic microenvironments would be created depending on the nature of each reaction. Consequently, the release of specific mediators derived from specific activated cells would explain the differences in the symptoms characterizing this pathological event.

3.2 The Vascular Wall and Homeostatic Factors

The endothelium plays a key role in the development of the reaction owing to its direct contact with blood. At the onset of anaphylaxis, many mediators released by the effector cells induce a direct effect on ECs. Among them, molecules such as histamine, cathepsin G, and PAF can produce vascular dilation and increase endothelial permeability [34,52,53,58,73,201].

On the other hand, products derived from the activation of the different blood systems are also capable of exerting an effect on the vascular wall. This can occur indirectly, for example, via complement-derived anaphylatoxins by inducing degranulation of effector cells [202,203], or directly, for example, via bradykinin, a potent vasoactive agent capable of inducing vasodilation and increasing vascular permeability [44,124]. In addition, a multitude of mediators released by effector cells, including the most relevant molecules (sT, chymase, histamine, and PAF), appear to be involved in the regulation of the complement, coagulation, and contact systems. Specifically, proteases play an important role by amplifying inflammation and activating the molecular cascades of complement and coagulation. For instance, chymase has been related to the coagulation pathway [48,56,61]. Similarly, sT and heparin can activate complement through an independent loop of the system's own convertases, thus



Figure 2. Relevant proteins identified in in vitro endothelial cells stimulated with serum from anaphylactic patients. Adapted from [115]. The complement, coagulation, and fibrinolytic systems stand out. Increased proteins are marked in red; decreased proteins are shown in blue. tPA indicates tissue plasminogen activator; uPA, urokinase; CXCL7, chemokine (C-X-C motif) ligand 7; HRG, histidine-rich glycoprotein; CPB2, carboxypeptidase B2; TSP1, thrombospondin-1; MMRN1, multimerin 1; THRB, thrombin; LG3BP, galectin-3-binding protein; FINC, fibronectin; APOH, apolipoprotein H; iPROC, inactive protein C; aPROC, active protein C; PROS1, protein S1; FV, VII, VIII, X, X, XI, aXI, factor V, VII, VIII, IX, X, XI, active XI; C1qC1sC1r, complement components 1q, 1s, 1r complex; C2, 2a, 2b, 3, 3a, 3b, 4, 4a, 4b, 5, 5a, 5b, complement components 2, 2a, 2b, 3, 3a, 3b, 4, 4a, 4b, 5, 5a, 5b, complement components 3b, 2b complex; C4Dp, complement components 4b binding protein; C6CC7C8C9, complement components 6c, 7, 8, 9 complex; VTNC, vitronectin; CFI, complement factor I; CFH, complement factor H; DAF, complement decay-accelerating factor.

inducing the release of C3a and C5a [53,204]. In turn, the complement system can amplify coagulation and inhibit fibrinolysis. Reciprocally, the coagulation system has the capacity to activate the complement system [116].

An in vitro study of ECs stimulated with sera from anaphylactic patients highlights the relevance of platelets and coagulation processes in anaphylaxis and shows alterations in proteins related not only to this system, but also to the complement and fibrinolytic systems (Figure 2). Most of the increased proteins identified are related to the coagulation process, although it is noteworthy that, among them, the FIX and FX proteins had previously been observed to be decreased in patients' sera, indicating that the endothelium is actively participating in the consumption of factors derived from these systems during the course of the reaction and thus explaining the decrease in these factors in serum at the cellular level [115].

3.3 Amplification of the Molecular Loops of Interleukins

Inflammation in anaphylaxis is partially due to the contribution of interleukins to the reactions. These molecules not only mediate key immune processes, but also lead activation to the resident cells, setting up positive and negative feedback loops (Figure 3).

IL-4 is highly relevant in the development of T_{H2} lymphocytes and the induction of IgE class-switching in B cells [205]. In turn, IL-13 activates IL-4 signalling pathways, induces IgE production, and activates recruitment of MCs and eosinophils, thus promoting their survival [205]. Both IL-4 and IL-13 exacerbate anaphylaxis through activation of a STAT 6-dependent pathway in murine models, increasing the sensitivity of target cells to vasoactive mediators [206]. In addition, both molecules could activate a phosphatidyl inositol-3 kinase (PI3K)-dependent pathway to induce expression of endothelial NO synthase and overproduction of NO [197]. Most of these processes lead to vascular permeability, vasodilation, and hypotension, thus influencing the severity of the episode. In addition, the fact that IL-4 and IL-13 shared the α chain of their IL-4 receptor (IL-4R α) has led to the use of treatment with anti-IL-4Ra, which is considered a relevant therapeutic tool in anaphylaxis [207]. It has been shown that IL-4 and IL-13 can also enhance the response to other anaphylactic mediators in human MCs and macrophages by increasing the expression of their receptors, such as cysteinyl-leukotriene receptors [208].

IL-33 acts directly on IgE class-switching in B lymphocytes by inducing IL-4 and promoting MC degranulation [209]. Importantly, interleukins could amplify other signalling pathways; for example, IL-33 induces release of VEGF from MCs, increasing vascular permeability and contributing

IL-18/IL-12





INFv

to inflammation [210]. Likewise, the effect of IL-6 can be mediated by its binding to IL-6Rs present on the surface of various cells or in its soluble form (sIL-6R) [205]. IL-6 is also able to induce ECs to synthesize several factors and proteins of the complement system [211,212]. Finally, in vitro approaches show the effect of IL-3 in basophils by increasing expression of the activation markers CD69 and CD203c and enhancing mediator release in response to FccR cross-linking [213,214].

In animal models, IL-18 has been shown to play a role in the initiation of T_{H2} responses by inducing IL-4 and IL-13 synthesis in MCs and basophils, leading to an increase in IgE production [215,216]. Conversely, coadministration of IL-18 and IL-12 induces the production of IFN- γ , stimulates T_{H1} -mediated immune responses, and inhibits IgE synthesis [216,217]. Therefore, the administration of these molecules in combination could be considered a treatment for severe allergic disorders [217].

On the other hand, IL-10 modulates MHCII and expression of costimulatory molecules on monocytes/ macrophages and dendritic cells, thus limiting the production of proinflammatory cytokines and chemokines [218-220]. Therefore, this mediator participates in the resolution of the systemic immune response. Specifically, peripheral T-cell tolerance in immunotherapy is mediated by upregulation of IL-10 by allergen-specific regulatory T cells [221]. In mouse models, the introduction of bioengineered microorganisms to deliver this protein to the intestine decreases food-induced anaphylaxis and prevents IgE-type sensitization to common food allergens [222]. IL-12 exhibits a similar function to that of IL-10. Its oral administration prevents and reverses peanut hypersensitivity. Furthermore, it was shown to reduce the release of histamine, specific IgE, and IgG1 in a mouse model of peanut anaphylaxis [223].

3.4 Communication Between Microenvironments: miRNAs and EVs

The data provided above demonstrate the importance of the different microenvironments in anaphylaxis. However, these are not isolated niches, since there is communication between them. Therefore, several molecules have been postulated as participants in the interaction between these microenvironments, including miRNAs and EVs, which have been described as key players in cell–cell communication during the anaphylactic reaction [181,188] (Figure 4).



Figure 4. miRNAs and extracellular vesicles (EVs) described in anaphylaxis. These molecules are involved in degranulation and vascular permeability (left) and in pro/anti-inflammatory effects (right). MC indicates mast cell; MON, monocyte; N, neutrophil; PI3K, phosphatidylinositol 3-kinases; COX-2, cyclooxygenase 2; SOCS1; suppressor of cytokine signaling 1; JAK2; Janus kinase 2; TNFR, tumor necrosis factor receptor; TGF- β , transforming growth factor β .

miR-155 regulates degranulation of MCs by modulating their calcium concentration through PI3K levels [179]. In addition, miR-21-3p could participate in this process by its action on signaling of Rac, a protein involved in MC degranulation [181,224,225]. In turn, analysis of EVs obtained during anaphylaxis revealed that the main function of their differential proteins was neutrophil degranulation [188]. Specifically, elevation of circulating levels of miR-451a has been associated with increased degranulation of the effector cells in the reaction [88].

Nevertheless, EVs obtained during anaphylaxis have also been reported to induce an increase in endothelial permeability in vitro [188]. In addition, they can act as transporters of various miRNAs, which exert their action on the target cell [226]. Two mi-RNAs have been characterized in anaphylaxis, namely, miR-135-5p and miR-154-5p, which can regulate allergic inflammation in a p62 protein–dependent manner [179,227].

Other miRNAs play a role in the regulation of inflammation during anaphylaxis. For instance, miR-26a and miR-26b regulate levels of COX-2, a key enzyme in this process and in the release of the cytokines involved in the reaction [177]. Conversely, miR-182-5p has been reported to induce an antiinflammatory effect, as it reduces the levels of IL-27, a cytokine involved in allergic inflammation [180]. Similarly, miR-122 was also found to participate in the inhibition of this process through the regulation of suppressor of cytokine signaling 1 and Janus kinase 2, both of which control the production of the anti-inflammatory mediator TGF-B [178]. In turn, miR-487b-3p modulates the inflammation underlying the reaction, as it participates in signaling of Janus kinase 2 and IL-9, a proinflammatory cytokine released by MCs [181]. Likewise, miR-21-3p regulates this process through its role in TNFR signaling, on which a wide variety of cytokines act [181].

4. Conclusions

Various molecules have been identified and proposed as biomarkers in anaphylaxis, although none of them has yet been extrapolated to clinical practice, probably because of the lack of studies connecting the clinical manifestations of the patient with the molecular mechanisms underlying the reaction. Accordingly, the recent increase in the understanding of the signaling pathways involved in anaphylaxis has led to new candidate molecular markers. Therefore, a more robust knowledge of the plethora of anaphylactic phenotypes and endotypes would lead to the identification and characterization of better diagnostic and predictive biomarkers that would in turn improve the clinical management of patients and their quality of life.

Acknowledgments

The figures were created with Biorender.com

Funding

This research was supported by grants from the Instituto de Salud Carlos III (PI18/00348, PI21/00158) and FEDER

Thematic Networks and Cooperative Research Centers RETICS ARADyAL (RD16/0006/0013). This work was also sustained by the SEAIC (19_A08) and Alfonso X el Sabio University Foundations. EN-B was funded from FOOD-AL (CM_P2018/BAAA-4574).

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- 1. Bilò MB, Martini M, Tontini C, Corsi A, Antonicelli L. Anaphylaxis. Eur Ann Allergy Clin Immunol. 2021;53(1):4-17.
- 2. Aronson JK, Ferner RE. Biomarkers-A General Review. Curr Protoc Pharmacol. 2017;76:9.23.1-9.23.17.
- Dribin TE, Schnadower D, Wang J, Camargo CA, Michelson KA, Shaker M, et al. Anaphylaxis knowledge gaps and future research priorities: A consensus report. J Allergy Clin Immunol. 2022;149(3):999-1009.
- Cardona V, Ansotegui IJ, Ebisawa M, El-Gamal Y, Fernandez Rivas M, Fineman S, et al. World allergy organization anaphylaxis guidance 2020. World Allergy Organ J. 202030;13(10):100472.
- Simons FER, Ardusso LRF, Bilò MB, El-Gamal YM, Ledford DK, Ring J, et al. World allergy organization guidelines for the assessment and management of anaphylaxis. World Allergy Organ J. 2011;4(2):13-37.
- Yu JE, Lin RY. The Epidemiology of Anaphylaxis. Clin Rev Allergy Immunol. 2018;54(3):366-74.
- Regateiro FS, Marques ML, Gomes ER. Drug-Induced Anaphylaxis: An Update on Epidemiology and Risk Factors. Int Arch Allergy Immunol. 2020;181(7):481-7.
- Li PH, Leung ASY, Li RMY, Leung TF, Lau CS, Wong GWK. Increasing incidence of anaphylaxis in Hong Kong from 2009 to 2019-discrepancies of anaphylaxis care between adult and paediatric patients. Clin Transl Allergy. 2020;10(1):51.
- Turner PJ, Gowland MH, Sharma V, lerodiakonou D, Harper N, Garcez T, et al. Increase in anaphylaxis-related hospitalizations but no increase in fatalities: an analysis of United Kingdom national anaphylaxis data, 1992-2012. J Allergy Clin Immunol. 2015;135(4):956-63.e1.
- Michelson KA, Dribin TE, Vyles D, Neuman MI. Trends in emergency care for anaphylaxis. J Allergy Clin Immunol Pract. 2020;8(2):767-8.e2.
- Tejedor-Alonso MA, Moro-Moro M, Múgica-García MV. Epidemiology of Anaphylaxis: Contributions From the Last 10 Years. J Investig Allergol Clin Immunol. 2015;25(3):163-75.
- Turner PJ, Worm M, Ansotegui IJ, El-Gamal Y, Rivas MF, Fineman S, et al. Time to revisit the definition and clinical criteria for anaphylaxis? World Allergy Organ J. 2019;12(10):100066.
- van der Linden PW, Hack CE, Poortman J, Vivié-Kipp YC, Struyvenberg A, van der Zwan JK. Insect-sting challenge in 138 patients: relation between clinical severity of anaphylaxis and mast cell activation. J Allergy Clin Immunol. 1992;90(1):110-8.
- De Schryver S, Halbrich M, Clarke A, La Vieille S, Eisman H, Alizadehfar R, et al. Tryptase levels in children presenting with anaphylaxis: Temporal trends and associated factors. J Allergy Clin Immunol. 2016;137(4):1138-42.

- Vitte J, Amadei L, Gouitaa M, Mezouar S, Zieleskiewicz L, Albanese J, et al. Paired acute-baseline serum tryptase levels in perioperative anaphylaxis: An observational study. Allergy. 2019;74(6):1157-65.
- 16. Vadas P, Perelman B, Liss G. Platelet-activating factor, histamine, and tryptase levels in human anaphylaxis. Journal of Allergy and Clinical Immunology. 2013;131(1):144-9.
- Brown SGA, Stone SF, Fatovich DM, Burrows SA, Holdgate A, CelenzaA, et al. Anaphylaxis: clinical patterns, mediator release, and severity. J Allergy Clin Immunol. 2013;132(5):1141-9.e5.
- Montañez MI, Mayorga C, Bogas G, Barrionuevo E, Fernandez-Santamaria R, Martin-Serrano A, et al. Epidemiology, Mechanisms, and Diagnosis of Drug-Induced Anaphylaxis. Front Immunol. 2017;8:614.
- 19. Passia E, Jandus P. Using Baseline and Peak Serum Tryptase Levels to Diagnose Anaphylaxis: a Review. Clin Rev Allergy Immunol. 2020;58(3):366-76.
- Sala-Cunill A, Cardona V, Labrador-Horrillo M, Luengo O, Esteso O, Garriga T, et al. Usefulness and limitations of sequential serum tryptase for the diagnosis of anaphylaxis in 102 patients. Int Arch Allergy Immunol. 2013;160(2):192-9.
- Lobbes H, Reynaud Q, Mainbourg S, Lega JC, Durieu I, Durupt S. [Tryptase: A practical guide for the physician]. Rev Med Interne. 2020;41(11):748-55.
- Borer-Reinhold M, Haeberli G, Bitzenhofer M, Jandus P, Hausmann O, Fricker M, et al. An increase in serum tryptase even below 11.4 ng/mL may indicate a mast cell-mediated hypersensitivity reaction: a prospective study in Hymenoptera venom allergic patients. Clin Exp Allergy. 2011;41(12):1777-83.
- Wongkaewpothong P, Pacharn P, Sripramong C, Boonchoo S, Piboonpocanun S, Visitsunthorn N, et al. The utility of serum tryptase in the diagnosis of food-induced anaphylaxis. Allergy Asthma Immunol Res. 2014;6(4):304-9.
- Valent P, Akin C, Arock M, Brockow K, Butterfield JH, Carter MC, et al. Definitions, criteria and global classification of mast cell disorders with special reference to mast cell activation syndromes: a consensus proposal. Int Arch Allergy Immunol. 2012;157(3):215-25.
- Nuñez-Borque E, Betancor D, Pastor-Vargas C, Fernández-Bravo S, Martin-Blazquez A, Casado-Navarro N, et al. Personalized diagnostic approach and indirect quantification of extravasation in human anaphylaxis. Allergy. 2022.
- Jimenez-Rodriguez TW, Garcia-Neuer M, Alenazy LA, Castells M. Anaphylaxis in the 21st century: phenotypes, endotypes, and biomarkers. J Asthma Allergy. 2018;11:121-42.
- Nuñez-Borque E, Fernandez-Bravo S, Yuste-Montalvo A, Esteban V. Pathophysiological, Cellular, and Molecular Events of the Vascular System in Anaphylaxis. Front Immunol. 2022;13:836222.
- Muñoz-Cano R, Pascal M, Araujo G, Goikoetxea MJ, Valero AL, Picado C, et al. Mechanisms, Cofactors, and Augmenting Factors Involved in Anaphylaxis. Front Immunol. 2017;8:1193.
- 29. Pastor-Vargas C, Esteban V. Editorial: New Insights In Anaphylaxis. Front Immunol. 2018;9:506.
- Gaudenzio N, Sibilano R, Marichal T, Starkl P, Reber LL, Cenac N, et al. Different activation signals induce distinct mast cell degranulation strategies. J Clin Invest. 2016;126(10):3981-98.

- Huber M, Cato ACB, Ainooson GK, Freichel M, Tsvilovskyy V, Jessberger R, et al. Regulation of the pleiotropic effects of tissue-resident mast cells. J Allergy Clin Immunol. 2019;144(4S):S31-45.
- 32. Yoo Y, Perzanowski MS. Allergic sensitization and the environment: latest update. Curr Allergy Asthma Rep. 2014;14(10):465.
- 33. Poulsen LK, Hummelshoj L. Triggers of IgE class switching and allergy development. Ann Med. 2007;39(6):440-56.
- 34. Reber LL, Hernandez JD, Galli SJ. The pathophysiology of anaphylaxis. J Allergy Clin Immunol. 2017;140(2):335-48.
- Peavy RD, Metcalfe DD. Understanding the mechanisms of anaphylaxis. Curr Opin Allergy Clin Immunol. 2008;8(4):310-5.
- 36. Simons FER, Frew AJ, Ansotegui IJ, Bochner BS, Golden DBK, Finkelman FD, et al. Risk assessment in anaphylaxis: current and future approaches. J Allergy Clin Immunol. 2007;120:S2-24.
- 37. Hoffman DR. Fatal reactions to hymenoptera stings. Allergy Asthma Proc. 2003;24(2):123-7.
- Golden DB, Marsh DG, Freidhoff LR, Kwiterovich KA, Addison B, Kagey-Sobotka A, et al. Natural history of Hymenoptera venom sensitivity in adults. J Allergy Clin Immunol. 1997;100(6 Pt 1):760-6.
- Finkelman FD, Rothenberg ME, Brandt EB, Morris SC, Strait RT. Molecular mechanisms of anaphylaxis: lessons from studies with murine models. J Allergy Clin Immunol. 2005;115(3):449-57; quiz 458.
- Strait RT, Morris SC, Finkelman FD. IgG-blocking antibodies inhibit IgE-mediated anaphylaxis in vivo through both antigen interception and Fc gamma RIIb cross-linking. J Clin Invest. 2006;116(3):833-41.
- 41. Bruhns P, Chollet-Martin S. Mechanisms of human drug-induced anaphylaxis. J Allergy Clin Immunol. 2021;147(4):1133-42.
- Jönsson F, Mancardi DA, Kita Y, Karasuyama H, Iannascoli B, Van Rooijen N, et al. Mouse and human neutrophils induce anaphylaxis. J Clin Invest. 2011;121(4):1484-96.
- Jönsson F, Chaisemartin L de, Granger V, Gouel-Chéron A, Gillis CM, Zhu Q, et al. An IgG-induced neutrophil activation pathway contributes to human drug-induced anaphylaxis. Sci Transl Med. 2019;11(500):eaat1479.
- 44. Cianferoni A. Non-IgE-mediated anaphylaxis. J Allergy Clin Immunol. 2021;147(4):1123-31.
- Guilliams M, Bruhns P, Saeys Y, Hammad H, Lambrecht BN. The function of Fcγ receptors in dendritic cells and macrophages. Nat Rev Immunol. 2014;14(2):94-108.
- 46. Qiao J, Al-Tamimi M, Baker RI, Andrews RK, Gardiner EE. The platelet Fc receptor, FcγRIIa. Immunol Rev. 2015;268(1):241-52.
- Francis A, Bosio E, Stone SF, Fatovich DM, Arendts G, Nagree Y, et al. Neutrophil activation during acute human anaphylaxis: analysis of MPO and sCD62L. Clin Exp Allergy. 2017;47(3):361-70.
- 48. Ogawa Y, Grant JA. Mediators of anaphylaxis. Immunol Allergy Clin North Am. 2007;27(2):249-60.
- 49. Jutel M, Akdis M, Akdis CA. Histamine, histamine receptors and their role in immune pathology. Clin Exp Allergy. 2009;39(12):1786-800.
- 50. Lieberman P. The basics of histamine biology. Ann Allergy Asthma Immunol. 2011;106:S2-5.

- 51. MacGlashan D. Histamine: A mediator of inflammation. J Allergy Clin Immunol. 2003;112:S53-9.
- 52. Brown SGA. The pathophysiology of shock in anaphylaxis. Immunol Allergy Clin North Am. 2007;27(2):165-75.
- Tomasiak-Łozowska MM, Klimek M, Lis A, Moniuszko M, Bodzenta-Łukaszyk A. Markers of anaphylaxis - a systematic review. Adv Med Sci. 2018;63(2):265-77.
- Stone SF, Cotterell C, Isbister GK, Holdgate A, Brown SGA, Emergency Department Anaphylaxis Investigators. Elevated serum cytokines during human anaphylaxis: Identification of potential mediators of acute allergic reactions. J Allergy Clin Immunol. 2009;124(4):786-92.e4.
- Lin RY, Schwartz LB, Curry A, Pesola GR, Knight RJ, Lee HS, et al. Histamine and tryptase levels in patients with acute allergic reactions: An emergency department-based study. J Allergy Clin Immunol. 2000;106(1 Pt 1):65-71.
- 56. Kabashima K, Nakashima C, Nonomura Y, Otsuka A, Cardamone C, Parente R, et al. Biomarkers for evaluation of mast cell and basophil activation. Immunol Rev. 2018;282(1):114-20.
- 57. Sala-Cunill A, Guilarte M, Cardona V. Phenotypes, endotypes and biomarkers in anaphylaxis: current insights. Curr Opin Allergy Clin Immunol. 2018;18(5):370-6.
- Nguyen SMT, Rupprecht CP, Haque A, Pattanaik D, Yusin J, Krishnaswamy G. Mechanisms Governing Anaphylaxis: Inflammatory Cells, Mediators, Endothelial Gap Junctions and Beyond. Int J Mol Sci. 2021;22(15):7785.
- 59. Simons FER. Anaphylaxis. J Allergy Clin Immunol. 2010;125: S161-81.
- Guo XJ, Wang YY, Zhang HY, Jin QQ, Gao CR. Mast cell tryptase and carboxypeptidase A expression in body fluid and gastrointestinal tract associated with drug-related fatal anaphylaxis. World J Gastroenterol. 2015;21(47):13288-93.
- 61. Guilarte M, Sala-Cunill A, Luengo O, Labrador-Horrillo M, Cardona V. The Mast Cell, Contact, and Coagulation System Connection in Anaphylaxis. Front Immunol. 2017;8:846.
- 62. Sala-Cunill A, Cardona V. Biomarkers of anaphylaxis, beyond tryptase. Curr Opin Allergy Clin Immunol. 2015;15(4):329-36.
- 63. Nishio H, Takai S, Miyazaki M, Horiuchi H, Osawa M, Uemura K, et al. Usefulness of serum mast cell-specific chymase levels for postmortem diagnosis of anaphylaxis. Int J Legal Med. 2005;119(6):331-4.
- 64. Laghlam D, Jozwiak M, Nguyen LS. Renin-Angiotensin-Aldosterone System and Immunomodulation: A State-of-the-Art Review. Cells. 2021;10(7):1767.
- Hermann K, Ring J. Association between the renin angiotensin system and anaphylaxis. Adv Exp Med Biol. 1995;377:299-309.
- Meier HL, Heck LW, Schulman ES, MacGlashan DW. Purified human mast cells and basophils release human elastase and cathepsin G by an IgE-mediated mechanism. Int Arch Allergy Appl Immunol. 1985;77(1-2):179-83.
- Burster T, Gärtner F, Knippschild U, Zhanapiya A. Activity-Based Probes to Utilize the Proteolytic Activity of Cathepsin G in Biological Samples. Front Chem. 2021;9:628295.
- Díaz-Perales A, Escribese MM, Garrido-Arandia M, Obeso D, Izquierdo-Alvarez E, Tome-Amat J, et al. The Role of Sphingolipids in Allergic Disorders. Front Allergy. 2021;2:675557.
- 69. Kulinski JM, Muñoz-Cano R, Olivera A. Sphingosine-1phosphate and other lipid mediators generated by mast

J Investig Allergol Clin Immunol 2022; Vol. 32(6): 419-437 doi: 10.18176/jiaci.0854 cells as critical players in allergy and mast cell function. Eur J Pharmacol. 20165;778:56-67.

- Montrucchio G, Alloatti G, Camussi G. Role of plateletactivating factor in cardiovascular pathophysiology. Physiol Rev. 2000;80(4):1669-99.
- 71. Burks AW. Factoring PAF in anaphylaxis. N Engl J Med. 2008;358(1):79-81.
- Braquet P, Paubert-Braquet M, Koltai M, Bourgain R, Bussolino F, Hosford D. Is there a case for PAF antagonists in the treatment of ischemic states? Trends Pharmacol Sci. 1989;10(1):23-30.
- Gill P, Jindal NL, Jagdis A, Vadas P. Platelets in the immune response: Revisiting platelet-activating factor in anaphylaxis. J Allergy Clin Immunol. 2015;135(6):1424-32.
- 74. Vadas P, Gold M, Perelman B, Liss GM, Lack G, Blyth T, et al. Platelet-activating factor, PAF acetylhydrolase, and severe anaphylaxis. N Engl J Med. 2008;358(1):28-35.
- 75. Farr RS, Cox CP, Wardlow ML, Jorgensen R. Preliminary studies of an acid-labile factor (ALF) in human sera that inactivates platelet-activating factor (PAF). Clin Immunol Immunopathol. 1980;15(3):318-30.
- Upton JEM, Hoang JA, Leon-Ponte M, Finkelstein Y, Du YJ, Adeli K, et al. Platelet-activating factor acetylhydrolase is a biomarker of severe anaphylaxis in children. Allergy. 2022;77(9):2665-76.
- Piwowarek KŁ, Rzeszotarska A, Korsak JŁ, Juszkiewicz A, Chciałowski A, Kruszewski J. Clinical significance of plasma PAF acetylhydrolase activity measurements as a biomarker of anaphylaxis: Cross-sectional study. PLoS One. 2021;16(8):e0256168.
- Gazit SL, Mariko B, Thérond P, Decouture B, Xiong Y, Couty L, et al. Platelet and Erythrocyte Sources of S1P Are Redundant for Vascular Development and Homeostasis, but Both Rendered Essential After Plasma S1P Depletion in Anaphylactic Shock. Circ Res. 2016;119(8):e110-26.
- 79. Takabe K, Paugh SW, Milstien S, Spiegel S. "Inside-out" signaling of sphingosine-1-phosphate: therapeutic targets. Pharmacol Rev. 2008;60(2):181-95.
- Oskeritzian CA, Price MM, Hait NC, Kapitonov D, Falanga YT, Morales JK, et al. Essential roles of sphingosine-1-phosphate receptor 2 in human mast cell activation, anaphylaxis, and pulmonary edema. J Exp Med. 2010;207(3):465-74.
- 81. Wilkerson BA, Argraves KM. The role of sphingosine-1phosphate in endothelial barrier function. Biochim Biophys Acta. 2014;1841(10):1403-12.
- Olivera A, Mizugishi K, Tikhonova A, Ciaccia L, Odom S, Proia RL, et al. The sphingosine kinase-sphingosine-1-phosphate axis is a determinant of mast cell function and anaphylaxis. Immunity. 2007;26(3):287-97.
- 83. Funk CD. Prostaglandins and leukotrienes: advances in eicosanoid biology. Science. 2001;294(5548):1871-5.
- Nassiri M, Eckermann O, Babina M, Edenharter G, Worm M. Serum levels of 9α,11β-PGF2 and cysteinyl leukotrienes are useful biomarkers of anaphylaxis. J Allergy Clin Immunol. 2016;137(1):312-4.e7.
- Orange RP, Murphy RC, Karnovsky ML, Austen KF. The physicochemical characteristics and purification of slow-reacting substance of anaphylaxis. J Immunol. 1973;110(3):760-70.

- Denzlinger C, Haberl C, Wilmanns W. Cysteinyl leukotriene production in anaphylactic reactions. Int Arch Allergy Immunol. 1995;108(2):158-64.
- Ono E, Taniguchi M, Mita H, Fukutomi Y, Higashi N, Miyazaki E, et al. Increased production of cysteinyl leukotrienes and prostaglandin D2 during human anaphylaxis. Clin Exp Allergy. 2009;39(1):72-80.
- Francuzik W, Pažur K, Dalke M, Dölle-Bierke S, Babina M, Worm M. Serological profiling reveals hsa-miR-451a as a possible biomarker of anaphylaxis. JCI Insight. 2022;7(7):e156669.
- Muñoz-Cano R, Pascal M, Bartra J, Picado C, Valero A, Kim DK, et al. Distinct transcriptome profiles differentiate nonsteroidal anti-inflammatory drug-dependent from nonsteroidal antiinflammatory drug-independent food-induced anaphylaxis. J Allergy Clin Immunol. 2016;137(1):137-46.
- 90. Muñoz-Cano R, San Bartolome C, Casas-Saucedo R, Araujo G, Gelis S, Ruano-Zaragoza M, et al. Immune-Mediated Mechanisms in Cofactor-Dependent Food Allergy and Anaphylaxis: Effect of Cofactors in Basophils and Mast Cells. Front Immunol. 2021;11:623071.
- Borish LC, Steinke JW. 2. Cytokines and chemokines. J Allergy Clin Immunol. 2003;111:S460-75.
- Commins SP, Borish L, Steinke JW. Immunologic messenger molecules: cytokines, interferons, and chemokines. J Allergy Clin Immunol. 2010;125:S53-72.
- Kendall JC, Li XH, Galli SJ, Gordon JR. Promotion of mouse fibroblast proliferation by IgE-dependent activation of mouse mast cells: role for mast cell tumor necrosis factor-alpha and transforming growth factor-beta 1. J Allergy Clin Immunol. 1997;99:113-23.
- 94. Mendez-Barbero N, Yuste-Montalvo A, Nuñez-Borque E, Jensen BM, Gutiérrez-Muñoz C, Tome-Amat J, et al. The TNF-like weak inducer of the apoptosis/fibroblast growth factor-inducible molecule 14 axis mediates histamine and platelet-activating factor-induced subcutaneous vascular leakage and anaphylactic shock. J Allergy Clin Immunol. 2020;145(2):583-96.e6.
- 95. Gordon JR, Galli SJ. Release of both preformed and newly synthesized tumor necrosis factor alpha (TNF-alpha)/cachectin by mouse mast cells stimulated via the Fc epsilon RI. A mechanism for the sustained action of mast cell-derived TNFalpha during IgE-dependent biological responses. J Exp Med. 1991;174(1):103-7.
- 96. Kim HM, Lee YM. Role of TGF-beta 1 on the IgE-dependent anaphylaxis reaction. J Immunol. 1999;162(8):4960-5.
- Ma S, Yin J. Imbalance of serum IL-10 and TGF-β in patients with pollen food syndrome. Allergol Immunopathol (Madr). 2014;42(3):198-205.
- Kak G, Raza M, Tiwari BK. Interferon-gamma (IFN-γ): Exploring its implications in infectious diseases. Biomol Concepts. 2018;9(1):64-79.
- Kubota Y, Koga T, Nakayama J. In vitro released interferongamma in the diagnosis of drug-induced anaphylaxis. Eur J Dermatol. 1999;9(7):559-60.
- Becher B, Tugues S, Greter M. GM-CSF: From Growth Factor to Central Mediator of Tissue Inflammation. Immunity. 2016;45(5):963-73.
- Eisman R, Surrey S, Ramachandran B, Schwartz E, Poncz M. Structural and functional comparison of the genes for human platelet factor 4 and PF4alt. Blood. 1990;76(2):336-44.

- 102. McManus LM, Morley CA, Levine SP, Pinckard RN. Platelet activating factor (PAF) induced release of platelet factor 4 (PF4) in vitro and during IgE anaphylaxis in the rabbit. J Immunol. 1979;123(6):2835-41.
- 103. Zlotnik A, Yoshie O. The chemokine superfamily revisited. Immunity. 2012;36(5):705-16.
- Korosec P, Turner PJ, Silar M, Kopac P, Kosnik M, Gibbs BF, et al. Basophils, high-affinity IgE receptors, and CCL2 in human anaphylaxis. J Allergy Clin Immunol. 2017;140(3):750-8.e15.
- 105. Vantur R, Rihar M, Koren A, Rijavec M, Kopac P, Bidovec-Stojkovic U, et al. Chemokines during anaphylaxis: the importance of CCL2 and CCL2-dependent chemotactic activity for basophils. Clin Transl Allergy. 2020;10(1):63.
- 106. Callesen KT, Poulsen LK, Garvey LH, Jensen BM. Comparing baseline and reaction samples of perioperative anaphylaxis patients reveals IL-6 and CCL2 as potential biomarkers. Clin Exp Allergy. 2021;51(9):1250-3.
- 107. Radman M, Hassanshahi G, Vazirinejad R, Arababadi MK, Karimabad MN, Khorramdelazad H, et al. Serum levels of the CC chemokines CCL2, CCL5, and CCL11 in food allergic children with different clinical manifestations. Inflammation. 2013;36(3):561-6.
- 108. Kaiser P, Rothwell L, Avery S, Balu S. Evolution of the interleukins. Dev Comp Immunol. 2004;28(5):375-94.
- 109. Bocci V. Interleukins. Clinical pharmacokinetics and practical implications. Clin Pharmacokinet. 1991;21(4):274-84.
- 110. Krishnaswamy G, Kelley J, Yerra L, Smith JK, Chi DS. Human endothelium as a source of multifunctional cytokines: molecular regulation and possible role in human disease. J Interferon Cytokine Res. 1999;19(2):91-104.
- 111. Francis A, Bosio E, Stone SF, Fatovich DM, Arendts G, MacDonald SPJ, et al. Markers Involved in Innate Immunity and Neutrophil Activation are Elevated during Acute Human Anaphylaxis: Validation of a Microarray Study. J Innate Immun. 2019;11(1):63-73.
- 112. Mican JA, Arora N, Burd PR, Metcalfe DD. Passive cutaneous anaphylaxis in mouse skin is associated with local accumulation of interleukin-6 mRNA and immunoreactive interleukin-6 protein. J Allergy Clin Immunol. 1992;90(5):815-24.
- 113. Isabwe GAC, Garcia Neuer M, de Las Vecillas Sanchez L, Lynch DM, Marquis K, Castells M. Hypersensitivity reactions to therapeutic monoclonal antibodies: Phenotypes and endotypes. J Allergy Clin Immunol. 2018;142(1):159-70.e2.
- 114. Galand C, Leyva-Castillo JM, Yoon J, Han A, Lee MS, McKenzie ANJ, et al. IL-33 promotes food anaphylaxis in epicutaneously sensitized mice by targeting mast cells. J Allergy Clin Immunol. 2016;138(5):1356-66.
- 115. Yuste-Montalvo A, Fernandez-Bravo S, Oliva T, Pastor-Vargas C, Betancor D, Goikoetxea MJ, et al. Proteomic and Biological Analysis of an In Vitro Human Endothelial System in Response to Drug Anaphylaxis. Front Immunol. 2021;12:692569.
- 116. Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. Nat Immunol. 2010;11(9):785-97.
- 117. Ling M, Murali M. Analysis of the Complement System in the Clinical Immunology Laboratory. Clin Lab Med. 2019;39(4):579-90.
- 118. Tichaczek-Goska D. Deficiencies and excessive human complement system activation in disorders of multifarious etiology. Adv Clin Exp Med. 2012;21(1):105-14.

- 119. Guo Q, Subramanian H, Gupta K, Ali H. Regulation of C3a receptor signaling in human mast cells by G protein coupled receptor kinases. PLoS One. 2011;6(7):e22559.
- 120. el-Lati SG, Dahinden CA, Church MK. Complement peptides C3a- and C5a-induced mediator release from dissociated human skin mast cells. J Invest Dermatol. 1994;102(5):803-6.
- 121. Nigrovic PA, Malbec O, Lu B, Markiewski MM, Kepley C, Gerard N, et al. C5a receptor enables participation of mast cells in immune complex arthritis independently of Fcγ receptor modulation. Arthritis Rheum. 2010;62(11):3322-33.
- 122. Szebeni J. Complement activation-related pseudoallergy: a stress reaction in blood triggered by nanomedicines and biologicals. Mol Immunol. 2014;61(2):163-73.
- 123. Blossom DB, Kallen AJ, Patel PR, Elward A, Robinson L, Gao G, et al. Outbreak of adverse reactions associated with contaminated heparin. N Engl J Med. 2008;359(25):2674-84.
- 124. Adam A, Montpas N, Keire D, Désormeaux A, Brown NJ, Marceau F, et al. Bradykinin forming capacity of oversulfated chondroitin sulfate contaminated heparin in vitro. Biomaterials. 2010;31(22):5741-8.
- 125. Corbier A, Le Berre N, Rampe D, Meng H, Lorenz M, Vicat P, et al. Oversulfated chondroitin sulfate and OSCS-contaminated heparin cause dose- and route-dependent hemodynamic effects in the rat. Toxicol Sci. 2011;121(2):417-27.
- 126. Bender L, Weidmann H, Rose-John S, Renné T, Long AT. Factor XII-Driven Inflammatory Reactions with Implications for Anaphylaxis. Front Immunol. 2017;8:1115.
- 127. Smith PL, Kagey-Sobotka A, Bleecker ER, Traystman R, Kaplan AP, Gralnick H, et al. Physiologic manifestations of human anaphylaxis. J Clin Invest. 1980;66(5):1072-80.
- 128. van der Linden PW, Hack CE, Eerenberg AJ, Struyvenberg A, van der Zwan JK. Activation of the contact system in insectsting anaphylaxis: association with the development of angioedema and shock. Blood. 1993;82(6):1732-9.
- 129. Sala-Cunill A, Björkqvist J, Senter R, Guilarte M, Cardona V, Labrador M, et al. Plasma contact system activation drives anaphylaxis in severe mast cell-mediated allergic reactions. J Allergy Clin Immunol. 2015;135(4):1031-43.e6.
- Lieberman P, Simons FER. Anaphylaxis and cardiovascular disease: therapeutic dilemmas. Clin Exp Allergy. 2015;45(8):1288-95.
- 131. Brown SGA. Cardiovascular aspects of anaphylaxis: implications for treatment and diagnosis. Curr Opin Allergy Clin Immunol. 2005;5(4):359-64.
- 132. Soufras GD, Kounis GN, Kounis NG. Brain injury due to anaphylactic shock: broadening manifestations of Kounis syndrome. Int Endod J. 2014;47(4):309-13.
- 133. Jolobe OMP. Kounis syndrome and anaphylaxis. Am J Emerg Med. 2022;56:264.
- 134. Cha YS, Kim H, Bang MH, Kim OH, Kim HI, Cha K, et al. Evaluation of myocardial injury through serum troponin I and echocardiography in anaphylaxis. Am J Emerg Med. 2016;34(2):140-4.
- 135. Lippi G, Buonocore R, Schirosa F, Cervellin G. Cardiac troponin I is increased in patients admitted to the emergency department with severe allergic reactions. A case-control study. Int J Cardiol. 2015;194:68-9.
- 136. Kounis NG, Cervellin G, Koniari I, Bonfanti L, Dousdampanis P, Charokopos N, et al. Anaphylactic cardiovascular collapse

J Investig Allergol Clin Immunol 2022; Vol. 32(6): 419-437 doi: 10.18176/jiaci.0854 and Kounis syndrome: systemic vasodilation or coronary vasoconstriction? Ann Transl Med. 2018;6(17):332.

- 137. Chaulin AM. Biology of Cardiac Troponins: Emphasis on Metabolism. Biology (Basel). 2022;11(3):429.
- 138. Callesen KT, Yuste-Montalvo A, Poulsen LK, Jensen BM, Esteban V. In Vitro Investigation of Vascular Permeability in Endothelial Cells from Human Artery, Vein and Lung Microvessels at Steady-State and Anaphylactic Conditions. Biomedicines. 2021;9(4):439.
- 139. Wawrzyniak M, Pich C, Gross B, Schütz F, Fleury S, Quemener S, et al. Endothelial, but not smooth muscle, peroxisome proliferator-activated receptor β/δ regulates vascular permeability and anaphylaxis. J Allergy Clin Immunol. 2015;135(6):1625-35.e5.
- 140. Godo S, Shimokawa H. Endothelial Functions. Arterioscler Thromb Vasc Biol. 2017;37(9):e108-14.
- 141. Touyz RM, Alves-Lopes R, Rios FJ, Camargo LL, Anagnostopoulou A, Arner A, et al. Vascular smooth muscle contraction in hypertension. Cardiovasc Res. 2018;114(4):529-39.
- 142. Félétou M, Vanhoutte PM. EDHF: an update. Clin Sci (Lond). 2009;117(4):139-55.
- 143. Evora PRB, Simon MR. Role of nitric oxide production in anaphylaxis and its relevance for the treatment of anaphylactic hypotension with methylene blue. Ann Allergy Asthma Immunol. 2007;99(4):306-13.
- 144. Russo D, Minutolo R, Clienti C, De Nicola L, Iodice C, Savino FA, et al. Endothelin-1 released by vascular smooth muscle cells enhances vascular responsiveness of rat mesenteric arterial bed exposed to high perfusion flow. Am J Hypertens. 1999;12(11 Pt 1):1119-23.
- 145. Chu L yun, Liou JY, Wu KK. Prostacyclin protects vascular integrity via PPAR/14-3-3 pathway. Prostaglandins Other Lipid Mediat. 2015;118-119:19-27.
- 146. Cyr AR, Huckaby LV, Shiva SS, Zuckerbraun BS. Nitric Oxide and Endothelial Dysfunction. Crit Care Clin. 2020;36(2):307-21.
- 147. Cauwels A, Janssen B, Buys E, Sips P, Brouckaert P. Anaphylactic shock depends on PI3K and eNOS-derived NO. J Clin Invest. 2006;116(8):2244-51.
- 148. Nakamura Y, Hashiba Y, Endo J, Furuie M, Isozaki A, Yagi K. Elevated exhaled nitric oxide in anaphylaxis with respiratory symptoms. Allergol Int. 2015;64(4):359-63.
- 149. Lambden S, Creagh-Brown BC, Hunt J, Summers C, Forni LG. Definitions and pathophysiology of vasoplegic shock. Crit Care. 2018;22(1):174.
- 150. Metz M, Schäfer B, Tsai M, Maurer M, Galli SJ. Evidence that the endothelin A receptor can enhance IgE-dependent anaphylaxis in mice. J Allergy Clin Immunol. 2011;128(2):424-6.e1.
- 151. Kizawa Y, Kotake H, Kusama T, Saito K, Murakami H. Antigeninduced elevation of immunoreactive endothelin-1 (ET-1) levels in ovalbumin-sensitized guinea pig airway tissue. Comp Biochem Physiol C Pharmacol Toxicol Endocrinol. 1999;122(2):239-43.
- Melincovici CS, Boşca AB, Şuşman S, Mărginean M, Mihu C, Istrate M, et al. Vascular endothelial growth factor (VEGF) key factor in normal and pathological angiogenesis. Rom J Morphol Embryol. 2018;59(2):455-67.
- 153. Takahashi T, Ono S, Ogawa K, Tamura M, Mizutani T. [A case of anaphylactoid shock occurring immediately after

the initiation of second intravenous administration of highdose immunoglobulin (IVIg) in a patient with Crow-Fukase syndrome]. Rinsho Shinkeigaku. 2003;43(6).

- 154. Catala M, Kubis N. Gross anatomy and development of the peripheral nervous system. Handb Clin Neurol. 2013;115:29-41.
- 155. Farley A, Johnstone C, Hendry C, McLafferty E. Nervous system: part 1. Nurs Stand. 2014;28(31):46-51.
- 156. Forsythe P. The nervous system as a critical regulator of immune responses underlying allergy. Curr Pharm Des. 2012;18(16):2290-304.
- 157. Kenney MJ, Ganta CK. Autonomic nervous system and immune system interactions. Compr Physiol. 2014;4(3):1177-200.
- 158. Gupta K, Harvima IT. Mast cell-neural interactions contribute to pain and itch. Immunol Rev. 2018;282(1):168-87.
- 159. Barrios VE, Jarosinski MA, Wright CD. Proteinase-activated receptor-2 mediates hyperresponsiveness in isolated guinea pig bronchi. Biochem Pharmacol. 2003;66(3):519-25.
- Escribese MM, Rosace D, Chivato T, Fernández TD, Corbí AL, Barber D. Alternative Anaphylactic Routes: The Potential Role of Macrophages. Front Immunol. 2017;8:515.
- 161. Fauvel JM. [Interventional cardiology]. Rev Prat. 1988;38:19-23.
- 162. Eum SY, Norel X, Lefort J, Labat C, Vargaftig BB, Brink C. Anaphylactic bronchoconstriction in BP2 mice: interactions between serotonin and acetylcholine. Br J Pharmacol. 1999;126(1):312-6.
- 163. Beutier H, Hechler B, Godon O, Wang Y, Gillis CM, de Chaisemartin L, et al. Platelets expressing IgG receptor FcγRIIA/CD32A determine the severity of experimental anaphylaxis. Sci Immunol. 2018;3(22):eaan5997.
- 164. Tilley SL, Wagoner VA, Salvatore CA, Jacobson MA, Koller BH. Adenosine and inosine increase cutaneous vasopermeability by activating A(3) receptors on mast cells. J Clin Invest. 2000;105(3):361-7.
- 165. Livingston M, Heaney LG, Ennis M. Adenosine, inflammation and asthma--a review. Inflamm Res. 2004;53(5):171-8.
- 166. Piper PJ, Collier HO. Release of catecholamines in the guinea-pig by substances involved in anaphylaxis. Nature. 1967;213(5078):838-40.
- 167. van der Linden PW, Struyvenberg A, Kraaijenhagen RJ, Hack CE, van der Zwan JK. Anaphylactic shock after insect-sting challenge in 138 persons with a previous insect-sting reaction. Ann Intern Med. 1993;118(3):161-8.
- 168. van den Blink B, Wijsenbeek MS, Hoogsteden HC. Serum biomarkers in idiopathic pulmonary fibrosis. Pulm Pharmacol Ther. 2010;23(6):515-20.
- Anderson NL, Anderson NG. The human plasma proteome: history, character, and diagnostic prospects. Mol Cell Proteomics. 2002;1(11):845-67.
- 170. Pettersson ME, Koppelman GH, Flokstra-de Blok BMJ, van Ginkel CD, Roozendaal C, Muller-Kobold AC, et al. Apolipoprotein B: a possible new biomarker for anaphylaxis. Ann Allergy Asthma Immunol. 2017;118(4):515-6.
- 171. Wittenberg M, Nassiri M, Francuzik W, Lehmann K, Babina M, Worm M. Serum levels of 9α,11β-PGF2 and apolipoprotein A1 achieve high predictive power as biomarkers of anaphylaxis. Allergy. 2017;72(11):1801-5.
- 172. Lu TX, Rothenberg ME. MicroRNA. J Allergy Clin Immunol. 2018;141(4):1202-7.

- 173. Saliminejad K, Khorram Khorshid HR, Soleymani Fard S, Ghaffari SH. An overview of microRNAs: Biology, functions, therapeutics, and analysis methods. J Cell Physiol. 2019;234(5):5451-65.
- 174. Mori MA, Ludwig RG, Garcia-Martin R, Brandão BB, Kahn CR. Extracellular miRNAs: From Biomarkers to Mediators of Physiology and Disease. Cell Metab. 2019;30(4):656-73.
- 175. Backes C, Meese E, Keller A. Specific miRNA Disease Biomarkers in Blood, Serum and Plasma: Challenges and Prospects. Mol Diagn Ther. 2016;20(6):509-18.
- 176. Biethahn K, Orinska Z, Vigorito E, Goyeneche-Patino DA, Mirghomizadeh F, Föger N, et al. miRNA-155 controls mast cell activation by regulating the PI3Kγ pathway and anaphylaxis in a mouse model. Allergy. 2014;69(6):752-62.
- 177. Kwon Y, Kim Y, Eom S, Kim M, Park D, Kim H, et al. MicroRNA-26a/-26b-COX-2-MIP-2 Loop Regulates Allergic Inflammation and Allergic Inflammation-promoted Enhanced Tumorigenic and Metastatic Potential of Cancer Cells. J Biol Chem. 2015;290(22):14245-66.
- 178. Noh K, Kim M, Kim Y, Kim H, Kim H, Byun J, et al. miR-122-SOCS1-JAK2 axis regulates allergic inflammation and allergic inflammation-promoted cellular interactions. Oncotarget. 2017;8(38):63155-76.
- 179. Kim M, Park Y, Kwon Y, Kim Y, Byun J, Jeong MS, et al. MiR-135-5p-p62 Axis Regulates Autophagic Flux, Tumorigenic Potential, and Cellular Interactions Mediated by Extracellular Vesicles During Allergic Inflammation. Front Immunol. 2019;10:738.
- 180. Kwon Y, Kim M, Kim Y, Jeong MS, Jung HS, Jeoung D. EGR3-HDAC6-IL-27 Axis Mediates Allergic Inflammation and Is Necessary for Tumorigenic Potential of Cancer Cells Enhanced by Allergic Inflammation-Promoted Cellular Interactions. Front Immunol. 2021;12:680441.
- 181. Nuñez-Borque E, Fernandez-Bravo S, Rodriguez Del Rio P, Alwashali EM, Lopez-Dominguez D, Gutierrez-Blazquez MD, et al. Increased miR-21-3p and miR-487b-3p serum levels during anaphylactic reaction in food allergic children. Pediatr Allergy Immunol. 2021;32(6):1296-306.
- 182. Simeone P, Bologna G, Lanuti P, Pierdomenico L, Guagnano MT, Pieragostino D, et al. Extracellular Vesicles as Signaling Mediators and Disease Biomarkers across Biological Barriers. Int J Mol Sci. 2020;21(7):E2514.
- Raposo G, Stoorvogel W. Extracellular vesicles: exosomes, microvesicles, and friends. J Cell Biol. 2013;200(4):373-83.
- Carretero-González A, Otero I, Carril-Ajuria L, de Velasco G, Manso L. Exosomes: Definition, Role in Tumor Development and Clinical Implications. Cancer Microenviron. 2018;11(1):13-21.
- Bobrie A, Colombo M, Raposo G, Théry C. Exosome secretion: molecular mechanisms and roles in immune responses. Traffic. 2011;12(12):1659-68.
- 186. Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. Cell Mol Life Sci. 2018;75(2):193-208.
- 187. Xu K, Liu Q, Wu K, Liu L, Zhao M, Yang H, et al. Extracellular vesicles as potential biomarkers and therapeutic approaches in autoimmune diseases. J Transl Med. 2020;18(1):432.
- 188. Nuñez-Borque E, Fernandez-Bravo S, Pastor-Vargas C, Alvarez-Llamas G, Gutierrez-Blazquez MD, Alwashali E, et al. Proteomic profile of extracellular vesicles in anaphylaxis and their role in vascular permeability. Allergy. 2021;76(7):2276-9.

- Barber D, Villaseñor A, Escribese MM. Metabolomics strategies to discover new biomarkers associated to severe allergic phenotypes. Asia Pac Allergy. 2019;9(4):e37.
- Rodriguez-Coira J, Villaseñor A, Izquierdo E, Huang M, Barker-Tejeda TC, Radzikowska U, et al. The Importance of Metabolism for Immune Homeostasis in Allergic Diseases. Front Immunol. 2021;12:692004.
- 191. Hu X, Wu GP, Zhang MH, Pan SQ, Wang RR, Ouyang JH, et al. GC-MS-based metabolic profiling reveals metabolic changes in anaphylaxis animal models. Anal Bioanal Chem. 2012;404(3):887-93.
- 192. Perales-Chorda C, Obeso D, Twomey L, Rojas-Benedicto A, Puchades-Carrasco L, Roca M, et al. Characterization of anaphylaxis reveals different metabolic changes depending on severity and triggers. Clin Exp Allergy. 2021;51(10):1295-309.
- 193. Marco-Martín G, La Rotta Hernández A, Vázquez de la Torre M, Higaki Y, Zubeldia JM, Baeza ML. Differences in the Anaphylactic Response between C3H/HeOuJ and BALB/c Mice. Int Arch Allergy Immunol. 2017;173(4):204-12.
- 194. Morafo V, Srivastava K, Huang CK, Kleiner G, Lee SY, Sampson HA, et al. Genetic susceptibility to food allergy is linked to differential TH2-TH1 responses in C3H/HeJ and BALB/c mice. J Allergy Clin Immunol. 2003;111(5):1122-8.
- 195. Subramanian H, Gupta K, Ali H. Roles of Mas-related G proteincoupled receptor X2 on mast cell-mediated host defense, pseudoallergic drug reactions, and chronic inflammatory diseases. J Allergy Clin Immunol. 2016;138(3):700-10.
- 196. Kalesnikoff J, Galli SJ. Anaphylaxis: mechanisms of mast cell activation. Chem Immunol Allergy. 2010;95:45-66.
- 197. Finkelman FD. Anaphylaxis: lessons from mouse models. J Allergy Clin Immunol. 2007;120(3):506-15.
- 198. Gilfillan AM, Tkaczyk C. Integrated signalling pathways for mast-cell activation. Nat Rev Immunol. 2006;6(3):218-30.
- 199. Elieh Ali Komi D, Wöhrl S, Bielory L. Mast Cell Biology at Molecular Level: a Comprehensive Review. Clinic Rev Allerg Immunol. 2020;58(3):342-65.
- 200. Gomez G, Zhao W, Schwartz LB. Disparity in FccRI-induced degranulation of primary human lung and skin mast cells exposed to adenosine. J Clin Immunol. 2011;31(3):479-87.
- 201. Gao S, Zhu H, Zuo X, Luo H. Cathepsin G and Its Role in Inflammation and Autoimmune Diseases. Arch Rheumatol. 2018;33(4):498-504.
- 202. Castells M. Diagnosis and management of anaphylaxis in precision medicine. J All Clin Immunol. 2017;140(2):321-33.
- 203. Fregonese L, Swan FJ, van Schadewijk A, Dolhnikoff M, Santos MA, Daha MR, et al. Expression of the anaphylatoxin receptors C3aR and C5aR is increased in fatal asthma. J Allergy Clin Immunol. 2005;115(6):1148-54.
- 204. Fukuoka Y, Xia HZ, Sanchez-Muñoz LB, Dellinger AL, Escribano L, Schwartz LB. Generation of anaphylatoxins by human betatryptase from C3, C4, and C5. J Immunol. 2008;180(9):6307-16.
- 205. Akdis M, Burgler S, Crameri R, Eiwegger T, Fujita H, Gomez E, et al. Interleukins, from 1 to 37, and interferon-γ: receptors, functions, and roles in diseases. J Allergy Clin Immunol. 2011;127(3):701-21.e1-70.
- 206. Strait RT, Morris SC, Smiley K, Urban JF, Finkelman FD. IL-4 exacerbates anaphylaxis. J Immunol. 2003;170(7):3835-42.

- 207. Bruton K, Spill P, Vohra S, Baribeau O, Manzoor S, Gadkar S, et al. Interrupting reactivation of immunologic memory diverts the allergic response and prevents anaphylaxis. J Allergy Clin Immunol. 2021;147(4):1381-92.
- Thivierge M, Stanková J, Rola-Pleszczynski M. IL-13 and IL-4 upregulate cysteinyl leukotriene 1 receptor expression in human monocytes and macrophages. J Immunol. 2001;167(5):2855-60.
- 209. Komai-Koma M, Brombacher F, Pushparaj PN, Arendse B, McSharry C, Alexander J, et al. Interleukin-33 amplifies IgE synthesis and triggers mast cell degranulation via interleukin-4 in naïve mice. Allergy. 2012;67(9):1118-26.
- 210. Kritas SK, Saggini A, Varvara G, Murmura G, Caraffa A, Antinolfi P, et al. Impact of mast cells on the skin. Int J Immunopathol Pharmacol. 2013;26(4):855-9.
- 211. Berge V, Johnson E, Berge KE. Interleukin-1 alpha, interleukin 6 and tumor necrosis factor alpha increase the synthesis and expression of the functional alternative and terminal complement pathways by human umbilical vein endothelial cells in vitro. APMIS. 1996;104(3):213-9.
- 212. Dauchel H, Julen N, Lemercier C, Daveau M, Ozanne D, Fontaine M, et al. Expression of complement alternative pathway proteins by endothelial cells. Differential regulation by interleukin 1 and glucocorticoids. Eur J Immunol. 1990;20(8):1669-75.
- 213. Hauswirth AW, Sonneck K, Florian S, Krauth MT, Bohm A, Sperr WR, et al. Interleukin-3 promotes the expression of E-NPP3/ CD203C on human blood basophils in healthy subjects and in patients with birch pollen allergy. Int J Immunopathol Pharmacol. 2007;20(2):267-78.
- 214. Suzukawa M, Komiya A, Yoshimura-Uchiyama C, Kawakami A, Koketsu R, Nagase H, et al. IgE- and FcepsilonRI-mediated enhancement of surface CD69 expression in basophils: role of low-level stimulation. Int Arch Allergy Immunol. 2007;143 Suppl 1:56-9.
- 215. Kruse S, Kuehr J, Moseler M, Kopp MV, Kurz T, Deichmann KA, et al. Polymorphisms in the IL 18 gene are associated with specific sensitization to common allergens and allergic rhinitis. J Allergy Clin Immunol. 2003;111(1):117-22.
- 216. Novak N, Kruse S, Potreck J, Maintz L, Jenneck C, Weidinger S, et al. Single nucleotide polymorphisms of the IL18 gene are associated with atopic eczema. J Allergy Clin Immunol. 2005;115(4):828-33.
- 217. Yoshimoto T, Tsutsui H, Tominaga K, Hoshino K, Okamura H, Akira S, et al. IL-18, although antiallergic when administered with IL-12, stimulates IL-4 and histamine release by basophils. Proc Natl Acad Sci U S A. 1999;96(24):13962-6.
- 218. Chadban SJ, Tesch GH, Foti R, Lan HY, Atkins RC, Nikolic-Paterson DJ. Interleukin-10 differentially modulates MHC class II expression by mesangial cells and macrophages in vitro and in vivo. Immunology. 1998;94(1):72-8.
- 219. Taylor A, Verhagen J, Blaser K, Akdis M, Akdis CA. Mechanisms of immune suppression by interleukin-10 and transforming growth factor-beta: the role of T regulatory cells. Immunology. 2006;117(4):433-42.
- 220. Akdis CA, Akdis M. Mechanisms and treatment of allergic disease in the big picture of regulatory T cells. J Allergy Clin Immunol. 2009;123(4):735-46.
- 221. Akdis M, Akdis CA. Therapeutic manipulation of immune tolerance in allergic disease. Nat Rev Drug Discov. 2009;8(8):645-60.

- 222. Frossard CP, Steidler L, Eigenmann PA. Oral administration of an IL-10-secreting Lactococcus lactis strain prevents food-induced IgE sensitization. J Allergy Clin Immunol. 2007;119(4):952-9.
- 223. Lee SY, Huang CK, Zhang TF, Schofield BH, Burks AW, Bannon GA, et al. Oral administration of IL-12 suppresses anaphylactic reactions in a murine model of peanut hypersensitivity. Clin Immunol. 2001;101(2):220-8.
- 224. Baier A, Ndoh VNE, Lacy P, Eitzen G. Rac1 and Rac2 control distinct events during antigen-stimulated mast cell exocytosis. J Leukoc Biol. 2014;95(5):763-74.
- 225. Sheshachalam A, Baier A, Eitzen G. The effect of Rho drugs on mast cell activation and degranulation. J Leukoc Biol. 2017;102(1):71-81.
- 226. Groot M, Lee H. Sorting Mechanisms for MicroRNAs into Extracellular Vesicles and Their Associated Diseases. Cells. 2020;9(4):E1044.

227. Kim M, Jo H, Kwon Y, Jeong MS, Jung HS, Kim Y, et al. MiR-154-5p-MCP1 Axis Regulates Allergic Inflammation by Mediating Cellular Interactions. Front Immunol. 2021;12:663726.

Manuscript received June 4, 2022; accepted for publication August 18, 2022.

Vanesa Esteban

Department of Allergy and Immunology IIS-Fundación Jiménez Díaz Avda Reyes Católicos 2 28040 Madrid, Spain E-mail: vesteban@fjd.es